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Three-dimensional reduced-graphene

oxide/Fe₃O₄/Hydroxypropyl-β-cyclodextrin nanocomposite enhanced electrochemical sensing for the simultaneous quantification of serotonin, dopamine and ascorbic acid

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Abstract

In the present work, three-dimensional reduced graphene oxide containing Fe₃O₄ nanoparticles have been successfully decorated with hydroxypropyl-β-cyclodextrin (HP-β-CD) to construct a novel nanocomposite (3D-rGO/Fe₃O₄/HP-β-CD). The 3D-rGO/Fe₃O₄/HP-β-CD was further coated on a glassy carbon electrode (GCE) to design an electrochemical sensing platform for detecting simultaneously of serotonin (5-HT), dopamine (DA), and ascorbic acid (AA). The interconnected porous reduced graphene oxide framework tightly anchored to Fe₃O₄ magnetic nanoparticles warrants good electrical conductivity and efficient catalytic activity. The HP-β-CD acts as a supramolecular host with high recognition ability for 5-HT, DA and AA. Benefiting from the above charming peculiarity, well-separated oxidation peaks and increased peak currents were observed for 5-HT, DA, and AA individually and in mixtures by differential pulse voltammetry (DPV). The following figures of merit were found for simultaneous electrochemical determination of 5-HT, DA, and AA: (a) Well separated peaks at around 0.316, 0.16 and -0.044 V (vs. glassy

carbon electrode, GCE); (b) linear responses in the 0.01 - 25 μ M, 0.02 - 25 μ M and 10 - 350 μ M; (c) detection limits of 3.3 nM, 6.7 nM and 3.3 μ M (S/N = 3), and (d) recoveries of 96.9%-103%, 97.3%-102% and 96.3%-105% from spiked serum samples, respectively, and relative standard deviation (RSD) all less than 4%.

Keywords Three-dimensional reduced-graphene oxide; Magnetic nanoparticles; Hydroxypropyl-β-cyclodextrin; Electrochemical sensing platform; Simultaneous determination

1. Introduction

Serotonin (5-hydroxytryptamine, 5-HT) and ascorbic acid (AA) frequently coexist in biological fluids, which make the extremely vital contribution to maintaining the functions of human metabolism, central nervous and circulatory system [1, 2]. Dopamine (DA), known as an important neurotransmitter, widely exists in the brain for message transfer [3]. In general, low levels of DA can bring about neurological disorders including Parkinson's disease, schizophrenia and HIV infection [4, 5]. For the treatment of these illnesses, there is a demand to supply the appropriate amounts of DA to the patients and further to monitor the changes of DA in their serum regularly. However, 5-HT and AA are always co-present in serum, causing some interference for the detection of DA. Thus, it is of especial importance to develop a rapid and simple assay to determine concentration of DA in the presence of 5-HT and AA.

Currently, electrochemical methods show the bigger promise in the detection of small biomolecules compared to other strategies including fluorometric, chromatographic, spectrophotometric, chemiluminescent and so on [6]. Due to the minimally invasive, low-cost and rapid diagnoses for patients, it has been tried as the effective means for the analyses of active molecules in the biological systems [7-9]. Nevertheless, the biggest barrier for simultaneous electrochemical determination of 5-HT, DA and AA is the similar oxidation potentials of these three targets on general electrodes resulting in an overlapping oxidation peak. Unluckily, there only exist a few composite materials as the sensitive coating of electrodes or attached to electrodes by electrochemically pre-treated method, which have been explored to determine these three analytes simultaneously, such as polymers [10], acetylcholine [11] and carbon-spheres [12]. Consequently, the research interest has focused on exploiting suitable functionalized materials to further promote the sensitivity and selectivity of electrode for simultaneously detecting of these three biomolecules.

