

Aqueous Supramolecular Assemblies of Luminescent Cyclometalated Gold (III) Amphiphiles with Biocompatibility

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KEYWORDS: *Supramolecular Assembly, Cyclometalated Gold (III) Complex, Gold Amphiphile, Biocompatible, Supramolecular Nanostructure*

Abstract:

Gold (III) complex-based amphiphiles in aqueous media have recently been demonstrated with high structural sensitivity to external stimulations, enabling new prospect for soft functional material applications. Here we demonstrate an advanced supramolecular assembly system of gold (III) amphiphiles reversibly controlled by counterions exchange. More importantly, the luminescent properties of the gold (III) amphiphiles are enhanced by implementation of σ -donating mono-alkynyl

21 ligand. Gold (III) amphiphiles demonstrate with good cytocompatibilities to human bone marrow-
22 derived mesenchymal stem cells.

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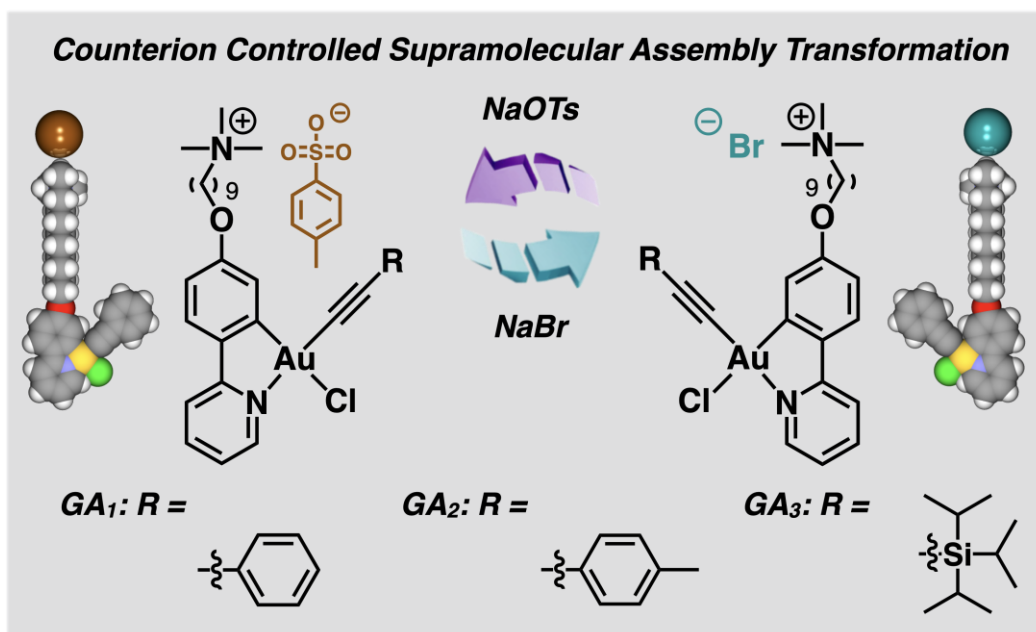
24 1. Introduction

25 Supramolecular assemblies are commonly found in biological systems to serve vital roles in
26 proper biological functions [1–3]. Synthetic supramolecular assemblies, as an alternative strategy to
27 its natural counterpart, provide structural versatility and advanced functions in response to non-
28 biocompatible external stimulations [4–7], *e.g.*, UV-light, pH, etc [8–10]. Amphiphile, featured with
29 a combination of hydrophilic and hydrophobic motifs, is a class of common synthetic organic
30 molecular designs in enabling tunable functions and potential aqueous solubility [11]. Organic
31 amphiphiles in aqueous media, such as chromophore-based [12–15], amino acid-based, and
32 photoresponsive molecular amphiphiles [4,16], have been extensively investigated in past few
33 decades. Metal-ligand amphiphiles have been demonstrated as a promising alternative strategy to
34 organic amphiphiles, due to the structural diversity and versatility of ligand modifications [17–21].
35 Fine adjustment of metal-ligand amphiphile design enables controls of supramolecular interactions
36 and its resulting supramolecular nanostructures [17,21].

