1Advances in lignin valorization towards bio-based chemicals and fuels:

2Lignin biorefinery

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18Abstract:

19Lignin is one of the most promising renewable sources for aromatic hydrocarbons, while 20effective depolymerization towards its constituent monomers is a particular challenge 21because of the structural complexity and stability. Intensive research efforts have been 22directed towards exploiting effective valorization of lignin for the production of bio-based 23platform chemicals and fuels. The present contribution aims to provide a critical review of 24key advances in the identification of exact lignin structure subjected to various fractionation 25technologies and demonstrate the key roles of lignin structures in depolymerization for 26unique functionalized products. Various technologies (e.g., thermocatalytic approaches, 27photocatalytic conversion, and mechanochemical depolymerization) are reviewed and 28evaluated in terms of feasibility and potential for further upgrading. Overall, advances in 29pristine lignin structure analysis and conversion technologies can facilitate recovery and 30subsequent utilization of lignin towards tailored commodity chemicals and fungible fuels. 31**Keywords:** Circular bioeconomy; Waste to resources; Sustainable development; Waste

32management/recycling; Bio-based chemicals; Thermocatalytic depolymerization.

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341. Introduction

35Biomass conversion to valuable chemicals and biofuels is expected to make a significant 36contribution to the sustainability of energy supply (Chen et al., 2017a; Yu and Tsang, 2017; 37Petridis & Smith, 2018; Xiong et al., 2019). Lignocellulose is the most abundant form of 38biomass, mainly composed of varying portions of lignin, hemicellulose, and cellulose 39depending on biomass types (Shylesh et al., 2017). Over the years, hemicellulose (20-30%) 40and cellulose (30-40%) have been well studied, and some mature technologies have been 41industrialized for the production of biofuels and important chemicals from the 42(hemi)cellulose (Maki-Arvela et al., 2014; Xiong et al., 2017; Cao et al., 2018a). For 43example, cellulosic ethanol is the key emerging industry for 2nd generation bio-fuel (Yoo et 44al., 2017a), and our recent studies showcase vegetable/food waste-derived carbohydrates can 45be upgraded to chemical building blocks (Chen et al., 2017b; Chen et al., 2018; Yang et al., 462019; Yu et al., 2018; Yu et al., 2019).

47 Lignin, the largest renewable source of aromatics biopolymer on Earth, which accounts to 4815 to 40% dry weight in most plants, however, is mostly regarded as a low-value by-product 49in most biorefinery processes (Cao et al., 2017). One major source of lignin is provided by 50the pulp and paper manufacturer, where only 5 % of waste lignin has been employed for low-51grade fuel for heat and power applications through combustion (Cao et al., 2018b). Efficient 52lignin valorization from paper processing and biorefinery processes followed by targeted 53upgrading could significantly increase the economic viability and environmental 54sustainability (Rahimi et al., 2014; Wu et al., 2018).

The greatest challenge for the lignin-conversion technologies lies in the highly irregular 56polymeric structure. The complex three-dimensional amorphous polymer consisting of 57methoxylated phenylpropanoid units are relatively intractable (Li et al., 2015). Valorization 58of lignin often required organosolvolysis pretreatment of lignocellulosic biomass or 59enzymatic processing to isolate native lignin followed by hydrogenation/dehydrogenation to 60produce the desired aromatic products such as monomers, dimers, oligomers and bio-oil 61(Anderson et al., 2017; Qi & Su, 2014). However, the lignin isolated from different processes 62is varied in terms of molecular weight, polydispersity, solubility, abundance in functional 63groups, and type of inter-subunit linkages.

64 The physiochemical properties and structure difference can change lignin recalcitrance and65 impact the valorization of lignin. Several researchers articulated the impact of fractionation

66technologies on the subsequent product recovery, which underlined the key roles and 67advantages of native lignin with mild modified structure involved in subsequent valorization 68as well as bio-oil quality and yield (Anderson et al., 2019; Ragauskas et al., 2014). The 69aromatic monomers yield from direct hydrogenolysis of native lignin in biomass can reach up 70to 40%-50%, which is 3 to 10 times higher than that of the extracted lignin with the 71significantly modified structure (Shuai et al., 2016; Shao et al., 2018).

Recent studies have put forward energy-efficient biorefineries for high product yields by 72 73thermocatalytic approaches. The catalytic conversion strategies are mainly divided into 74acid/base catalyzed depolymerization, oxidative/reductive depolymerization, pyrolysis, and 75gasification (Shao et al., 2017; Wang et al., 2019a; Wang et al., 2019b; Luo et al., 2016). The 76advances in the catalytic processes have been based on the design of high-performance 77catalysts (e.g., well-defined and nanosized metal-composed materials) and advanced 78technologies, such as microwave assisted reaction, hydrothermal liquefaction, photocatalysis, 79mechanochemical depolymerization (Dabral et al., 2018b; Schutyser et al., 2018b; Sun et al., 802018). However, the poor stability of heterogeneous catalysts and complicated mechanism of 81homogeneous catalytic reactions need to be further studied. The bio-catalytic process is 82 regarded as an eco-friendly approach for the lignin depolymerization by taking advantage of 83the mild condition and environmental adaptability (Xu et al., 2018). However, the required 84specific condition and additional purification have brought difficulties and challenges for 85industrial production (Zhang et al., 2016). Overall, the previous works have highlighted a 86broad vision of promising lignin valorization emphasizing fractionation technologies and 87catalytic conversion.