3D-graphene-based nanocomposites are considered to be a promising electrode material for electrochemical biosensor, thanking to their ultrahigh surface-area-to-volume ratio, improved electron transfer and mass transport [13-17]. Fe₃O₄, magnetic nanoparticles, possess considerable catalytic ability, together with great surface activity and favorable magnetic properties [18], it has been used to functionalize three-dimensional reduced graphene oxide (3D-rGO). By the preparation of Fe₃O₄ loaded 3D-rGO materials (3D-rGO/Fe₃O₄), it can enhance their existing merits synergistically [19-21]. Nevertheless, 3D-rGO/Fe₃O₄ composites suffer from the disadvantages of poor water solubility and lack of selectivity. This still hinders their great promise in electrochemical aspects, so it is highly desired to explore certain modifiers to improve their hydrophilicity, dispersion and recognizability. Interestingly, cyclodextrins (CDs) have commonly emerged as a functional water-soluble unit to improve the dispersibility and stability of graphene family nanomaterials and achieve remarkable electrochemical behavior by supramolecular selectivity [22, 23]. The CD-related host-guest interactions have potentially exerted a profound effect on the electrochemical sensing [24-26]. It has been demonstrated that CD and CD derivatives are highly efficient in specific molecular recognition and stereodifferentiation [27, 28], even revealing the inclusion interaction towards 5-HT [29], DA [30] and AA [30].

Herein, we develop an efficient strategy for the generation of HP-β-CD functionalized 3D-rGO/Fe₃O₄ (3D-rGO/Fe₃O₄/HP-β-CD) and explore its adhibition in the simultaneous electrochemical detection of 5-HT, DA and AA, as shown in scheme 1. The 3D-rGO/Fe₃O₄/HP-β-CD possesses a 3D network structure, high specific surface area, exceptional electronic conductivity and satisfactory capacity for biomolecules by virtue of the perfect unification great peculiarity of the three conjugated material. Thus, the 3D-rGO/Fe₃O₄/HP-β-CD coated glassy carbon electrode (GCE) exhibits a strong catalytic property for the electrooxidation of 5-HT, DA and AA, which can be applied to simultaneously detect of these three molecules by differential pulse voltammetry (DPV) mode. This sensing platform displays high sensitivity, wide linear range and low detection limit in simultaneous determination. Additionally, this electrochemical sensor was also selected to detect real sample with satisfactory recoveries, indicating a great potential of 3D-rGO/Fe₃O₄/HP-β-CD nanocomposite as electrode material in electroanalytical practices.

Experimental section

Materials and instruments

Ammonia solution (NH₃·H₂O, 25%), HP-β-CD, ferric chloride hexahydrate (FeCl₃·6H₂O), graphite powder, L-cysteine (L-Cys), 1-(3-dimethylaminopropyl)-3-ethylcarbodiimidehydro chloride (EDC·HCl), 4-dimethylaminopyridine (DMAP) were offered by Shanghai Macklin Biochemical Co. Ltd (http://www.macklin.cn/). 5-HT, DA and AA were provided by Aladdin Industrial Corporation from Shanghai (http://www.aladdin-e.com/). The human hepatoblastoma HepG2 Cells were taken from Shanxi Dayi Hospital. The ultrapure water has been supplied by Millipore system. The size and morphology of the nanomaterials were measured by SEM (S-4800, Hitachi, Japan) besides transmission electron microscopy (TEM, Jem-2100F, Jeol). The XRD (Bruker, D8 Advance), FT-IR (Bruker, Tensor), Raman (Horiba 800) and XPS (Thermo Scientific, ESCALAB 250Xi) have also been used to character the structure and elemental composition of nanocomposites. The electrochemical experiments were accomplished using an electrochemical workstation (CHI 660E, Chenhua, China) in the identical system encompassing three-electrode with our previous work [15]. All the details of electrochemical experiment information have been described in Electronic Supporting Material (ESM).

Fabrication of 3D-rGO/Fe₃O₄/HP-β-CD

Firstly, 3D-rGO/Fe₃O₄ (prepared in ESM) was sonicated in ultrapure water for several minutes so that it was completely dispersed. After activation the surface of carboxylic groups from 3D-rGO/Fe₃O₄ using EDC•HCl (46 mg, 0.24 mmol) for 3h, the appropriate amounts of HP-β-CD powder was added into above solution with a combination of DMAP (29 mg, 0.24 mmol) to make ester bonds between the carboxyl groups of 3D-rGO/Fe₃O₄ and hydroxyl groups of HP-β-CD. Then the vial was kept stirring for 24 h at room temperature to give 3D-rGO/Fe₃O₄/HP-β-CD. Finally, the nano-composites were washed with ultrapure water and anhydrous ethanol, collected with the aid of an external magnet repeatedly, and freeze-dried to yield a black powder.