37 Gold (I) and gold (III) complexes, featured with excellent luminescent properties and aqueous
38 stability, have been developed for applications in catalysis [22–26], bioconjugation methods [27–32],
39 and optoelectronic materials [33,34]. Gold complexes-based amphiphiles was firstly reported by Che
40 and co-workers [35], in using of cyclometalated gold (III) complexes linked to polyethylene glycol
41 (PEG) chain for improved aqueous solubility. Namely, the gold (III) amphiphiles form micellar
42 nanostructures for *in-vitro* phototoxicity [35]. Kinetically controlled supramolecular assemblies of
43 charged cyclometalated gold (III) amphiphiles in ACN/water media were reported by the same group
44 [36]. Charged cyclometalated gold (III) amphiphiles were further demonstrated by Yam, showing

45 intrinsic multiple responsiveness and organic gel-sol processes [37]. It is noted that these reported
46 cyclometalated gold (III) amphiphiles were highly sensitive to minor change of molecular structure
47 and subsequently induced supramolecular structural transformations [35–37]. On the basis of these
48 pioneering cyclometalated gold (III) amphiphile designs [38–42], we reported the first reversibly
49 counterion controlled supramolecular assembly transformations of cyclometalated gold (III)
50 amphiphiles in aqueous media [43]. However, the luminescent properties and cytocompatibilities of
51 this reversibly counterion controlled supramolecular assembly system remain unexplored.

52 Pioneered by Yam, cyclometalated gold (III) alkynyl complexes have been demonstrated
53 extensively with enhanced ambident luminescent properties, due to the strong σ -donating alkynyl
54 ligands rendering the metal center more electron rich [37,44]. Herein, we design and synthesize a new
55 series of cyclometalated gold (III) amphiphiles (GA) with enhanced luminescent properties in
56 aqueous media and biocompatibility. Cyclometalated gold (III) complex core is connected to a
57 quaternary ammonium ion with an alkyl-linker, inducing molecular phase separation for improved
58 aqueous solubility. Strong σ -donating alkynyl ligands are directly bonded to the cyclometalated gold
59 (III) complex core, enabling ambident luminescence of GAs. Large aspect ratio of supramolecular
60 nanostructures is formed upon dissolution of GAs into aqueous media. The packing parameters of
61 GAs can be tuned reversibly by counterions. Cytocompatibilities of GAs in aqueous media has been
62 clearly demonstrated with limited cytotoxicity at low concentration of GAs. By elucidating conditions
63 of supramolecular assembling processes, luminescent properties, and cytocompatibilities of GAs, it
64 could open up new prospects toward fabrications of stimuli-controlled supramolecular soft functional
65 materials.



66

67 **Scheme 1.** Schematic illustration of the design of cyclometalated gold (III) amphiphiles.

68

69 2. Materials and methods

70 2.1. Materials

71 All commercial reagents are purchased from Acros Organics, Sigma Aldrich and Tokyo Chemical
 72 Industry Co. Ltd, and were used as received unless otherwise specified. All reactions were performed
 73 under nitrogen unless otherwise specified. Analytical thin layer chromatography (TLC) was
 74 performed with Macherey-Nagel Silica gel 60 UV254 aluminium plates and visualization was
 75 accomplished by UV light (254 / 365 nm) or staining with phosphomolybdic acid followed by
 76 heating. Flash column chromatography was performed using Macherey-Nagel Silica gel 60 (230–400
 77 mesh). Deuterated solvents were purchased from Cambridge Isotope Laboratories Inc.

78

79 2.3. UV-vis Spectroscopy

80 UV-vis measurements were performed on Agilent Cary 60 UV-Visible Spectrophotometer with a 1
81 cm path length quartz cuvette. A Luma 40/8453 temperature-controlled cuvette holder with four
82 optical ports was mounted in the sample compartment of Agilent Cary 60 UV-Visible
83 Spectrophotometer. Measurement of all samples were carried out at 20 °C.

84

85 *2.4. Preparation of aqueous sample*

86 **GAs** (1 wt.%) was dissolved in fresh deionized water (DI water). The solution was heated at 50 °C
87 for 5 min, then slowly cooled to 20 °C at a rate of 1.0 °C/min to form assembled structure. For TEM
88 study, the annealed solution was further diluted to 0.2 wt.%.