The importance of native lignin structure in biosynthesis has been gradually recognized, 88 89which could govern downstream conversion for value-added chemicals (Anderson et al., 902019; Lancefield et al., 2018; Zaheer & Kempe, 2015). Therefore, this review focuses on the 91interplay of following important aspects during the course of lignin valorization, where 92various depolymerization technologies were described in relation to initial lignin structures in 93order to establish structure-activity relationship in lignin valorization (Fig. 1): (i) 94lignocellulose fractionation: identification of the exact structure of lignin with various 95extraction process; (ii) lignin depolymerization: mechanistic understanding and 96depolymerization technologies towards valuable aromatics and fuels; and (iii) further 97upgrading towards targeted chemicals.

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992. Lignin

1002.1 Structure and properties

101Lignin is mainly an irregular three-dimensional polymer created by free radical coupling 102among three monomers, syringyl (S), guaiacyl (G), and p-hydroxyphenyl (H) alcohol, which 103results in a variety of C-O and C-C linkages (Schutyser et al., 2018b). Unlike hemicellulose, 104where the biopolymer is relatively easy to be decomposed to pentose and hexose sugars with 105the typical polymerization degree in the range of 50-300 units, the highly polymerized lignin 106(up to 10,000 units) generates more rigid structure that is recalcitrant to chemical attack (Sun 107et al., 2018; Matsagar et al., 2017). The highly complex and unspecified structure of lignin 108has hampered lignin valorization for a long time. Structural studies revealed that β -O-4 109linkage is the most representative linkage in the native lignin, which makes up approximately

11050% of total linkages, together with other common bonding patterns including β -5 (1-12%), 111 β - β (5-12%), 5-5 (1-9%), 4-O-5, and β -1 (Li et al., 2015; Pu et al., 2011). Therefore, the 112cleavage of β -O-4 has been a key for lignin depolymerization strategies.

113 The structure and composition of lignin largely depend on the source of biomass. For 114example, softwood lignin contains a high portion of G units (80% - 90%), hardwood lignin 115has a mixture of G (25%-50%) and S (50%-70%) units, and grass lignin comprises a mixture 116of G (25-50%), H (10%-25%), S (25-50%) aromatic units (Li et al., 2015). Structural analysis 117of lignin and identification of nature linkages in lignin are essential for its efficient 118valorization to renewable fuels, materials or chemicals. Typically, the S/G-rich lignin is 119favorable to maximize monomer yields in biorefining applications (Rahimi et al., 2014; Shuai 120et al., 2016) because high S/G ratio can lessen the formation of inactive C-C bonds which 121leads to undesired condensed lignin during lignin depolymerization (Anderson et al., 2019). 122The lack of S units and the rich of G units in lignin may predict the abundance of β -O-aryl 123ether linkages during fractionation, which may undergo fast monomer coupling again.

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1252.2 Structural characterization

Nevertheless, the precise structures of lignin are also subjected to the fractionation and 127extraction methods from lignocellulosic biomass. For example, the conventional kraft pulping 128process can damage lignin structure showing significantly reduced levels of native lignin 129linkages and increased content of phenolic groups (Lancefield et al., 2018). The lignin 130structural modification induced by fractionation conditions is an important aspect to be 131concerned before subsequent valorization to value-added chemicals. The presence of 132different functionalities and units in lignin, as well as the molecular weight and 133polydispersity, needs to be analyzed to tailor the lignin valorization. A variety of 134characterization methods can be applied for different purposes, for instance, gel permeation 135chromatography (GPC) to determine molecular weights, FTIR spectroscopy to identify 136functional groups (Anderson et al., 2016). More recently, nuclear magnetic resonance (NMR) 137spectroscopic techniques such as 1D ¹³C, ³¹P spectroscopies and 2D HSQC/HMQC 138correlation have been extensively used for the elucidation of lignin structure and 139functionalities (Gioia et al., 2018; Pu et al., 2011). These findings have clearly shown that 140distinctive lignin structures upon various extraction methods. Even though NMR can 141elucidate lignin subunit composition and lignin interunit linkage distribution, there is still a 142lack of effective analysis of intermediate products during the reaction. Advanced *in situ* 143characterization and theoretical calculation could provide detail understanding of molecular 144reaction dynamics in the complex biorefineries.