Fabrication of 3D-rGO/Fe₃O₄/HP-β-CD/GCE

After carefully polishing the bare glassy carbon electrode (GCE, 3 mm in diameter) using alumina powder, and washing it with doubly deionized water and ethanol, the electrode was naturally dried. 3D-rGO/Fe₃O₄/HP-β-CD was dispersed in ultrapure water by ultrasonication, and 5 μL of the resulting 3D-rGO/Fe₃O₄/HP-β-CD suspension solution (2 mg·mL⁻¹) was modified onto clean GCE surface, and finally dried by infrared lamp. The 3D-rGO/Fe₃O₄ and 3D-rGO modified GCE were constructed with similar preparation process. The other electrochemical experiment information are presented in ESM.

Real sample analysis

The 3D-rGO/Fe₃O₄/HP- β -CD modified electrode was applied to detect 5-HT, DA and AA from human serum sample (provided by Shanxi Dayi Hospital) to evaluate its reliability for practical applications. Briefly, interfering proteins in pristine serum removed by centrifugation at 8000 rpm. Next, the serum samples were diluted 1:5 and 1:50 with 0.1 M phosphate buffer (PB) (pH = 7.0). 10 mL of diluted serum solution was transferred to the electrochemical cell for DPV analysis using 3D-rGO/Fe₃O₄/HP- β -CD/GCE to obtain the original amount of 5-HT, DA and AA in serum. Further tests were conducted for quantitative analysis with the standard addition method by adding different volumes of the standard solution of 5-HT, DA and AA. Then, the spiked 5-HT, DA and AA were also analyzed by DPV.

Results and discussion

Characterization of the 3D-rGO/Fe $_3$ O $_4$ /HP- β -CD

The surface morphology and size distribution of the 3D-rGO/Fe₃O₄/HP-β-CD were conducted by TEM besides SEM. From the SEM photograph in Fig. 1A, a well-defined polyporous and interconnected 3D framework was shaped by the crosslinked coordination and haphazard orientation of stacked rGO sheets. However, compared with the 3D-rGO/Fe₃O₄ (Fig. S1†), the introduction of HP-β-CD in 3D-rGO/Fe₃O₄/HP-β-CD hybrids failed to influence the 3D structure of our nanocomposite. Clearly, the TEM image (Fig. 1B) shows typical crumpled structure caused by a large amount of 3D-rGO nanosheets with many wrinkles. The dark spots corresponding to Fe₃O₄ nanoparticles seem to be uniformly embedded into the network of 3D-rGO. As can been seen from the HRTEM photograph (Fig. 1C), the thickness of the graphene sheets is

about 3 nm, and the average diameter of the Fe₃O₄ nanoparticle is estimated to be about 8 nm.

Further information is provided for the HP-β-CD successful cross-linked 3D-rGO/Fe₃O₄. FT-IR spectrogram of 3D-rGO/Fe₃O₄/HP-β-CD as well as pristine HP-β-CD, 3D-rGO and 3D-rGO/Fe₃O₄ are depicted in Fig. 2A. For HP-β-CD, the stretching vibration of O-H is exhibited a broad and strong peak at 3400 cm⁻¹, and the peak in position of 1647 cm⁻¹ is attributable to C=C stretching vibrations. The significant peaks between 1200 cm⁻¹ and 1000 cm⁻¹ are considered as the feature of HP-β-CD. Compared to 3D-rGO, the appearance of C=O at 1720 cm⁻¹ for 3D-rGO/Fe₃O₄ suggests that the OH groups of Fe₃O₄ reacted with the COOH groups situated at the sides of the 3D-rGO layers. Two featured bands at 1175 cm⁻¹ and 1020 cm⁻¹ are owed to the epoxy C-O and alkoxy C-O stretching vibrations, respectively. Besides, it also shows the bands due to the stretching vibrations (2927 cm⁻¹, 2971 cm⁻¹) and bending vibration (742 cm⁻¹) of C-H on the phenyl ring. Corresponding to the bare Fe₃O₄, the representative peak of stretching vibration of Fe-O bond at around 533 cm⁻¹ indicates that the Fe₃O₄ is successfully bound to the 3D-rGO. Thus, the typical cyclodextrin and Fe₃O₄ absorption features of 3D-rGO/Fe₃O₄/HP-β-CD exhibiting spectrogram conclude in the FT-IR the successful generation of $3D-rGO/Fe_3O_4/HP-\beta-CD$.