89

90 *2.5. Transmission Electron Microscopy (TEM)*

91 TEM was performed on a JEOL Model JEM-2010 Transmission Electron Microscope with hair pin
92 type tungsten filament operating at 120 kV equipped with Gatan 794 CCD camera. TEM sample were
93 prepared by depositing sample solutions (5.0 μ L) onto a carbon grid (Micro to Nano, EMR Carbon
94 support film on copper, 400 square mesh) for 20 s. The sample solution was removed by blotting and
95 UranylLess EM stain solution (Electron Microscopy Science, 5.0 μ L) was directly deposited onto the
96 grid for 20 s and the stain was removed by blotting.

97

98 *2.6. Dynamic Light Scattering (DLS)*

99 Dynamic Light Scattering intensities of each sample were measured on a Wyatt Technology DynaPro
100 NanoStar. The scattering intensities were recorded as a parameter for assembly size, given that the
101 objects in solution are anisotropic and the models used by Wyatt software are fitting for spherical
102 objects. According to previously described procedures [12], the critical aggregation concentration

103 (CAC) of **GAs** is determined by the scattering intensities of the solutions of **GAs** (concentration: 1.0
104 $\times 10^{-3}$ to 1.0 mM) at 20 °C. The scattering rate was normalized by the concentration of the solution
105 to yield the molar scattering intensity (M Counts s⁻¹ m⁻¹). Ten replications were performed, and the
106 data was averaged to show the molar scattering intensity and corresponding standard deviation.

107

108 2.7 Fluorescence

109 Fluorescence measurements were performed on an Agilent G9800AA Cary Eclipse fluorescence
110 spectrophotometer with a 1 cm path length quartz cuvette. All measurements were carried out at room
111 temperature.

112

113 2.8 Cytotoxicity test

114 The cytotoxicity of **GAs** were characterized by 3-(4,5-dimethylthiazol-2-yl)-5-(3-
115 carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium (MTS) assay. Human bone-marrow
116 Mesenchymal Stem Cells (hBM-MSCs) were seeded in 96-well plate as a density of 3000 cells per
117 well. After incubated in growth media which contains Minimum Essential Medium (MEM Alpha, no
118 phenol red, Gibco, USA), 10% Fetal Bovine Serum (FBS, Gibco, USA) and 1% Antibiotic-
119 Antimycotic (Gibco, USA) for 12 h, different concentration of **GAs** was added and incubated with
120 cells for 24 h, respectively. For MTS assay, MTS solution was added to each well and incubated for
121 2 h at 37 °C and 5% CO₂. The absorbance of each well was measured at 490 nm using LEDETECT
122 96 microplate reader.

123

124

125

126 3. RESULT AND DISCUSSION

127 3.1. Design and Synthesis of GAs

128 GAs were designed with cyclometalated gold (III) complex core, functionalized with a σ -
129 donating alkynyl ligand, attached with a quaternary ammonium ion motif with a nonyl-linker with a
130 tosylate counterion (Scheme 1). It is noted that extended conjugated system decrease aqueous
131 solubilities of the resulting supramolecular assemblies, hence, mono-alkynyl ligand is employed in
132 this current design. The general synthetic route for gold (III) amphiphiles **GA**₁, **GA**₂, and **GA**₃ is
133 shown in supporting information. According to previously reported procedure [45–48], GA precursor
134 **1** can be obtained and purified in a satisfactory isolated yield. Dichloro-gold (III) complex core of **1**
135 was added with alkynes **2** in the presence of triethylamine and catalytic amount of copper iodide,
136 affording GAs with a mono-alkynyl ligand in modest yields (29–46%) after multiple column
137 purifications. The chemical structures of newly prepared gold (III) amphiphiles **GA**₁, **GA**₂, and **GA**₃
138 were characterized unambiguously using ¹H, ¹³C NMR and high-resolution mass spectrometry
139 (Figures S11–S15).