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1462.3 Solid effect of lignin and catalyst

147 Poor solubility of lignin inevitably hinders the mass transfer and reaction efficiency during 148conversion, which also induces the complexity of the liquid-solid multi-phase system. 149Heterogeneous catalysts such as metal-based catalysts (e.g., Ni, Cu, Fe, and Pt-based 150catalysts) have been widely adopted for hydrogenation/dehydrogenation of lignin to obtain 151aromatic monomers in view of their high selectivity and reactivity in cleavage of C-O and C-152C bonds (Jongerius et al., 2013; Opris et al., 2017). However, the lack of effective contact 153between the solid catalyst and the biomass remains an issue, resulting in high energy154consumption mixing and limited conversion. In the current valorization system, employment 155of appropriate solvents and selection of the desired temperature is the key to overcome this 156limitation. A classical high temperature over 200 °C together with an organic solvent or 157under the assistance of acid/base is used to enhance the lignin solubility for subsequent 158conversion. However, the harsh conditions with high reaction temperature and high reaction 159pressure ($H_2/O_2/N_2$) make the active aromatic intermediates of lignin, such as reactive 160benzylic ketone at C_{α} position, susceptible to repolymerization and re-condensation (Shuai et 161al., 2016). Several strategies attempted to block reactive benzylic species to prevent the 162repolymerization, and the proposed mechanism will be described in detail below. Therefore, 163lignin depolymerization with a tradeoff between selectivity and activity remains a grand 164challenge for the production of value-added chemicals.

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1663. Fractionation

167 Separation of the desired composition from lignocellulose biomass is the prerequisite of 168lignin valorization. Extracting a soluble and uncondensed lignin substrate during biomass 169pretreatment can facilitate the production of lignin monomers and be compatible with other 170biorefining strategies. Research efforts are underway for: (i) maximal delignification and 171efficient removal of hemicellulosic fractions; (ii) advanced isolation of lignin in industrial 172applications for targeted components; and (iii) practical and efficient fractionation to improve 173biorefinery economic feasibility. Yet, lignin underwent organosolvolysis, enzymatic 174processing or recovered from pulping and paper industry presents various structures as a 175result of different fractionation methods, which indicate the potential influence on further 176 valorization.

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1783.1 Modified lignin

Modified lignin is primarily obtained from pulping industrial processing known as Kraft 179 180lignin and sulfite lignin, where the lignocellulosic biomass undergoes harsh treatments of 181high temperature and strong acid or base to improve product recovery. For the acid sulfite 182pulping process, the high reaction temperature (120 °C - 160 °C) and application of SO₃²⁻ or 183HSO₃⁻ under the acidic environment (72% sulfuric acid for extraction of Klason lignin) can 184significantly destroy the structure of native lignin (Matsagar et al. 2018a). The incorporated 185sulfonate groups (4% - 8%) in the modified lignin can improve the water solubility. However, 186the formed sulfonate groups could lead to the poisoning of metal-based catalyst, which may 187inhibit the catalytic conversion of lignin to produce aromatic chemicals. The pulping process 188under the base condition is typically named as the Kraft process (Hu et al. 2018). The 189hydrogen sulfide and hydroxyl anions can break β-O-4 linkages at 160 °C - 180 °C, where 190repolymerization of C-C bond formation occurs simultaneously (Renders et al., 2016a). The **191**obtained Kraft lignin has a lower molecular weight and enhanced water solubility as a result 1920f decreased content in β -O-4 linkages and increased amount of OH groups, while the 193exceptionally stable C-C linkages in the modified lignin increase the difficulty to produce 194related aromatic monomers. Therefore, novel catalytic reaction for the efficient valorization 195of modified lignin to valuable chemicals and fuels should be developed. The native lignin 196produced more likely from an experimental scale possesses a higher content of β -O-4 and β - β 197linkages that favors C-O cleavage.

1993.2 Native lignin

Employment of neutral and mild conditions in extraction facilitates the isolation of native 200 201lignin. Organic solvents such as alcohols and tetrahydrofuran have been utilized for 202promising biorefinery fractionation by improving biomass dissolution (Yoo et al., 2017b; 203Zhang et al., 2016). The high content of alcohol used in lignin extraction could suppress self-204condensation and yield high-quality native lignin (Lancefield et al., 2017). NMR HSQC 205correlation analysis revealed that the native structure can be obtained from n-butanol isolated 206lignin, making them particularly suitable for depolymerization to aromatic monomers. 207Compared to the Kraft process, organosolv lignin is more environmental-friendly in terms of 208generating less amount of acid or base wastes. Ionic liquids have been recently regarded as 209green solvents to enhance biomass dissolution, which possesses multiple advantages 210including lower melting points, tunable physical properties, negligible vapor pressure, and 211outstanding thermal stability (Liu et al., 2016, Matsagar et al. 2018b). These advanced 212properties can benefit the extraction of native lignin under the mild condition. For example, 213Prado et al. demonstrated the extraction of native lignin using triethylammonium hydrogen 214sulfate at 120 °C, which was subjected to oxidative reaction and produced phenolic-rich bio-215oil (Prado et al., 2016). Under these mild conditions, the native structure of lignin is protected 216 from re-condensation.

