

1 **Polychlorinated biphenyls in agricultural soils from the Yangtze River Delta of**
2 **China: Regional contamination characteristics, combined ecological effects and**
3 **human health risks**

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28 **ABSTRACT**

29 The current contamination status of polychlorinated biphenyls (PCBs) was studied in the
30 agricultural soils of the Yangtze River Delta (YRD), one of the largest economic zones in China.
31 The concentrations of PCBs ranged from <0.1 to 130 ng/g dry weight. Higher concentrations of
32 PCBs were observed in the 0 e30 cm surface layers relative to the subsurface soils. A distinct
33 spatial distribution was observed with a declining concentration gradient from the northwest to the
34 southeast of the region. The composition of PCBs in the soils was consistent with the Chinese
35 commercial PCB mixtures, but different from the compositions in global background soil. Local
36 sources including large-scale use and disposal of PCB- containing products were the main potential
37 sources to soil. The ecological effects and human health risks associated with combined persistent
38 organic pollutants, including PCBs, organochlorine pesticides (OCPs), phthalate esters (PAEs) and
39 polybrominated diphenyl ethers (PBDEs), were further estimated. The four toxic organic
40 compounds and seven physicochemical parameters together could only explain 12.7% of the
41 variation in microbial community composition, suggesting the soil ecosystem function was not
42 strongly influenced by the combined pollution at low concentrations. However, the potential health
43 risks to residents via multiple pathways were notably higher for PCBs than other chemicals. The
44 potential risks were mainly derived from PCB-126, 81, and 169.

45

46 **INTRODUCTION**

47 Polychlorinated biphenyls (PCBs) are a well-known class of persistent organic pollutants (POPs),
48 and have been used as dielectric and heat-exchange fluids, flame-retardants, plasticizers, and
49 pesticide additives (Breivik et al., 2007). Despite being banned for several decades, the widespread

50 production and use of PCBs before their legal restriction has led to ubiquitous contamination of
51 the environment, presenting potential ecological and human health risks especially for metabolites
52 upon PCBs transformation (Gutleb et al., 2010; Su et al., 2012; Sun et al., 2016a).

53 Soil is an important reservoir for many POPs including PCBs (Doick et al., 2005; Zhang et al.,
54 2011; Zhong and Zhu, 2013), and is a secondary emission source to air (Bidleman and Leone,
55 2004; Cabrerizo et al., 2011) and water (Zhang et al., 2003). Even relatively low concentrations
56 of PCBs in soils can be bioaccumulated gradually via food chains with potentially adverse effects
57 on humans (Sirot et al., 2012). Globally, nearly 21,000 tons of PCBs have been discharged into
58 soils (Meijer et al., 2003). In China, about 10,000 tons of commercial PCB were produced in the
59 past (Zhang et al., 2013), which have polluted soils and other environmental components across
60 the country (Bao et al., 2012; Ren et al., 2007; Zheng et al., 2016).

61 The Yangtze River Delta (YRD) is one of the three largest regional economic zones in China. A
62 large number of industrial and commercial enterprises contribute to regional economic
63 development, but also pose a serious threat to the environment quality. Our recent study found that
64 the agricultural soil in the YRD is contaminated with organochlorine pesticides (OCPs), phthalate
65 esters (PAEs) and polybrominated diphenyl ethers (PBDEs) (Sun et al., 2016b). It is important to
66 investigate the regional-scale distribution and the associated risks of PCBs to manage
67 environmental risks and facilitate sustainable industrial and economic development.
68 Environmental pollution has been found to cause drastic changes in microbial community activity
69 and affect the functions of soil ecosystems (Brantner and Senko, 2014; Turpeinen et al., 2004).
70 Yet there are limited studies on the responses of soil microbiota to the combined pollution and its
71 ecological consequences (Liu et al., 2015). The potential risks of combined pollution (including
72 PCBs, OCPs, PAEs, and PBDEs) to human health via multiple pathways have not been well

73 understood. Although an individual pollutant cannot impose significant health risk to human, a
74 long-term chronic exposure to a mixture of contaminants could potentially cause serious
75 carcinogenic and non-cancer effects (Sumpter and Johnson, 2005).