140 3.2. Luminescent Properties and Counterion Controlled Supramolecular Assemblies 141 Transformations of **GA**₁

142 A freshly prepared aqueous solution of **GA**₁ (11.6 mM, 1.0 weight%) was heated to 50 °C and
143 slowly cooled to 20 °C at a rate of 1.0 °C/min, *i.e.*, thermal annealing process. The aqueous solution
144 was diluted into a range of concentrations from 0.01 to 1.0 mM for the determination of the critical
145 aggregation concentration (CAC) by using a dynamic light scattering (DLS). The CAC of **GA**₁ was
146 determined as 50 mM (Figure S1). An aqueous solution of **GA**₁ (400 μ M) was heated to 50 °C and
147 slowly cooled to 20 °C at a rate of 1.0 °C/min, studied by UV-vis absorption spectroscopy (Figure
148 1a). The absorption maximum at 355 nm of **GA**₁ increased in the cooling process with a formation
149 of bathochromic-shifted shoulder band appearing at ~450 nm (Figure 1a, red-line). Though higher

150 concentration of **GA**₁ was required (1.0 mM) for adequate luminescence, an emission band at 470–
151 600 nm was observed, which was excited by 360 nm (Figures 1b). The vibronic structured emission
152 band originates from the intra-ligand excited state of the cyclometalated ligand. [37, 39–41, 44, 47–
153 48]. Additionally, **GA**₁ was dissolved in dichloromethane/methanol (1:1 volume ratio) to give an
154 absorption band at 300–390 nm with absorption maximum at 355 nm (Figure S2a). Using the identical
155 solvent system, **GA**₁ (1.0 mM) gives much lower intensity emission band at 420–600 nm (Figure
156 S2b) than that of **GA**₁ in aqueous media (Figure 1b). Given that GA precursor **1** in aqueous media
157 showed limited luminescent properties even at high concentration (Figure S3), the results indicated
158 that implementation of strong a σ -donating alkynyl ligand to the cyclometalated gold (III) complex
159 core can enhance the luminescent properties of **GA**₁ in aqueous media.

160 A thermal annealed aqueous solution of **GA**₁ (2.32 mM) was examined with negative-stained
161 transmission electron microscopy (TEM), revealing supramolecular nanofibers with hundreds of
162 nanometres to micrometres in length and ~ 9 nm in diameter (Figure 1c). The supramolecular
163 nanostructures of aqueous solution of **GA**₁ are highly similar to that of observed in GA precursor **1**
164 as reported previously, except the diameter of the supramolecular nanofibers increased in **GA**₁.
165 Thermal annealed supramolecular nanofibers of **GA**₁ (400 μ M, final concentration) was added with
166 aqueous solution of sodium bromide (2.0 equiv.) and subsequently heated to 50 °C and slowly cooled
167 to 20 °C at a rate of 1.0 °C/min. The absorption spectrum of the obtained solution shown absorption
168 maximum at 355 nm was significantly reduced with a shoulder band at 400–500 nm formed (Figure
169 1d). An aqueous solution of **GA**₁ (1.0 mM), prepared from the identical method in the presence of
170 sodium bromide, shows an emission band at 400–600 nm (Figure 1e) with significant reduced
171 intensity than that of observed in Figure 1b. The results indicated that counterion exchange from
172 tosylate to bromide induces increasement of packing parameters of **GA**₁ and subsequently changes
173 the packing structure from less organized nanofibers to well-organized nanotubes. In this connection,
174 luminescent property of **GA**₁ was partially quenched with the increased aggregation in the resulting
175 well-organized nanotubular structures. The TEM image of the solution **GA**₁ (2.32 mM), prepared

from above method, revealed entangled nanotubular nanostructures with outer diameter ~19 nm in the presence of sodium bromide (Figure 1f). Counterion controlled supramolecular transformation of **GA1** is clearly demonstrated, more importantly, it provides as a control of luminescent properties of **GA1** by simply counterion bromide addition. Furthermore, the nanotubes of **GA1** were subsequently reformed after addition of sodium tosylate (2.0 equiv.) as further counterion exchange (Figure S4), showing partial supramolecular transformation of nanotubes to nanofibers with a mixture of irregular aggregates. Incomplete supramolecular transformation of **GA1** is possibly attributed to the increased ion strength after a series of sodium salts additions.