217 Milled-wood lignin (MWL) is one of the most representative native lignin in the plant cell 218wall since it is isolated under room temperature in the absence of any strong acid or base 219species (Yang et al., 2016). The biomass is first ball-milled and then extracted by the mixture 220of 1,4-dioxane and water (96/4, v/v). The recovery of lignin is mainly associated with ball 221milling time, where the yield increases from 16 % to 60 % when the ball milling time 222increases from one day to one week. However, this method mainly focuses on the recovery of 223lignin, which also includes some compromises like time-consuming and introducing a large 224number of carbohydrates coupled with the isolated lignin. To obtain a higher purity of native 225lignin, cellulolytic enzyme lignin (CEL) obtained from MWL or lignin-carbohydrate complex 226which were treated with cellulolytic enzyme under the mild conditions (pH 4.5-5.0) is most 227favorable (Yoo et al., 2017b; Zhang et al., 2016). The overall yield of purified lignin can 228reach up to 50 % with a high content of β -O-4 linkages (84.4 per 100Ar), which depicts a 229comparative advantage in biorefining applications (Yang et al., 2016).

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2314. Lignin depolymerization

232 In this section, we provide the discussion of the catalytic depolymerization, 233mechanochemistry, photocatalytic, and thermal technologies for the conversion of various 234types of lignin. An overview of these state-of-the-art methods is shown in Fig. 2. The 235oxidative depolymerization of lignin mainly exhibited the production of syringaldehyde; and 236vanillin through oxidative cleavage is one of the oldest methods reported for lignin 237depolymerization (Sun et al., 2018). The vanillin yield depends on the oxygen concentration 238and pH of the system. Whereas the reductive depolymerization of lignin usually leads to the 239formation of aliphatic compounds mainly 4-propylcyclohexanol. The hydrogenation and 240deoxygenation carried out for birch wood sawdust produce a mixture of C8 and C9 241cycloalkanes (Yan et al., 2008). The reductive depolymerization generally uses pure hydrogen 242gas, hydrogen donor chemicals (such as formic acid, sodium borohydride), or hydrogen 243donor solvents for the production fuels and bio-oil (Sun et al., 2018; Matsagar et al., 2019). 244The commercial Pd/C catalyst reported for the lignin fragmentation under ambient hydrogen 245pressure results in the formation of phenols and ethyl-substituted arenes (Gao et al., 2016). 246Similarly, the acid/base catalyzed depolymerization of lignin mainly produces bio-oil and 247phenolic lignin monomers as the main products (Deepa et al., 2014; Chaudhary et al., 2017). 248Furthermore, thermochemical methods such as pyrolysis and gasification used for lignin 249biomass leads to the production of bio-oil and syngas. Various methods used for lignin 250depolymerization shows that the product distribution, the types of lignin monomers produced, 251and their yields are heavily influenced by the method used for the lignin depolymerization.

252 Catalytic conversion of lignin for the sustainable utilization of biomass has drawn ever-253increasing attention. The key points in the design of the efficient catalytic process is to 254maximize the value of starting materials while minimizing waste generation and energy 255consumptions. Summary of advancing catalytic strategies including acid/base catalyzed and 256oxidative/reductive depolymerization process are shown in Table 1 (Entry 1-4: acid-catalyzed 257depolymerization; entry 5-8: base-catalyzed depolymerization; entry 9-12: oxidative 258depolymerization; and entry 13-16: reductive depolymerization). Thus far, reductive 259depolymerization has primarily been employed for the catalytic conversion of native lignin in 260high activity and selectivity. However, the high reaction temperature and high pressure (H₂/ 261N₂) required in the reductive reaction may lead to low functionalized aromatics unavoidably. 262Another important aspect for hydrogenation of lignin is the uncontrolled process in the 263presence of metal-based catalysts resulting in excessive hydrogenation of the aromatic rings. 264In comparison, the oxidative depolymerization involved milder conditions, the attractive 265advantage is the production of functionalized aromatics that allows facile separation and 266subsequent valorization.

Most methods for lignin depolymerization still focused on the cleavage of β-O-4 linkage, 268which makes up about 50% of all linkages in native lignin and has a low bond dissociation 26950-70 kcal/mol (Shuai et al., 2018). The cleavage of β-O-4 linkage generated aromatic 270monomers and reactive intermediates that are prone to subsequent repolymerization, forming 271bio-oil or biochar. The major challenge in developing efficient conversion of lignin is to 272prevent interunit C-C coupling by self-condensation and improve aromatic monomers yield. 273Intensive research efforts have been devoted to preventing repolymerization during lignin 274conversion. The addition of formaldehyde can block the reactive benzylic positions with a 275protecting agent during pretreatment, thus effectively preventing lignin condensation (Shuai 276et al., 2016). The oxidation of the C_α alcohol to a ketone can happen in the presence of formic 277acid at 110 °C, which yielded more than 60 wt% of aromatics by promoting cleavage of the β-278O-4 linkage (Rahimi et al., 2014).