76 The aims of this work are (i) to investigate the concentration, composition and spatial distributions
77 of PCBs in soils of the YRD region; and (ii) to assess the ecological effect and human health risk
78 of combined pollution in soils and to provide a basis for assessing soil quality in the YRD during
79 the course of rapid economic development.

80

81 **EXPERIMENTAL METHODS**

82 **Sample collection**

83 The YRD economic region comprises the Shanghai Municipality, southern Jiangsu and northern
84 Zhejiang Provinces in eastern China, covers 100,000 km² and has a population of about 100 million
85 (China Today, 2013). An evenly distributed sampling network composed of 241 sites was schemed
86 to cover an area of approximately 45,800 km² (Fig. 1). Detailed information was described in a
87 previous publication (Sun et al., 2016b). During June 2014, a total of 241 topsoil samples (0e15
88 cm depth) and six soil profiles were collected from various farmlands in the YRD region. The land-
89 use types included paddy fields, vegetable fields, forests, uncultivated lands, and other agricultural
90 uses. The soil profiles were excavated to a depth of 80 cm. Soils were taken from the front of the
91 soil trenches at 10 cm intervals, each interval provides approximately 500 g soil samples. The soil
92 samples were collected using a stainless steel scoop, packed in aluminum foil, sealed in Kraft bags,
93 and freeze-dried in the laboratory.

94

95 **Extraction and analysis of PCBs**

96 The extraction and cleanup procedures for PCBs were adapted from the reported methods (Wang
97 et al., 2013). All freeze-dried samples were ground and sieved through a stainless steel 75- mesh
98 sieve and stored at 20 C before analysis. A portion of soil samples (5 g) were spiked with recovery
99 surrogate PCB-65, followed by ultrasonic extraction using hexane/dichloromethane (1:1 v/v, 40
100 mL) for 30 min. The procedure was repeated two more times. The extracts were combined and
101 concentrated to 1 mL, and then subjected to cleanup by a multilayer silica column as follows (from
102 bottom to top): 1 g silica, 4 g basic silica (1.2%, w/w), 1 g silica, 8 g acidic silica (30%, w/w), 2 g
103 silica, and 4 g anhydrous sodium sulfate. The column was pre-washed with 80 mL of hexane. The
104 extract was eluted with 100 mL of hexane, and was thereafter concentrated by rotary evaporation
105 to a final volume of 200 μ L prior to instrumental analysis.

106 The target compounds measured in all samples include 12 dioxin-like PCBs (DL-PCBs) (including
107 PCB-77, 81, 105, 114, 118, 123, 126, 156, 157, 167, 169, and 189), and six indicator PCBs
108 (including PCB-28, 52, 101, 138, 153, and 180). Determination of PCBs was performed on gas
109 chromatography/mass spectrometry (GC/MS) (7890B/5977A, Agilent Technologies, Santa Clara,
110 CA, USA) with an electron ionization source. A DB-5 MS capillary column was used (30 m 0.25
111 mm i.d. with 0.25 μ m film thickness). All data were obtained in the selective ion monitoring (SIM)
112 mode. High purity helium was used as carrier gas with a flow rate of 1.0 mL/min. Oven temperature
113 program was as follows: initial column temperature at 80°C for 3 min, ramp to 150°C at 15°C/min
114 and hold for 2 min, increase to 270°C at 2.5 °C/min and hold for 3 min, increase to 300°C at 15 °C
115 /min and hold for 5 min.

116

117 **Quality assurance and quality control**

118 A procedural blank, a spiked blank, and a sample duplicate were processed in parallel with each
119 batch of ten samples. No targeted compound was found in the blanks. The recovery rates of PCBs
120 in the spiked samples ranged from 86.1% to 94.4%. The variations in concentrations of PCBs in
121 duplicates were lower than 20% (n = 3). Five-point standard calibration curves were employed for
122 quantitative analysis. The recovery rates of surrogate standards were 81.7% - 103%. The
123 concentrations were not corrected with recovery rate. The limit of detection (LOD) of PCBs was
124 defined on a signal- to-noise ratio of three using the lowest concentration standard and ranged
125 between 0.03 and 0.10 ng/g.