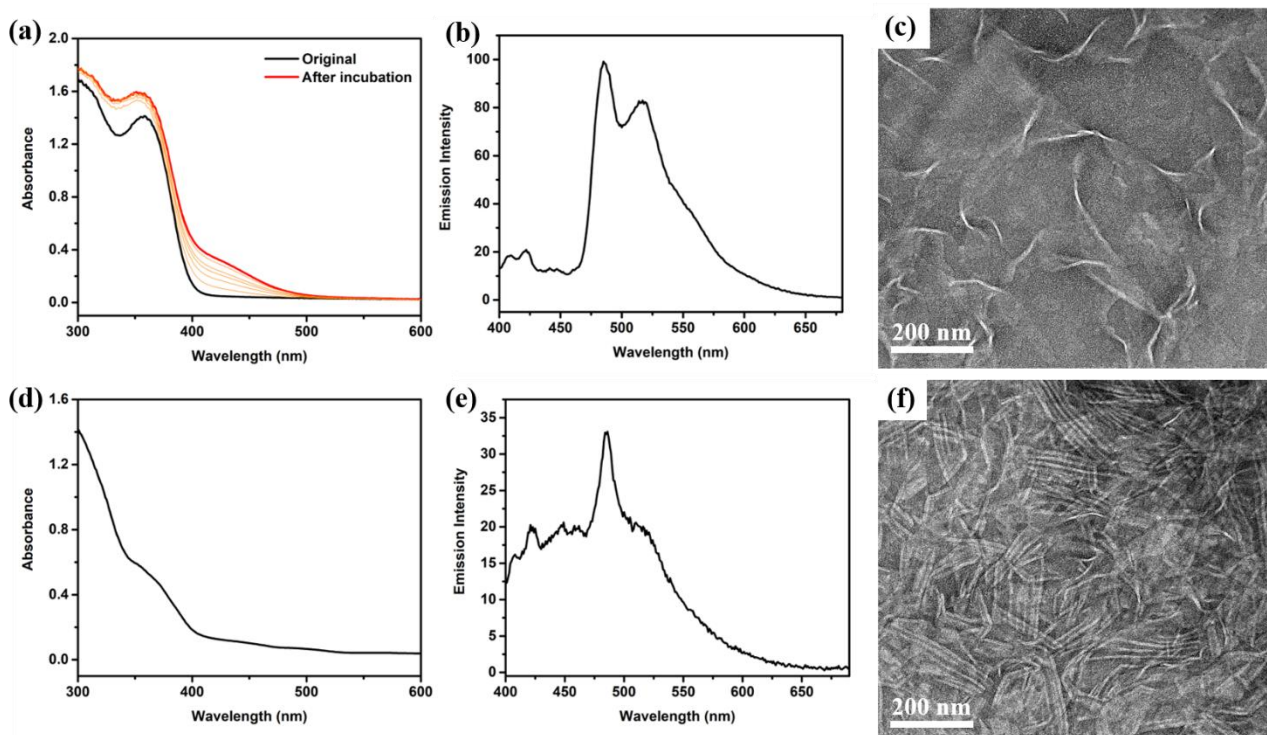
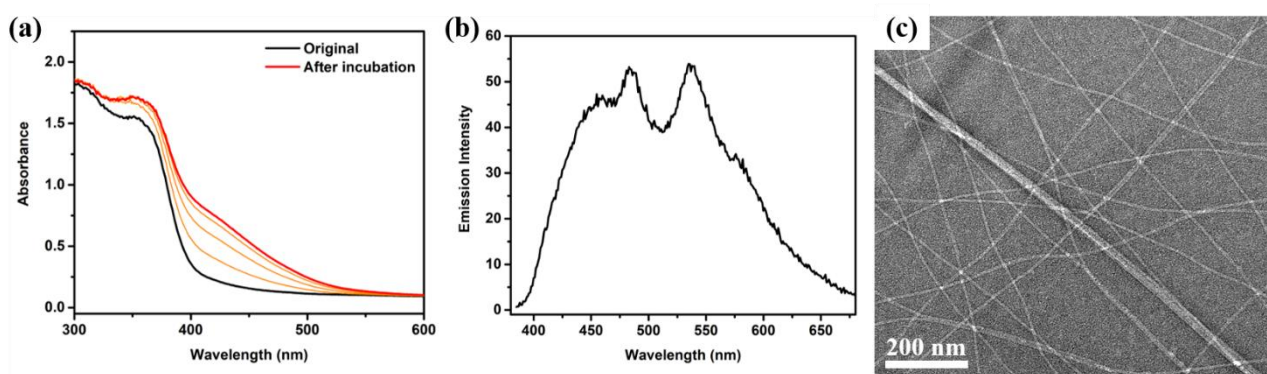


Fig. 1. (a) UV-vis absorption spectra of **GA1** (400 μ M) in DI water before (black-line) and after incubation, (red-line). (b) Emission spectrum of compound **GA1** (1 mM) in DI water. (c) TEM image of thermal annealed solution of **GA1** (2.32 mM). (d) UV-vis absorption spectrum of **GA1** (400 μ M) in DI water after addition of bromide counterion. (e) Emission spectrum of compound **GA1** (1 mM) in DI water after addition of bromide counterion. (f) TEM image of thermal annealed solution of **GA1** after addition of bromide counterion (2.32 mM).