For the most abundant Kraft lignin by-product generated from paper industry byproduct, it 280contained 11.0 wt% of methoxy groups. The selective utilization of methoxy groups to 281produce valuable compounds may provide a new method for the comprehensive utilization of 282lignin resources (Table 1, entry 17-18). Recently, it has been reported that CO and water can 283react with methoxy group in kraft lignin and organosolv lignin to form acetic acid over RhCl₃ 284catalyst. This reaction was conducted in ionic liquids at a mild reaction temperature (120 °C-285140 °C), and 87.5% yield of acetic acid without by-product can be achieved (Mei et al., 2862017). In another case, 99% purity of terephthalic acid can be obtained by the de-287methoxylation of lignin and the introduction of desired carboxy groups from obtained bio-oils 288(Song et al., 2019). The selective removal of methoxy in lignin enable the functionalization 289of complex aromatic chemicals into the target chemicals.

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2914.1 Acid/Base-catalyzed depolymerization

292 Dilute aqueous acids (e.g., H_2SO_4 , H_3PO_4 , and HCl), organic acids (e.g., formic acid) and 293solid acids (e.g., zeolite, and SiO₂-Al₂O₃ oxides) are known to catalyze depolymerization of 294lignin or lignocellulose at a mild temperature (Dong et al., 2014; Qu et al., 2015; Nandiwale 295et al., 2018; Wu et al., 2019). Introduction of dilute acid can improve the degree of 296delignification without significant structure destruction, and 40% of monomers yield was 297reached when using H_3PO_4 (Renders et al., 2016a). The proposed acid-catalyzed 298depolymerization mechanisms have been studied based on the native lignin (Lahive et al., 2992016; Shuai et al., 2016). In the presence of solid acid catalysts, the amount of acidic sites 300over the catalyst surface and the pore structure has a strong impact on lignin deoxygenation. 301The FTIR and NMR studies identified that high yield (52% - 60%) of aromatic monomers 302can be attributed to the cleavage of β-O-4 and α-O-4 ether linkages over the solid acid 303catalysts in the H₂O/CH₃OH (1/5 v/v) solvent mixture (Deepa & Dhepe, 2014).

Base-catalyzed depolymerization can be achieved by using various inexpensive and readily 305available reagents, such as NaOH, KOH, K₂CO₃, CaO and MgO, which are known to 306promote delignification by breaking ester linkages combined with enhanced lignin dissolution 307(Stiefel et al., 2017; Yoo et al., 2017b; Zhang et al., 2018a). Base-catalyzed depolymerization 308can break down the β -O-4 linkages by two stages, i.e., aerobic oxidation of the C α alcohol to 309the carbonyl group, and cleavage of β -O-4 bond followed by aromatic monomers formation 310(Dabral et al., 2018a; Schutyser et al., 2018b). Under the condition of a strong base, high 311reaction temperature and long reaction time may theoretically enhance the production of 312aromatic monomers, while the formation of solid residue also increases due to 313repolymerization. Severe repolymerization and oligomerization are the main problems in 314base-catalyzed reaction. In homogeneous reaction, the selectivity and yield of bio-based 315chemicals are particularly dependent on the reaction temperature, pressure, time, 316concentration of various base, and substrate/solvent ratio (Qi et al., 2017; Renders et al., 3172016a). Regarding solid bases, the strength of basic sites plays an important role in the 318enhancement of bio-oil and aromatic monomers yield. The enhanced concentration of basic 319sites over the NaX (Si/Al 1.2/1) catalyst promoted the conversion of alkaline lignin to 320produce low molecular aromatic products (51% yield) with abundant functional groups (e.g., 321-OCH₃, -CHO, and -CH₃ groups) (Chaudhary & Dhepe, 2017).

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3234.2 Oxidative depolymerization

324 From a general point of view, oxidative depolymerization of lignin is a valorization 325strategy that focuses on producing polyfunctional aromatic compounds, ranging from 326aromatic aldehydes to carboxylic acids, such as vanillin, syringaldehyde, and aromatic acid, 327which are alternative to fossil fuels derived chemicals. The oxidative cracking reaction 328includes the cleavage of the β -O-4 bonds, C-C bonds, aromatic rings, or other linkages within 329the lignin. Homogeneous transition metal complexes, metal oxides, oxygen, and hydrogen 330peroxide are the most popular oxidative reagents. The presence of NaOH solution and 331hydrogen peroxide is required to increase the rate of lignin degradation and the yields of 332phenolic compounds (Dai et al., 2018; Prado et al., 2016). The high percentage of vanillin 333product was at approximately 350 mg/L under the microwave assisted oxidative reaction by 334using Cu(OH)₂ and Fe₂O₃ catalysts in a NaOH/H₂O₂ system (Panyadee et al., 2018). In this 335regard, the relatively low yield of oxidation of lignin is the major limitation.