126

127 **Microbiological analysis**

128 Microbiological analysis was conducted for all the 241 topsoil samples. Extraction and analysis of
129 microbial phospholipid fatty acids (PLFA) were performed with reference to the reported method
130 (He et al., 2013). Briefly, PLFAs were extracted from soil sample using a mixture of chloroform-
131 methanol-citrate buffer and then separated by solid-phase extraction cartridges. PLFAs were
132 analyzed by GC fitted with MIDI Sherlock microbial identification system. The microbial biomass
133 of bacteria, fungi, actinomycetes, gram-positive bacteria, and gram-negative bacteria were
134 separately quantified on the basis of the detected 42 fatty acids.

135

136 **Physicochemical analysis**

137 The water content in the soil samples was determined by weighing samples before and after oven-
138 drying at 105°C for 24 h. The sieved and freeze-dried samples were used for subsequent analyses.
139 The soil pH was measured using a pH meter (Mettlertoledo Instruments, Shanghai, China) with a
140 soil/water ratio of 1: 2.5. The total organic carbon (TOC) (using Vavio EL III elemental analyzer,
141 Elementar, Hanau, Germany), total nitrogen (using Rapid N cube, Elementar, Hanau, Germany),
142 and total phosphorus (using UV-1800, Shimadzu Instruments, Suzhou, China) were analyzed
143 according to standard methods (Page et al., 1982). The concentra- tions of zinc and copper were
144 measured by inductively coupled plasma mass spectrometry (ICP-MS; NexION 300x ,
145 PerkinElmer, MA, USA) after acid digestion.

146

147 **Health risk assessment**

148 The non-cancer and carcinogenic risks of selected pollutants to human were evaluated with the
149 methods recommended by the United States Environmental Protection Agency (USEPA, 1997).
150 The average daily intake dose (ADD, mg kg⁻¹ day⁻¹) via non-dietary (including soil ingestion,
151 inhalation and dermal contact) and di- etary (i.e., intake of agricultural products harvested from
152 the soils) exposure routes (Niu et al., 2013) were estimated using the equations presented in
153 Supporting Information (SI). The non-cancer risks of pollutants via non-dietary and dietary
154 pathways were represented with the hazard index (HI). It is suggested that when $HI < 1$ the non-
155 cancer risk is negligible. In the carcinogenic risk evaluation, the carcinogenic risk of a pollutant
156 was classified as “very low” when the risk value was lower than 10^{-6} , “relatively low” in the range
157 of 10^{-6} and 10^{-4} , “moderate” in the range of 10^{-4} and 10^{-3} , “high” between 10^{-3} and 10^{-1} , and “very
158 high” when exceeding 10^{-1} (USEPA, 2009; Niu et al., 2013). This is a standard approach as a part

159 of tiered risk assessment, which could help to identify contaminant hotspots and provide a good
160 reference for site-specific, detailed assessment if needed in the future.

161

162 **Statistical analysis**

163 Statistical analyses were performed using SPSS 18.0, Origin 8.0, and R (<http://www.r-project.org>).

164 Statistical significance was considered as $p < 0.05$. Spatial distributions of PCBs were predicted

165 using universal Kriging in ArcGIS 10.2. Canonical correspondence analysis and partial canonical

166 correspondence analysis were used to determine the contribution of selected variables to the

167 variations of microbial community composition.

168

169 **RESULTS AND DISCUSSION**

170 **Concentrations and composition of PCBs**

171 A summary of the concentrations (ng/g dw) of PCB homologue groups and the sum of quantified

172 PCBs in 241 topsoil samples is presented in Table SI-1. The concentrations of total PCBs in the

173 YRD agricultural soils ranged from <0.1 to 130 ng/g, with a mean of

174 20.2 ng/g and a detection rate of 76.8%, indicating that PCBs were widely dispersed in the YRD

175 soils. The total PCBs levels in the present study were higher than those in soil of Qinghai-Tibet

176 Plateau (0.22×10^1 to 1.96×10^2 ng/g) (Gai et al., 2014), Hong Kong (0.07×10^1 to 9.87×10^1 ng/g) (Zhang et al., 2007), and

177 Germany (0.95×10^3 to 3.84×10^4 ng/g) (Manz et al., 2001), while they were lower than those in the soils of

178 the Pearl River Delta in South China (0.3×10^2 to 202×10^2 ng/g) (Zhang et al., 2013), the Iowa State in USA

179 (3×10^2 to 200×10^2 ng/g) (Martinez et al., 2012), and the Greater London in the UK (9×10^2 to 2600×10^2 ng/g) (Vane et

180 al., 2014). The levels of PCBs in these soils were similar to those found in the soils of Mongolia
181 (0.53e114 ng/g) (Mamontova et al., 2013) and European cities (0.15e86 ng/g) (Cachada et al.,
182 2009). According to the commonly used screening levels for resident soil (USEPA, 2016), PCB-
183 77, PCB-81, PCB-126, and PCB-169 in 2, 11, 34, and 10 sampling sites, respectively, exceeded
184 the allowable concentrations.