3.3. Luminescent Properties and Supramolecular Assemblies of Structurally Derived GAs

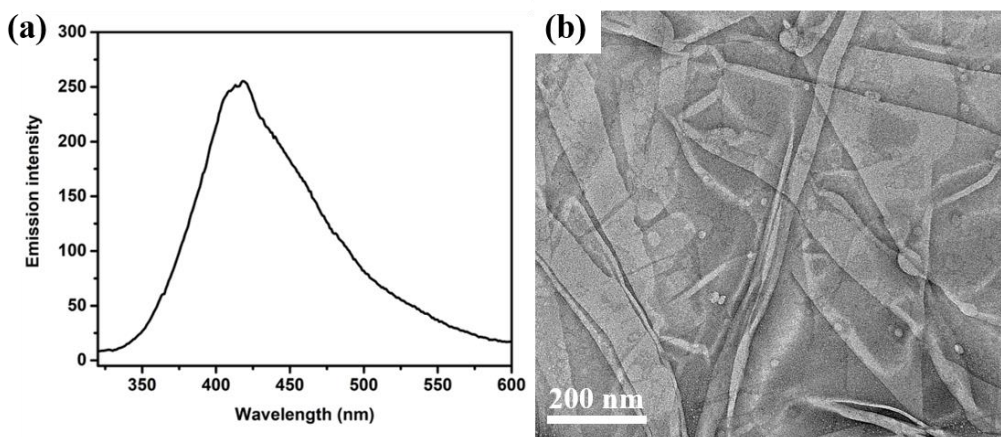
192 **GA₂**, a structurally derived GAs implemented with a 4-methylphenylacetylene ligand, was
 193 prepared according to the similar procedure, affording mono-alkynyl **GA₂**. **GA₂** was dissolved in
 194 organic media dichloromethane/methanol (1:1) to show strong absorption band at 320–390 nm with
 195 an absorption maximum at 356 nm (Figure S5a), while the identical solution shows luminescent
 196 properties with emission band at 450–550 nm (Figure S5b). In contrast, an aqueous solution of **GA₂**,
 197 thermally annealed at 50 °C and slowly cooled to 20 °C at a rate of 1.0 °C/min, showed enhanced
 198 absorption band at 320–500 nm (Figure 2a). Notably, the luminescent intensity of an aqueous solution
 199 of **GA₂** (1.0 mM) was reduced, possibly due to subtle change of packing structure with slightly
 200 increased steric hindrance of the alkynyl ligand. A pair of emission maxima were observed at 470 nm
 201 and 530 nm (Figure 2b), indicating aggregated GA can show blue-green color emission upon 320 nm
 202 excitation. Similar to **GA₁**, the vibronic structured emission band was also observed. The CAC of
 203 **GA₂** was determined as 50 mM (Figure S6). A thermally annealed aqueous solution of **GA₂** (2.28
 204 mM) was imaged under TEM (Figure 2c), revealing nanofibers structure with diameter ~ 6 nm
 205 essentially identical to that of observed in GA precursor **1** in aqueous media. Thermal annealed
 206 supramolecular nanofibers of **GA₂** (2.28 μ M, final concentration) was added with aqueous solution
 207 of sodium bromide (2.0 equiv.) and subsequently heated to 50 °C and slowly cooled to 20 °C at a rate
 208 of 1.0 °C/min. Nanofibers were transformed into a mixture of bundled nanofibers with vesicles
 209 (Figure S7), indicating supramolecular transformation of **GA₂** could be controlled by counterion
 210 addition.