Nowadays, most of the studies focus on the oxidation of lignin model compounds. The 337studies of diaromatic structures can provide a better understanding of the selectivity and 338activity of monomers in various reaction systems (Lahive et al., 2016). Both β-O-4 linkage 339and S/G ratios, strongly correlated to lignin structures and properties, have been considered 340as the key factors for a variety of monomer yields (Yoo et al., 2017a). The model compounds 341with specific β-O-4 linkage and varying S/G ratio were most commonly reported (Deng et al., 3422015). When Co-based catalysts with nitrogen modified carbon support were applied in the 343oxidation of lignin model compounds, 96% yield of phenol was obtained without oxidative 344coupling (Luo et al., 2018). The reactivity of copper and vanadium-based catalysts for the 345oxidation of lignin model compounds were also explored, in which intermediate ketone 346benzoin methyl ether was formed over the vanadium catalyst and transformed into benzoic 347acid and methyl benzoate, whereas benzaldehyde and methyl benzoate were produced over 348the copper catalysts without intermediate formation (Sedai et al., 2011). The development of 349model compounds studies could provide insight into the reaction mechanism.

350 In contrast, industrial waste lignin contains mostly interunit C-C bonds, which are formed 351during biomass pretreatment or pulping processing with strong acids or bases. Higher bond 352dissociation energies of C-C bonds (75-118 kcal/mol) in comparison to those of C-O linkages 353(50-70 kcal/mol) poses challenges to lignin depolymerization under mild conditions. 354Commercial CoS₂ catalyst can break the inactive C-C linkage under reasonably low 355temperature (250 °C) (Shuai et al., 2018). A representative methylene-linked C-C model 356dimer achieved 88% yield of mainly two aromatic monomers within 1.5 h (Wang et al., 3572016). A two-step strategy for oxidative cleavage of lignin C-C bond to aromatic monomers 358was employed under oxygen in acetonitrile. The first step was the conversion of β-O-4 359alcohol to ketone and the second step was for the active and selective oxygenation of C_β-H. 360This oxidative process decreased the C_α-C_β (around 100 kJ/mol) bond energy, thus facilitating 361the cleavage of the C-C bond. Typically, the selectivity and activity based on the lignin 362model compounds showed extremely high values for producing monomers, which, however, 363may not be representative considering the high complexity of real lignin. New techniques for 364the most structurally relevant and well-defined lignin should be developed.

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3664.3 Reductive depolymerization

367 Reductive depolymerization is an effective strategy for lignin conversion owing to its 368excellent selectivity in lignin C-O and C-C bond cleavage (Van den Bosch et al., 2015). The 369products generated from this method own a high ratio of hydrogen to carbon (H/C), 370endowing it with great potential to produce bio-fuel. The char formation during lignin 371hydrogenolysis is relatively minor by reductive stabilization of reactive intermediates 372(Renders et al., 2016b). For instance, Pd/C catalyzed depolymerization can achieve up to 25 373wt% yield of syringol/guaiacol chemicals and suggested no significant loss of activity and 374selectivity after four cycles (Ouyang et al., 2018). The use of H-donating solvents, such as 375formic acid, and other stabilizing compounds, such as alcohol, has recently been shown to 376cleave β -O-4 linkage and stabilize the resulting reactive species *via* reductive pathways (Shao 377et al., 2018; Wu et al., 2019).

Studies on non-precious-metal-catalyzed depolymerization of lignin are in promising 378 379progress. Low-cost nanosized MoOx/CNT for the hydrogenolysis of enzymatic lignin under 380H₂ atmosphere produced up to 47 wt% aromatic monomers (Xiao et al., 2017). Ni-based 381catalysts have been widely applied in the industrial catalysis due to its cost-efficient and high 382 activity. For the reductive conversion of lignin, selectively cleavage of β -O-4 in the 383hydrodeoxygenation process can be promoted over Ni-based catalysts. HBEA zeolite with 384abundant Brønsted acid sites supported Ni-based catalysts could achieve selective 385hydrodeoxygenation of organosolv lignin to low molecular weight compounds (Kasakov et 386al., 2015). The proposed pathway of hydrogenolysis of β -O-4 bonds and aromatic rings to 387cyclic alcohols over Ni-based catalysts were identified by GC-MS and in situ IR analysis. 388Nevertheless, there are several studies on whether the addition of metal promotor to Ni 389catalysts has a positive effect on the catalytic activity. Previous studies reported that the 390" dilution effect" of NiFe alloy effectively reduce Ni particle size, thus providing more active 391sites for the reaction. Furthermore, the formed Fe²⁺/Fe⁰ redox cycle can contribute to 392maintaining Ni⁰ state to improve the catalytic stability of Ni-based catalysts (Kim et al., 2017; 393Margossian et al., 2017). Bimetallic Ni-Fe/AC catalyst demonstrated high efficiency in β-O-4 394cleavage from lignin model compounds without hydrogenation of aromatic rings. A yield of 39539.5 wt% aromatic monomers and 88 % selectivity to propylguaiacol were achieved under H₂ 396atmosphere when using birch wood sawdust as the lignin substrate (Zhai et al., 2017).