Raman spectroscopy is employed to identify the quality and structural characterization of graphene family materials [31]. Fig. 2B exhibits the various Raman spectrogram of GO, 3D-rGO, 3D-rGO/Fe₃O₄ and 3D-rGO/Fe₃O₄/HP-β-CD. For original GO, there exist two distinct bands approximately at 1346 cm⁻¹ (D band, for defect or disorder) and 1600 cm⁻¹ (G band), respectively. The ratio of D-/G-intensity (I_D/I_G) was expected to quantify the degree of disorder, and it was calculated to be 0.92 for GO. For 3D-rGO curve, I_D/I_G ratio increases on account of the synthesis of 3D framework in the process of reduction of GO resulting in the increasing of carbon cavities and disordered degree. In the case of 3D-rGO/Fe₃O₄, an enhanced I_D/I_G ratio (1.42) confirms the introduction of Fe₃O₄ nanoparticles onto the 3D network. After 3D-rGO/Fe₃O₄ was functionalized with HP-β-CD, it was observed that the I_D/I_G value distinctly decreases (1.01) in comparison with 3D-rGO/Fe₃O₄ as a result of the partial disappearance of defects during chemical functionalization. Additionally, a relatively weaker 2D band (2701 cm⁻¹) and a strong G band (1589 cm⁻¹) obviously lead to I_G/I_{2D} >1, indicating the fact that 3D-rGO/Fe₃O₄/HP-β-CD is multi-layered. Besides, XRD (Fig. S2) and XPS (Fig. S3) results also provide some crucial evidences for the successful

synthesis of 3D-rGO/Fe₃O₄/HP- β -CD. And biocompatibility patterns were performed and illustrated the nontoxicity of 3D-rGO/Fe₃O₄/HP- β -CD under the tested concentration conditions (Fig. S4).

Electrochemical measurement of the sensor fabrication

EIS was employed to determine the electrochemical characteristics during preparation process, shown in Fig. 3A. A Randles equivalent circuit was applied to fit the EIS data. As depicted in the inset of Fig. 3A, the circuit includes the electrolyte resistance between the working and reference electrodes (R_s), the double layer capacitance of electrode and electrolyte (C), Warburg impedance (Z_w), and electron transfer resistance (R_{et}) by the diffusion of ions from the electrolyte to the interface. The EIS value R_{et} of bare GCE is the largest (320 Ω, RSD: 2.3%), illustrating that the electron transfer of bare GCE is the most difficult one among the four nanomaterials. As to 3D-rGO/Fe₃O₄/GCE, its semicircle diameter is significantly smaller compared to the value of 3D-rGO/GCE, and the R_{et} of 68 Ω (RSD: 2.5%) was indeed less than that of 3D-rGO/GCE (166 Ω, RSD: 3.2%). This may result from the synergy linking of Fe₃O₄ and 3D-rGO that makes the interfacial electron transfer easy for 3D-rGO/Fe₃O₄/GCE. When GCE was modified by 3D-rGO/Fe₃O₄/HP-β-CD, the lowest R_{et} (15 Ω, RSD: 4.6%) value is obtained among all the modified electrodes, indicating that the grafting of HP-β-CD contributes to enhancing the transfer rate of 3D-rGO/Fe₃O₄/HP-β-CD nanohybrids.

Additionally, the effective surface areas (A) of 3D-rGO/Fe₃O₄/HP- β -CD/GCE and bare GCE were also explored by chronocoulometry in 0.1 mM K₃[Fe(CN)₆] containing 1.0 M KCl (diffusion coefficient, $D = 7.6 \times 10^{-6}$ cm²·s⁻¹, at 25 °C [32]). The corresponding values of A were calculated to be 0.7439 and 0.1758 cm², respectively, which obviously proves the effective area of the 3D-rGO/Fe₃O₄/HP- β -CD/GCE is approximately 4-fold high as much as that of the bare GCE. This further demonstrates that 3D-rGO/Fe₃O₄/HP- β -CD modified GCE can improve greatly surface area of the electrode (Fig. S5).