185 The detection rates of all the 18 PCB congeners ranged from 4.1% (PCB-153) to 45.2% (PCB-81).
186 The concentration of the sum of six indicator PCBs (PCB-28, 52, 101, 138, 153 and 180) among
187 the 241 sites varied from <0.1 to 119 ng/g, with a mean value of 17.1 ng/g, accounting for 80.2%
188 of the total PCBs concentrations. The concentrations of these six indicator PCBs were significantly
189 correlated with the concentrations of total PCBs ($R = 0.916$, $p < 0.01$, $n = 241$), thus they could be
190 used to represent the extent of PCBs contamination in the soil environment.

191 The major PCB homologue group residing in the YRD topsoil was tetra-PCB, followed by tri-
192 PCB and penta-PCB, which was consistent with the fact that 70% of the PCBs technical mixtures
193 produced globally were tri-, tetra- and penta-PCBs (Breivik et al., 2002). The detected composition
194 of PCBs was dominated by the low- chlorinated PCBs, while the high-chlorinated congeners, such
195 as hepta-PCBs, were detected at significantly lower concentrations in the YRD soil samples.
196 Although the global background soil was dominated by hexa-PCBs and penta-PCBs (Ren et al.,
197 2007), this study showed that the PCBs homologue compositions of the YRD agricultural soils
198 were different and hexa-PCBs were mainly found in the top 10e30 cm surface soil only (Fig. SI-
199 1).

200 The concentration of the sum of 12 dioxin-like PCBs among the 241 sites ranged from <0.1 to
201 78.9 ng/g, with a mean value of 7.5 ng/g. The World Health Organization toxic equivalency factors
202 (Van den Berg et al., 2006) were used to calculate the toxicity equivalents (TEQs) of the DL-PCBs.

203 The TEQ values ranged from below LOD to 379 ng-TEQ/kg, with the mean values of 9.5 ng-
204 TEQ/kg. The non ortho-PCB congeners, PCB-126 and PCB-169, presented the highest TEQ values
205 (i.e., highest toxic potency) among PCB homologous, accounting for >90% of the total TEQ. The
206 TEQ concentration was notably higher than the data reported in other Chinese cities, such as
207 Beijing (0.35 ng/kg) (Wu et al., 2011) and Dalian (1.37 ng/kg) (Wang et al., 2008).

208 The vertical concentrations and distributions of PCBs at different depths in six selected agricultural
209 soil profiles collected from the YRD are shown in Fig. SI-1. The higher concentrations of PCBs
210 were observed in the 0 - 30 cm surface layers relatively to the subsurface soils, because the upper
211 30 cm of the soil was a plow layer formed through frequent cultivation and plowing activities with
212 regular irrigation. A rapid decline of PCB concentrations to marginal level or even below the
213 detection limit was observed when the soil depth was greater than 30 cm, which was consistent
214 with previous findings (Cousins et al., 1999; Wang et al., 2006). This indicated that PCBs in
215 agricultural soils were less likely to contaminate the groundwater and subsurface environment.

216

217 **Spatial distributions and potential sources**

218 Distinct spatial distribution patterns of PCBs were observed (Fig. 1), where the concentrations of
219 PCBs were much higher in Jiangsu Province than those in Zhejiang Province and Shanghai
220 Municipality. A declining concentration gradient from the north- west direction was identified.
221 The highest concentrations of PCBs were found in Danyang, Jurong, and Changzhou cities, at the
222 longitude range between 119° and 120°, latitude range between 31° and 32°.