211
 212 **Fig. 2.** (a) UV-vis absorption spectra of **GA₂** (400 μ M) in DI water before (black-line) and after

213 incubation, (red-line). (b) Emission spectrum of compound **GA**₂ (1 mM) in DI water. (c) TEM image
214 of thermal annealed solution of **GA**₂ (2.28 mM).

215 Given that cyclometalated gold (III) complex implemented with (triisopropylsilyl)acetylene
216 ligands enhance its luminescent properties [47], **GA**₃ was prepared from the identical synthetic
217 method with (triisopropylsilyl)acetylene ligands, giving a mixture of mono-alkynyl GA (**GA**₃) and
218 di-alkynyl GA (**GA**₃') of **GA**₃ in a ratio of 1:5 (Figure S8). It is noted that multiple column
219 chromatography separations have been performed but mono-alkynyl **GA**₃ could not be further
220 purified from di-alkynyl **GA**₃'. Absorption spectra of **GA**₃ in both organic (Figure S9a) and aqueous
221 (Figure S9b) are essentially identical to that of observed in **GA**₁ (Figure 1a). At 1.0 mM of mixture
222 of **GA**₃ in aqueous media, minor light scattering was observed, possibly due to reduced aqueous
223 solubility in the presence of higher ratio of di-alkynyl **GA**₃'. Significant enhanced luminescent
224 properties of **GA**₃ (1.0 mM) in aqueous media were observed along with an emission band at 350–
225 550 nm (λ_{max} = 420 nm) (Figure 3a), possibly due to high ratio of di-alkynyl **GA**₃'. Similar enhanced
226 luminescent intensity of **GA**₃ (1.0 mM) in DCM/MeOH (1:1 volume ratio) was observed (Figure
227 S9c). The CAC of **GA**₃ was determined as 20 mM (Figure S10). A thermal annealed aqueous solution
228 of **GA**₃ (1.97 mM) was imaged TEM to show a mixture of supramolecular nanosheet-like structures
229 and solid particles with hundreds of nanometers in diameter (Figure 3b). Nanosheet-like structures of
230 **GA**₃ is potentially constructed from the mono-alkynyl **GA**₃, while solid particles should be originated
231 from the low aqueous solubility di-alkynyl **GA**₃'. Although the complications of resulting
232 supramolecular assemblies of **GA**₃ remained challenging in study of the supramolecular
233 transformations, the results indicated the structural modifications, attaching σ -donating alkynyl ligand
234 to the cyclometalated gold (III) complex core, can improve the luminescent properties in the resulting
235 supramolecular assemblies.



236

237 **Fig. 3.** (a) Emission spectrum of compound **GA₃** (1 mM) in DI water. (b) TEM image of thermal
238 annealed solution of **GA₃** (1.97 mM).

239 3.4. Cytocompatibility of GAs

240 Aqueous solutions of **GA₁**, **GA₂**, and GA precursor **1** were prepared after thermal annealing
241 process and subjected for cytocompatibility studies, except **GA₃** due to its lower aqueous solubility.
242 An MTS mitochondrial activity assay was employed for determining cell viability of human bone
243 marrow-derived mesenchymal stem cells (hBM-MSCs) in the presence of aqueous solution of GA
244 precursor **1** in concentration range of 40–100 μ M (Figure 4a). In Figure 4a, the results were
245 normalized to viability of hBM-MSCs in the absence of GAs. High cell viabilities (~90%) of hBM-
246 MSCs over 24 h of culture with the solution of GA precursor **1** in concentration range of 40–100 μ M.
247 According to identical method, the cell viability of **GA₁** was investigated (Figure 4b), showing
248 increased cytotoxicity over 60 μ M of **GA₁**. **GA₂** showed reduced cytotoxicity to hBM-MSCs upon
249 incubation in concentration range of 40–100 μ M (Figure 4c). In considering the nanostructures of
250 GAs, GA precursor **1** and **GA₂** showed essentially identical nanofibers, while the nanofibers of **GA₁**

are larger in diameter. The results indicated a plausible dependence between the cytotoxicity of GAs and the resulting nanostructures of GAs.