3D-rGO/Fe₃O₄/HP-β-CD/GCE for electrochemical detection of 5-HT, DA and AA

The possibility of simultaneous detection of 5-HT, DA and AA has been investigated by

using 3D-rGO/Fe₃O₄/HP-β-CD/GCE. Fig. 3B displays DPVs of bare GCE, 3D-rGO/GCE, 3D-rGO/Fe₃O₄/GCE and 3D-rGO/Fe₃O₄/HP-β-CD/GCE in 0.1 M PB, in which 10 μM 5-HT, 40 μM DA and 400 μM AA co-existed. Only a rather broad oxidation peak appeared at bare GCE, which leads to peak potentials for these three analytes using bare GCE are indistinguishable, further revealing the poor selectivity towards these three biomolecules. It is very apparent that the distinctly distinguishable peaks at 0.316, 0.16 and -0.044 V using 3D-rGO/Fe₃O₄/HP-β-CD/GCE corresponds to the respectively electrochemistry oxidation of 5-HT, DA and AA. The difference in anode peak potential between 5-HT and DA, DA and AA, 5-HT and AA are 0.156 V, 0.204 V and 0.36 V, respectively, which ensure the simultaneous detection of these three biomolecules by 3D-rGO/Fe₃O₄/HP-β-CD/GCE. In fact, the well-defined anodic peaks for three targets can also be given by 3D-rGO/Fe₃O₄/GCE and 3D-rGO/GCE, but the oxidation current for three molecules are much lower than those at 3D-rGO/Fe₃O₄/HP-β-CD/GCE. The increased current response at 3D-rGO/Fe₃O₄/HP-β-CD/GCE might be ascribed to the fact that HP-β-CD can give a unique host-guest interaction for the detected biomolecules, which evidently boost the electrochemical sensitivity and selectivity of electrode. Considering that 3D-rGO/Fe₃O₄/HP-β-CD/GCE displays the highest current response in terms of DPVs (Fig. 3B) and CVs (Fig. S6), the 3D-rGO/Fe₃O₄/HP-β-CD/GCE was applied to independent and simultaneous detection of 5-HT, DA and AA in the follow-up measurements via DPV mode. In addition, the exploration of the oxidation mechanism for analytes was performed, as shown in Fig. S7 and Fig. S8.

Optimization of determination conditions

To achieve the better sensing behavior for simultaneous determination of 5-HT, DA and AA, the following parameters were optimized in the experiment: (a) pH value of PB; (b) accumulation time; (c) volume of 3D-rGO/Fe₃O₄/HP- β -CD dispersion. Respective data and figures are provided in the ESM (Fig. S9-S12). Experimental determination conditions were found to give best results: (a) Optimal pH of PB: 7.0; (b) Optimal accumulation time: 6 min; (c) Optimal amount of the modifier: 5 μ L.

Electrochemical quantification of 5-HT, DA, AA

Furthermore, under the optimum experimental conditions mentioned above, the individual

and simultaneous quantitative determination of 5-HT, DA or AA was implemented respectively. Fig. 4 presents DPVs results of independent 5-HT (A), DA (B), AA (C) and three molecules mixed solution (D) achieved at according various concentrations by 3D-rGO/Fe₃O₄/HP-β-CD/GCE. And Fig. S13 displays the calibration plots corresponding to Fig. 4. A significant observation was made in terms of three analytes that the great linearity was obtained in various concentration ranges with the detection limits whether individual or simultaneous quantitative detection of 5-HT, DA and AA. All the quantitative analysis results of individual and simultaneous detection have been described in Table 1.

Noticeably, the similar slope of linear analytical curve for each molecule was acquired in the simultaneous and individual tests, which indicates that the 5-HT, DA and AA do not interfere each other in the simultaneous quantitative determination at 3D-rGO/Fe₃O₄/HP-β-CD/GCE. Table 2 shows that, in contrast to other sensors [8, 11, 12, 22, 23, 33-40], the 3D-rGO/Fe₃O₄/HP-β-CD-modified GCE shows superior distinguishability and high sensitivity.