223 The composition of PCBs in the YRD soils was consistent with the Chinese commercial PCB
224 mixtures, which contained more low- molecular-weight PCBs (tri-CBs 40.4%, tetra-CBs 31.1%)

225 than global PCB products (tri-CBs 25.2%, tetra-CBs 24.7%) (Ren et al., 2007). The soil PCB
226 concentrations were strongly influenced by proximity to the sources. The large-scale use and
227 disposal of PCB-containing products, such as lubricants, dielectric fluids, transformers, capac-
228 itors, and plasticizers by the industries in these areas were potential sources for the high PCB levels
229 in soils (Breivik et al., 2002; Ren et al., 2007). Moreover, PCBs in building materials such as joint
230 sealing materials, plaster and paint might enter surrounding soils (Herrick et al., 2007). On the
231 other hand, long-range atmospheric transport and deposition of PCBs could not be excluded,
232 especially for low-molecular-weight PCBs (Meijer et al., 2003). Some micro-organisms might
233 also degrade highly chlorinated PCBs via dechlorination reaction into lower chlorinated PCBs as
234 residues in soils (Abraham et al., 2002).

235 The results also showed that the concentration of PCBs was positively correlated with pH ($R =$
236 0.149 , $p < 0.05$, $n = 241$), while there was no significant correlation with TOC ($R = - 0.099$, $p =$
237 0.126 , $n = 241$). The mean concentrations of total PCBs categorized with respect to land uses
238 ranked as follows: paddy fields (19.2 ng/g, $n = 44$) > vegetable fields (15.7 ng/g, $n = 48$) >
239 uncultivated lands (13.5 ng/g, $n = 26$) > forests (11.6 ng/g, $n = 52$). The anaerobic condition in
240 paddy fields was likely to hinder possible degradation of PCBs, though no statistically significant
241 difference was observed between different land-use types ($p > 0.05$). The relationship was also
242 analyzed between population density of sampling sites and the concentrations of PCBs. Positive
243 correlation was observed between PCBs and population density ($R = 0.140$, $p < 0.05$, $n = 241$),
244 suggesting that dense population, associated with high urbanization and industrialization, was
245 partly responsible for the elevated levels of PCBs in soils.

247 **Combined effects on microbial communities**

248 In the microbial communities of the studied soils, bacteria were predominant and made up $63.9 \pm$
249 13.7% of the total biomass. There was no significant difference between overall microbial diversity
250 in the collected agricultural soils ($p > 0.05$). As shown in Fig. 2, both fungi and actinomycete were
251 positively correlated with the concentrations of PBDEs and total phosphorus, and negatively
252 correlated with the concentrations of PAEs, PCBs and copper, whereas bacteria were positively
253 correlated with the pH, and the concentrations of PCBs, PAEs, and copper. Compared to gram-
254 positive bacteria, gram-negative bacteria were more correlated with pH and water.

255 Canonical correspondence analysis was applied to identify the combination of environmental
256 variables that could best fit the community composition patterns. The subset of 11 variables was
257 identified (i.e., seven soil physicochemical parameters including water content, pH, TOC, total
258 nitrogen, total phosphorus, copper, and zinc, and four kinds of toxic organic compounds including
259 PCBs, PBDEs, OCPs, and PAEs) to explain the variations in microbial community composition
260 in the soils. The results showed that the 11 selected variables together could explain 12.7% of the
261 observed variation in the community composition (Fig. 2). The subsequent partial canonical
262 correspondence analysis revealed that the seven physicochemical parameters, the four kinds of
263 toxic organic com- pounds, and the interaction effect of all these variables explained 11.4% , 0.86%
264 and 0.44% of the variation, respectively (Fig. 2). Thus, soil physicochemical parameters played a
265 more important role in regulating microbial communities than the combination of selected organic
266 pollutants in the YRD soils. Water content and pH were the primary factors, displaying 5.03% and
267 2.06% explanation among the predictor variables.

268 Liu et al. (2015) found significant differences in microbial composition between the contaminated
269 soils and reference soils due to the ecotoxicological effects of combined pollution associated with

270 crude e-waste processing. In comparison, the soils of this study were collected from farmland that
271 had not been severely polluted by point pollution sources. As the 241 sampling sites were evenly
272 distributed in the YRD region, these results suggested that the microbial communities in
273 agricultural soils were not strongly influenced by the combined pollution at low concentrations.
274 Other factors might have higher impact on soil microbial communities. It has been reported that
275 soil type, vegetation, climate, and land management practice also contribute to the diversity of
276 microbial communities (Fisk et al., 2003; Liu et al., 2000; Lupwayi et al., 2001). Such information
277 should be obtained and considered in future studies to capture more of the variation.