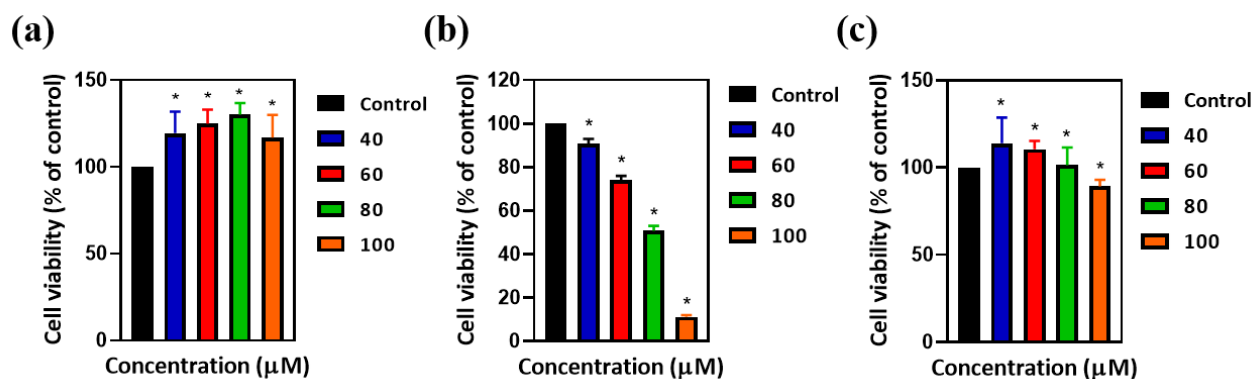


Fig. 4. (a) hBM-MSCs viability after incubated with (a) GA precursor **1**, (b) **GA₁** and (c) **GA₂** gradient solutions for 24 h, respectively. All values are mean \pm standard deviation of $n = 3$. * denotes $P < 0.05$, which represents for statistically difference compared to the control group.

4. CONCLUSION

Gold (III) amphiphiles, functionalized with σ -donating mono-alkynyl ligands, were designed to show enhanced luminescent properties in both organic and aqueous media. Nanofibers of **GA₁** and **GA₂** were confirmed with TEM and shown CAC below 50 μ M by DLS. Upon addition of NaBr, counterion exchanges enable supramolecular assembly transformations of **GA₁** from nanofibers to nanotubes, which is reversible upon further NaOTs counterion addition. Cyto compatibilities of **GA₁**, **GA₂**, and GA precursor **1** were clearly shown that supramolecular assemblies of GAs have limited cytotoxicity to the hBM-MSCs. The current approach would open up new generations of multi-functional supramolecular assembly of gold (III) amphiphiles.

Author Contributions

M.-H. Chau performed all the assembly characterizations for GAs in this work, while A. K.-H. Chan performed all organic synthesis and characterizations. Y. Ren carried out all cytotoxicity study. J. J.

269 Jiang performed preliminary study of GAs. M. K. Wong commented on the GA work. F. K.-C. Leung
270 co-wrote, conceived and supervised the research. All authors discussed the results and commented
271 on the manuscript.

272 **Conflicts of interest**

273 There are no conflicts to declare.

274 **Acknowledgements**

275 This work was supported financially by the Croucher Foundation (Croucher Innovation Award-2021
276 to F.K.C.L.), the Hong Kong Research Grant Council (GRF 15305822 to F.K.C.L.), (GRF 15300019
277 and 15300520 to M.K.W.), and The Hong Kong Polytechnic University (W08A, ZVST). We
278 acknowledge the technical support from UCEA and ULS of PolyU.

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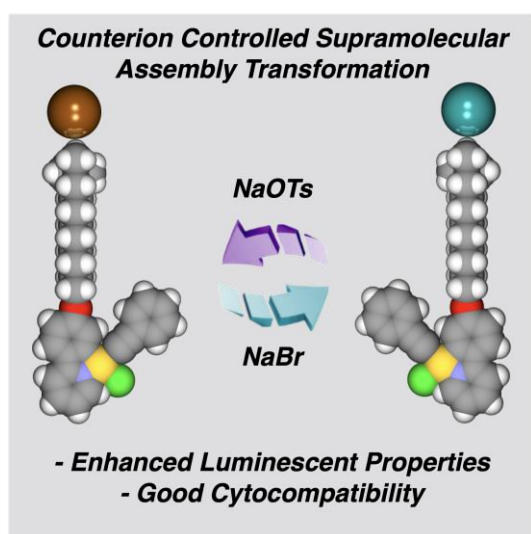
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426 **Table of Content / Graphical Abstract**



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