Repeatability and stability and practical sample analysis

Repeatability of 3D-rGO/Fe₃O₄/HP-β-CD/GCE was estimated from the DPV response to simultaneously detecting of 20 μM 5-HT, 10 μM DA and 300 μM AA by parallel four measurements with the same electrode (Fig. S14). The relative standard deviations (RSD) of 5-HT, DA and AA have been calculated to be 3.9%, 2.6%, 3.2%, displaying satisfactory repeatability of the modified electrode. In addition, after the 3D-rGO/Fe₃O₄/HP-β-CD/GCE was kept in PB (0.1 M, pH 7) under 4 °C about 1 month, no obvious current response decay has been perceived (Fig. S15), exhibiting the stability the 3D-rGO/Fe $_3O_4$ /HP- β -CD/GCE. good of investigate the practicability of the 3D-rGO/Fe₃O₄/HP-β-CD/GCE, simultaneously detecting of sample and AA has been performed in human serum 3D-rGO/Fe₃O₄/HP-β-CD/GCE with the standard addition method. The mixed samples containing various concentrations of three small biomolecules are gradually added to the serum for the DPV recording. The quantitative results (Table S1) are calculated by the DPV detection of 5-HT, DA and AA in varied diluted serum samples by 3D-rGO/Fe₃O₄/HP-β-CD/GCE. Under the detected conditions, the concentrations of 5-HT and AA are respectively found to be 0.75 and 59.5 µM, whereas DA fails to be found in the pristine serum sample. The acceptable recoveries of 96.9%-103% were obtained for 5-HT, 97.3%-102% for DA and 96.3%-105% for AA, which confirm that the 3D-rGO/Fe₃O₄/HP-β-CD/GCE is credible for the practical sample detections.

Summary

A sensitive assay has been constructed by using 3D-rGO/Fe₃O₄/HP-β-CD modified GCE, and successfully applied to simultaneously detect of 5-HT, DA and AA. The 3D-rGO/Fe₃O₄ has a specific 3D-network structure, high conductivity and efficient catalytic activity, while HP-β-CD exhibits host-guest interactions with small biomolecules. Based on these unique characteristics, the 3D-rGO/Fe₃O₄/HP-β-CD biosensor provides enhanced peak current responses towards single 5-HT, DA and AA solution with wide linear ranges and limits of detection as low as 6.7 nM, 3.3 nM and 5 μM, however, the corresponding value are 3.3 nM, 6.7 nM and 3.3 μM for ternary mixture, respectively. Further application for the detections of 5-HT, DA and AA in serum samples was also demonstrated with satisfactory results. Thus, we anticipate that the present development has laid the foundation of potential application of the 3D graphene nanohybrid-based electrochemical biosensor in clinical detection and medical testing.

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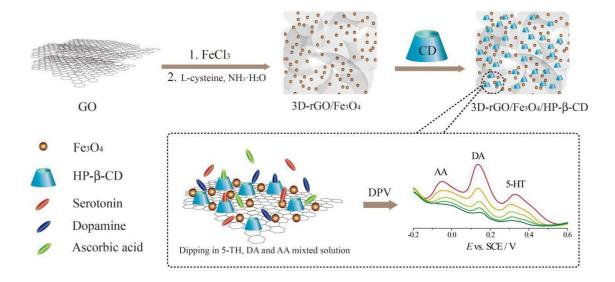
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Scheme 1. Schematic diagram for constructing 3D-rGO/Fe₃O₄/HP-β-CD, and simultaneous detecting serotonin (5-HT), dopamine (DA) and ascorbic acid (AA) by an electrochemical strategy.

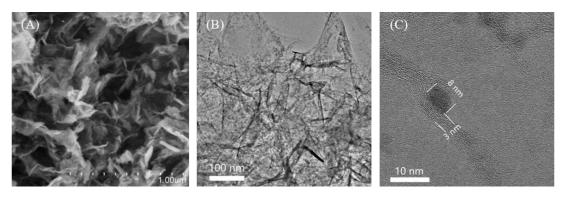


Fig. 1. SEM (A), TEM (B) and HRTEM(C) photographs of 3D-rGO/Fe $_3$ O $_4$ /HP- β -CD

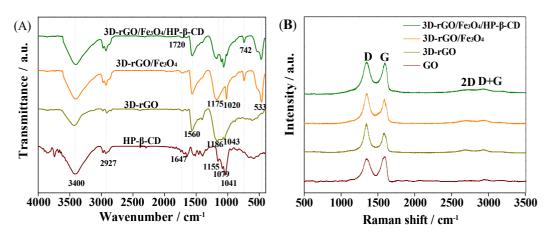


Fig. 2. (A) FT-IR spectrogram of HP- β -CD, 3D-rGO, 3D-rGO/Fe₃O₄ and 3D-rGO/Fe₃O₄/HP- β -CD; (B) XRD analysis of the 3D-rGO/Fe₃O₄ and 3D-rGO/Fe₃O₄/HP- β -CD.