278

279 **Human health risk assessments**

280 The non-cancer risks and carcinogenic risk of chemicals to residents via multiple pathways were
281 assessed. The non-cancer risks to children were higher than those to adults (Fig. 3). The average
282 non-cancer risk of PCBs (HI = 44.0 for children, and 24.7 for adults) was overwhelmingly high,
283 followed by much lower risk due to OCPs (HI = 0.048 for children, and 0.026 for adults), PAEs
284 (HI = 0.036 for children, and 0.020 for adults) and PBDEs (HI = 0.017 for children, and 0.009 for
285 adults). Among the measured PCB congeners, the average HI of individual congeners in arable
286 soils was the highest for PCB-126 (HI = 37.1 for children, and 20.8 for adults), followed by PCB-
287 81 (HI = 5.74 for children, and 3.23 for adults), and PCB-169 (HI = 0.506 for children, and 0.280
288 for adults). The HI of other chemicals descended in the order of bis (2-ethylhexyl) phthalate
289 (DEHP) (HI = 0.028 for children, and 0.016 for adults) > γ -hexachlorocyclohexane (γ -HCH) (HI
290 0.016 for children, and 0.009 for adults) > BDE-47 (HI = 0.015 for children, and 0.008 for adults).
291 The estimated intake doses of PCBs exceeded the acceptable levels (HI > 1) for children and adults
292 in 55% and 52% of the soil samples, respectively. The estimated intake doses of PBDEs exceeded

293 the acceptable levels for children and adults in 0.83% and 0.42% of the soil samples, respectively.
294 None of the samples pose non-cancer risk from PAEs and OCPs.

295 On the contrary, the carcinogenic risks to adults were higher than those to children (Fig. SI-2). The
296 average carcinogenic risk of PCBs (3.42×10^{-4} for children and 8.32×10^{-4} for adults) to residents
297 was also the highest, followed by OCPs (2.30×10^{-6} for children and 5.53×10^{-6} for adults), PAEs
298 (6.78×10^{-7} for children and 1.63×10^{-6} for adults), and PBDEs (2.49×10^{-10} for children and
299 5.96×10^{-10} for adults). The high risks were mainly derived from PCB-126, PCB-81, and PCB-
300 169, respectively. Approximately 36% and 33% of the samples showed very low carcinogenic
301 risks ($<10^{-6}$) of PCBs to children and adults, respectively. The carcinogenic risks of PCBs in a
302 large number of samples (40% for children and 29% for adults) were between 10^{-6} and 10^{-4} ,
303 implying relatively low carcinogenic risks to residents. Moderate cancer risks (10^{-4} - 10^{-3}) of PCBs
304 to children and adults were found in 15% and 22% of the samples, respectively. Notably, 9% and
305 16% of the samples posed high cancer risks (10^{-3} - 10^{-1}) to children and adults, respectively. In
306 contrast, the carcinogenic risks posed by other chemical in the soils via non-dietary and dietary
307 routes were all on the low side ($<10^{-6}$).

308 The combined risks were estimated by summing the risks of all individual compounds in an
309 additive manner, which was implicitly assumed in the standard risk assessment of USEPA (Niu et
310 al., 2013). This may be a limitation of the cumulative risk interpretation. As shown by the findings
311 of this study, the combined risks of pollutants were dominated by PCBs. Human health risks
312 through intake of agricultural products contributed for over 99% of the total risks, suggesting that
313 the food consumption would be the primary contributor. A large number of soil samples in this
314 study were collected from farmlands inside villages or around farmers' houses. People living in

315 rural areas may frequently contact with these soils. The health risks of coexisting chemicals to
316 residents via multiple pathways should arouse more concern.