Recently, a chemical reduction of lignin fragments isolated from the lignocellulosic Recently, a chemical reduction of lignin fragments isolated from the lignocellulosic Recently, a chemical reduction of lignin (CUB) processes based on early-399stage catalytic conversion of lignin (ECCL) (Rinaldi et al., 2016). This method offered two 400distinct easily-separable and stable fractions, i.e., highly-aromatic lignin (lignin oil) and 401holocellulose. The holocellulose can be utilized as a hydrogen source for hydrodeoxygenation 402(HDO) reaction through gasification. The lignin oil made by HDO reaction consists of 403aromatics or alkane fuels depending on the reaction conditions (Cao et al., 2018c). The ECCL 404offered a new way for the efficient deconstruction of lignocellulosic biomass with a high 405atom economy. The CUB process can be carried out using either inexpensive Rany Ni 406catalyst or noble metal-supported catalyst, which leads to a reduction of reactive lignin 407fragments (i.e., reactive functional groups are deactivated) such as reduction of aldehyde 408functional groups into alcohol.

409 The product distribution and yields of biomass-based chemicals significantly depend on 410the biomass type, catalyst type, and reaction conditions of depolymerization. The lignin with 411higher S-units require less severe reaction conditions so it allows for a higher lignin monomer 412yield (Rinaldi et al., 2016, Van et al., 2015). A higher number of S-units in lignin would 413decrease the repolymerization during the reaction. Furthermore, a higher concentration of S-414units decreases the relative concentration of G-units, which reduces the complexity of the 415condensing units in lignin. The CUB based on ECCL is the best example of efficient catalytic 416delignification of lignin with higher S-unit concentration under less severe reaction 417conditions, thus it is a promising approach for lignin research in the future.

4194.4 Thermal depolymerization

4204.4.1 Microwave reaction

421 Microwave-assisted depolymerization is one of the most attractive techniques for the 422valorization of biomass owing to its rapid heating rate and energy efficiency (Yu et al., 4232017b). Table 2 outlines advantages and disadvantages of various techniques for lignin 424depolymerization. Microwave irradiation provides heating directly inside the reaction 425medium via efficient hot spots, which are more amenable to produce three high-quality 426products: bio-oil, gas, and biochar (Fan et al., 2013; Luo et al., 2017; Zhou et al., 2018). 427Microwave-assisted solvolysis of lignin and other biomass has been widely investigated over 428suitable catalysts under relatively mild reaction conditions (< 200 °C) (Shao et al., 2017). In 429 comparison with the traditional hydrothermal process, microwave-assisted depolymerization 430of lignin could give higher bio-oil yield (Zhou et al., 2018). Recently, microwave-assisted 431depolymerization of alkaline lignin was reported over hydrotalcite-based CuNiAl mixed 432oxides (Zhou et al., 2018), in which 60% of bio-oil was achieved at 160 °C within 80 min. 433The bio-oil contains phenolic monomers such as guaiacol, p-hydroxyacetophenon, 434hydroxyacetovanillon, and syringaldehyde as the main products. Microwave heating was also 435used for the isolation of lignin from softwood with dilute H₂SO₄ (Zhou et al., 2017), which 436offered high lignin yield (82 wt%) and purity (93%) within 10 min at 190 °C. This illustrates 437an advantage over conventional thermal treatments that usually take days. For the oxidative 438degradation of lignin model compounds, microwave irradiation could effectively accelerate 439the oxidative degradation rate (Pan et al., 2015). However, the scaling up of biorefinery using

440microwave heating requires further investigation as there are limited application experiences 441of industrial-scale reaction vessels.