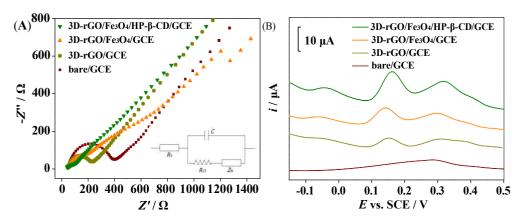


Fig. 3. (A) Nyquist plots of bare GCE, 3D-rGO, 3D-rGO/Fe₃O₄ and 3D-rGO/Fe₃O₄/HP- β -CD modified GCE in a 10.0 mL 0.1 M PB containing 5 mM [Fe(CN)₆]^{3-/4-} and 0.1 M KCl (pH 7.0) (Amplitude: 0.005 V, potential: 0.188 V, frequency range:10 mHz - 100 kHz.). (B) DPVs of 3D-rGO/Fe₃O₄/HP- β -CD/GCE, 3D-rGO/Fe₃O₄/GCE, 3D-rGO and bare GCE in 0.1 M PB of pH 7.0 containing 10 μM 5-HT, 40 μM DA, and 400 μM AA. (Amplitude: 0.05 V, increment potential: 0.004 V, pulse width: 0.05 s, pulse period: 0.5 s, quiet time: 2 s.)

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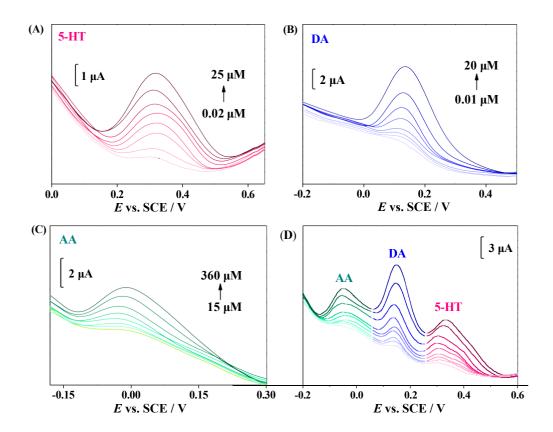


Fig. 4. Individual quantitative detection of 5-HT (A) or DA (B) or AA (C) and simultaneously quantitative detection of 5-HT, DA and AA (D) by 3D-rGO/Fe₃O₄/HP- β -CD/GCE in 0.1 M PB of pH 7.0. (Amplitude: 0.05 V, increment potential: 0.004 V, pulse width: 0.05 s, pulse period: 0.5 s, quiet time: 2 s.)

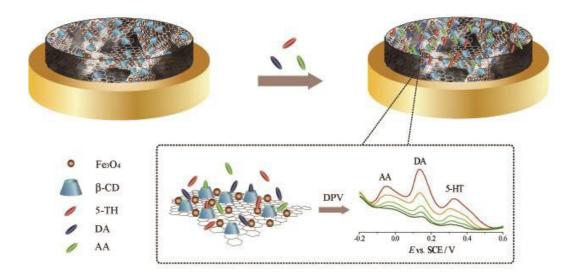
 $\textbf{Table 1.} \ Determination \ data \ analysis \ of individual \ and \ simultaneous \ detection \ of 5-HT, DA \ and AA.$