317

318 **CONCLUSION**

319 In this study, 241 topsoil samples and six soil profiles from agricultural fields in the YRD region
320 (approximately 45,800 km²) were collected and analyzed to reveal the status of PCB contamination
321 in one of regional economic zones of China. PCBs were found widespread contaminants in the
322 YRD region. The higher PCBs concentrations were observed in the 0 - 30 cm surface soils. PCBs
323 compositions in the YRD agricultural soils were consistent with the Chinese commercial PCB
324 mixtures, suggesting that local sources were potentially the main input of PCBs to soil. The mean
325 concentrations of PCBs measured in different land uses were in the following order: paddy field
326 > vegetable fields > uncultivated lands > forests. The subset of selected soil physicochemical
327 parameters better explained the variation in microbial community composition than the
328 combination of selected organic pollutants in soil. The risks of chemicals to residents via multiple
329 pathways ranked as PCBs > OCPs > PAEs > PBDEs, of which PCB-126, 81, and 169 showed
330 much higher risks than other compounds. Notably, PCBs in 9% and 16% of the samples posed
331 potential cancer risks (10^{-3} - 10^{-1}) to children and adults, respectively. The non-cancer risks of
332 PCBs exceeded acceptable levels (HI > 1) for children and adults in 55% and 52% of the soil
333 samples, respectively. Ubiquitous occurrence of coexisting organic compounds and their health
334 risks to residents should warrant more studies in the future.

335

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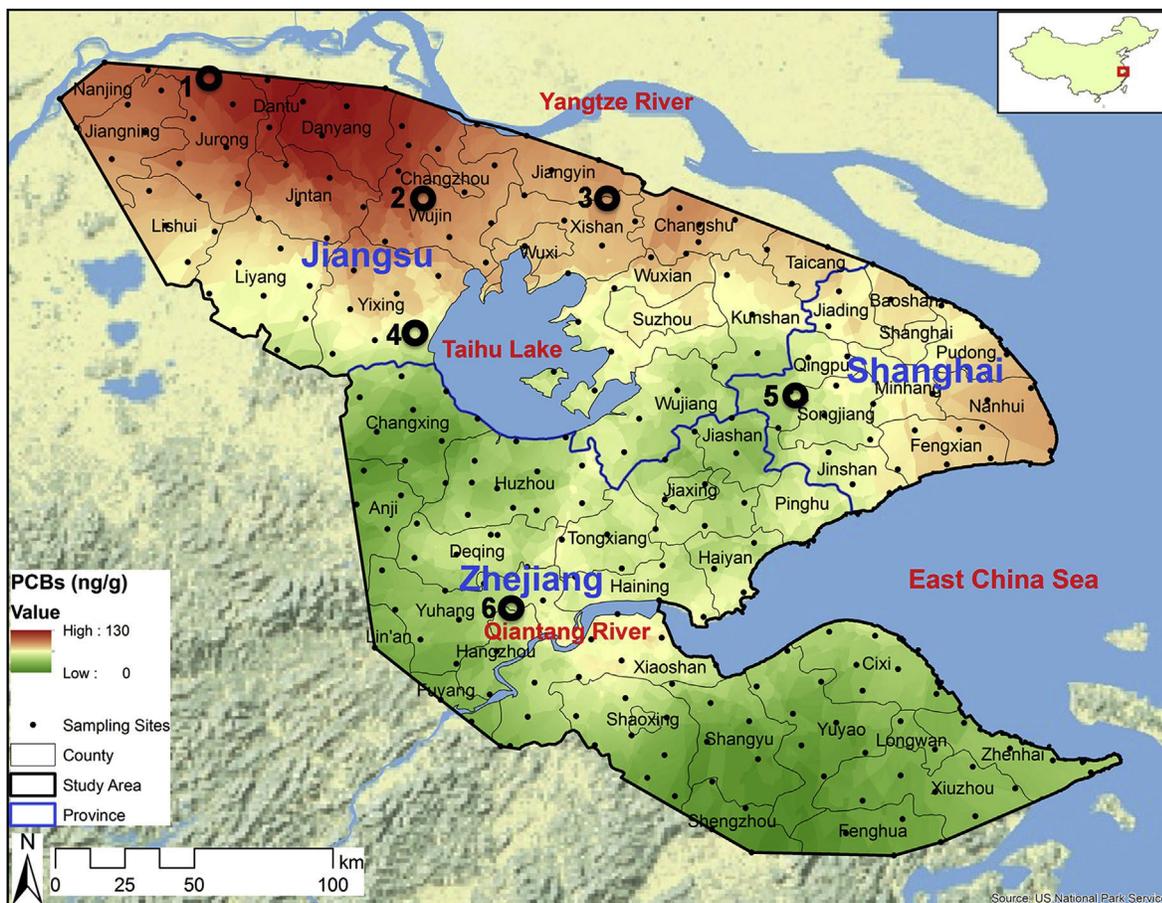
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343 **Appendix A. Supplementary data**