| Detection way | Analytes | Linear calibration curves | Linearity range/μΜ | Detection limits | |
|---|----------|-------------------------------------|----------------------|------------------|--|
| | | (linear correlation coefficient) | Emourity range, part | (S/N=3) | |
| Individual quantitative detection | 5-HT | $i (\mu A) = 0.33 + 0.11c (\mu M)$ | 0.02-25 | 6.7 nM | |
| | | $(R^2=0.9995)$ | 0.02-23 | O. / IIIVI | |
| | DA | $i (\mu A) = 0.20 + 0.39c (\mu M)$ | 0.01-20 | 3.3 nM | |
| | | $(R^2=0.9985)$ | 0.01-20 | | |
| | AA | $i (\mu A) = 0.22 + 0.01c (\mu M)$ | 15 260 | 5 M | |
| | | $(R^2=0.9990)$ | 15-360 | 5 μΜ | |
| Simultaneous quantitative detection | 5-HT | $i (\mu A) = 0.35 + 0.12c (\mu M)$ | 0.01-25 | 3.3 nM | |
| | | $(R^2=0.9987)$ | 0.01-23 | 3.3 nivi | |
| | DA | $i (\mu A) = 0.23 + 0.35c (\mu M)$ | 0.02.25 | (7. M | |
| | | $(R^2=0.9984)$ | 0.02-25 | 6.7 nM | |
| | AA | $i (\mu A) = 0.20 + 0.011c (\mu M)$ | 10.250 | 22.35 | |
| | | $(R^2=0.9988)$ | 10-350 | 3.3 μΜ | |

 Table 2. Comparison of analytical performance over various modified electrodes

| | Linear range (μM) | | Detection limit (μM) | | Detection | | | |
|--|-------------------|------------|----------------------|--------|-----------|------|--------------|------|
| Electrode | 5-HT | DA | AA | 5-HT | DA | AA | way | Kei. |
| EPPGE | 0.1-100 | 0.2-25 | 0.5-60 | 0.06 | 0.09 | 0.2 | simultaneous | 33 |
| ACh/GCE | 1-30 | 0.7-5 | 7-90 | 0.5 | 0.3 | 0.9 | simultaneous | 11 |
| CS/GCE | 40-750 | 20-150 | 300-2000 | 0.7 | 2 | 0.6 | simultaneous | 12 |
| GCE/P-Arg/ErGO/ | 0.01-0.5 | 0.001-0.05 | _ | 0.03 | 0.001 | _ | individual | 34 |
| AuNP | 1.0-10 | 1.0-50 | | | | | | |
| DNA-PPyox/CFE | 0.01-1 | 0.3-10 | _ | 0.007 | 0.05 | _ | individual | 35 |
| ERGO-P/GCE | 0.1-300 | 0.1-500 | _ | 0.0049 | 0.035 | _ | individual | 36 |
| NiO/CNT/PEDOT/ | 0.3-35 | 0.03-20 | _ | 0.063 | 0.026 | _ | simultaneous | 37 |
| GCE | | | | | | | | |
| MWNT-IE | 1-15 | 0.5-10 | _ | 0.2 | 0.1 | _ | simultaneous | 38 |
| CD/PNAANI/CNT | 4-200 | 4-200 | _ | <1 | <1 | _ | simultaneous | 23 |
| /CPE | | | | | | | | |
| 3DGLCFs/GCE | _ | 0.05-10 | 12.5-400 | _ | 0.01 | 2 | individual | 39 |
| β-CD/AuNPs/EDG | _ | 0.5-120 | 50-900 | _ | 0.024 | 2.0 | simultaneous | 22 |
| O/GCE | | | | | | | | |
| GO/TmPO ₄ /GCE | _ | 2-20 | 100-1000 | _ | 0.785 | 39.0 | simultaneous | 40 |
| AgNW-rGO/SPCE | _ | 40-450 | 45-1550 | _ | 0.26 | 0.81 | simultaneous | 8 |
| 3D-rGO/Fe ₃ O ₄ /HP- | 0.02-25 | 0.01-20 | 15-360 | 0.0067 | 0.0033 | 5 | individual | This |
| β-CD/GCE | 0.01-25 | 0.02-25 | 10-350 | 0.0033 | 0.067 | 3.3 | simultaneous | work |

Graphical abstract



Schematic presentation of simultaneous detecting serotonin (5-HT), dopamine (DA) and ascorbic acid (AA) for three-dimensional reduced-graphene oxide/Fe₃O₄/hydroxypropyl- β -cyclodextrin (3D-rGO/Fe₃O₄/HP- β -CD) by differential pulse voltammetry (DPV) approach.