344 Supplementary data related to this article can be found at
345 <http://dx.doi.org/10.1016/j.chemosphere.2016.08.038>

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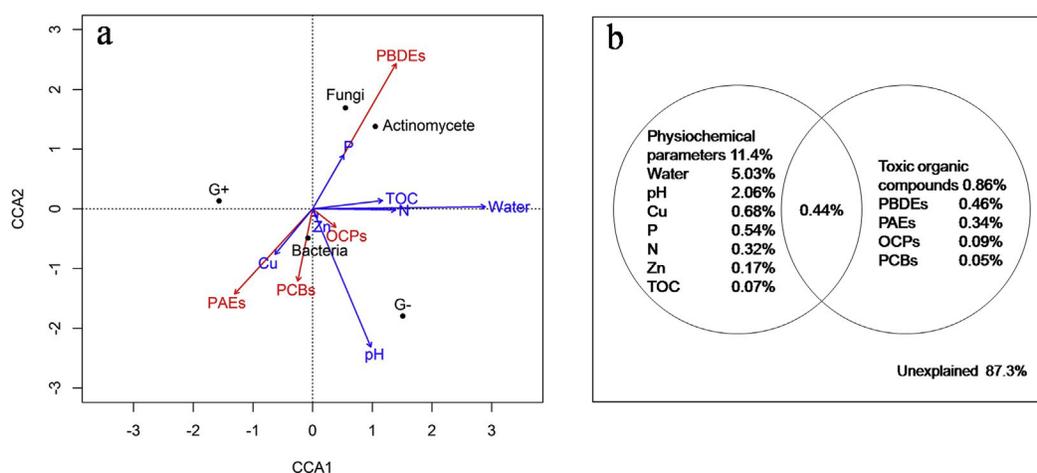
347 **LIST OF FIGURES**



348

349 Fig. 1. Sampling sites and spatial distribution of the total concentrations of PCBs in the agricultural
 350 soils of the YRD region. The six sampling sites with soil profiles are marked with bold numbers.
 351 The legend labels, “High: 130” and “Low: 0”, represent the highest and lowest concentrations.

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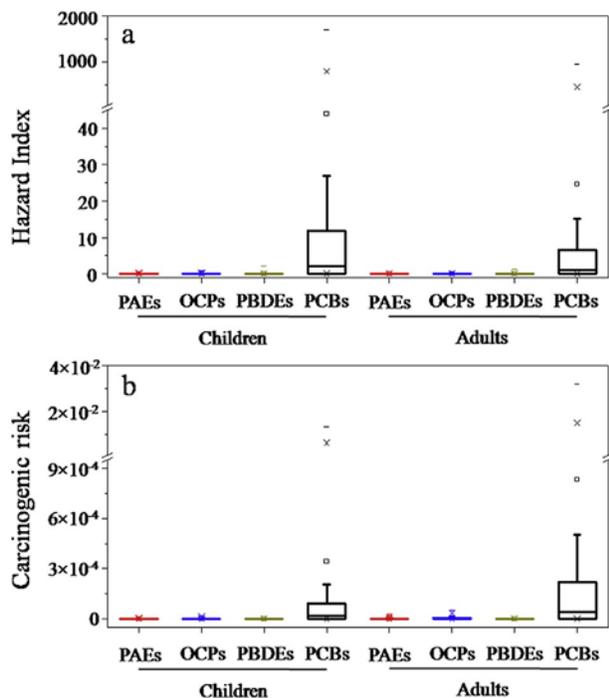


353

354 Fig. 2. (a) Canonical correspondence analysis of microbial data and the subset of 11 environmental
 355 variables (including water content, pH, TOC, total nitrogen, total phosphorus, copper, zinc, PCBs,
 356 PBDEs, OCPs, and PAEs); and (b) partial canonical correspondence analysis of the effects of

357 seven physicochemical parameters and four groups of organic compounds on the microbial
358 community composition in the YRD soils.

359



360

361 Fig. 3. Comparison of non-cancer exposure risk (a) and carcinogenic exposure risks (b) to adults
362 and children among selected toxic organic compounds including PCBs, OCPs, PBDEs, and PAEs.

363

364

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