4424.4.2 Hydrothermal liquefaction

443 Hydrothermal liquefaction (HTL) of biomass comprises the thermochemical conversion of 444a broad range of bio-based materials into a liquid product using water, the greenest solvent, as 445the reaction media, thus representing a promising and sustainable option for biomass 446valorization (Qian et al., 2018; Zhang et al., 2019). This process is typically conducted under 447subcritical conditions: moderate temperatures (150-350 °C) and relatively high pressures 448(0.5-25 MPa) (Younas et al., 2017). HTL is a thermochemical route allowing the conversion 449of biomass into bio-oil (Cao et al., 2016). The choice of catalysts and solvents plays a critical 450role in determining to which degree delignification takes place and to what extent the lignin 451conversion (Renders et al., 2016a). A base catalyst is often added to enhance the performance 452of the HTL process. Depolymerization of Kraft lignin in subcritical water by using ZrO₂, 453K₂CO₃, and KOH as catalysts could yield high percentage of bio-oils (Belkheiri et al., 2018). 454Subcritical conditions (300-370 °C) were compared with supercritical conditions (390-450 455°C) to further investigate the temperature effect on lignin decomposition, in which 456supercritical conditions tremendously increased the char formation although a higher degree 457of depolymerization was achieved (Yong & Matsumura, 2013). The HTL is a superior 458process than pyrolysis because bio-oil produced using the HTL process has relatively low 459oxygen content (i.e., high energy density bio-oil) than pyrolysis oil. Moreover, HTL can be 460used for wet biomass, unlike pyrolysis and gasification treatment that requires pre-drying of 461feedstock.

4624.4.3 Pyrolysis and gasification

463Pyrolysis and gasification are two of the most commonly used thermochemical methods for 464the conversion of biomass, lignin, and other residual waste into syngas, bio-oil, and biochar 465(Beneroso et al., 2017; Lee et al., 2017a; Lee et al., 2017b; You et al., 2017). Pyrolysis is 466often carried out in a range of 400 °C to 800 °C in the absence of oxidizing agents (oxygen or 467air) or under limited amount of oxygen/air (Yoo et al., 2018). The product distribution is 468greatly influenced by the temperature and time of pyrolysis. For example, in a slow pyrolysis 469(low temperature and long residence time) the charcoal yield increases while in fast pyrolysis 470(high temperature (>800 °C) and short residence time (< 2 s)) the bio-oil yield increases with 471many lignin-derived aromatic products. The efficiency of fast pyrolysis reaction is dependent 472upon reaction temperature, residence time, sweep gas, and flow rate. Microwave-assisted 473pyrolysis can provide better quality bio-products in shorter timespan compared to 474conventional pyrolysis (Beneroso et al., 2017). Pyrolysis temperature and microwave 475irradiation time are the key factors to produce a high yield of bio-oil and low content of 476biochar (Dai et al., 2017). In pyrolysis the product selectivity is low and separation of product 477is a challenge, which may limit its practical application for lignin depolymerization.

478 The gasification provides an alternative for the production of synthetic petroleum from 479biomass through subsequent conversion of syngas using the Fischer–Tropsch process. 480Gasification is a thermochemical conversion by partial oxidation under high temperature (> 481800 °C) (Yang et al., 2018). For biomass gasification, using air as oxidizing agent provides 482the producer gas, while the use of pure oxygen as oxidizing agent provides the syngas. The 483presence of H₂ and CO in gasification can provide a reductive condition that is favourable for 484lignin depolymerization. Acid pretreatment helps to break down the highly complex 485structures of lignin and improve the quality of the bio-oil with high alkyl-phenols content 486(Duan et al., 2018). However, for the lignin gasification, the production of coke and tar from 487aromatics is the major problem (Kawamoto et al., 2017), which may require properly 488designed catalysts to address the issue.

489

4904.5 Photocatalytic depolymerization

491 Current conversion strategies rely on high-temperature hydrogenation of lignin in presence 492of various metal-based catalysts. In nature, lignin is produced by photosynthesis and can be 493depolymerized to chemicals via photochemical reaction (Luo et al., 2016). There are few 494studies on the conversion of native lignin *via* solar energy-driven at room temperature. The β-495O-4 linkages in the lignin structure could be cleaved by an electron-hole coupled photo-redox 496mechanism over the CdS quantum dots, while cellulose/hemicellulose remained almost intact 497(Wu et al., 2018). Photocatalytic oxidation of lignin model compound is reported by merging 498visible light photo-redox and Pd catalysis. This dual catalytic method exhibited an efficient 499oxidation of lignin model compound to high yield of oxidized products under room 500temperature (Kärkäs et al., 2016). Furthermore, the photocatalytic oxidation-hydrogenolysis 501of β -O-4 model compounds is reported through dual light wavelength switching strategy for 502the selective cleavage of the C-O bond (Luo et al., 2016). In this one-pot method, 503photocatalytic oxidation of α-C–OH bonds occurs over Pd/ZnIn₂S₄ while hydrogenolysis of 504C–O bonds happens over TiO₂–NaOAc. An organo.metallic Ir catalyst could reductively 505cleave β-O-4 ketones *via* light stimulation (Nguyen et al., 2014). The concept of using solar

506light energy at room temperature and atmospheric pressure *via* photocatalysis to selectively 507convert lignin into value-added chemicals represents an innovative route in terms of energy 508and environmental aspects.

509

5104.6 Mechanochemical depolymerization

Mechanochemical activation generates high energy micro-environment due to the localized 511 512pressure and frictional heating provided by the kinetic energy (Muñoz-Batista et al., 2018). In 513comparison with the conventional thermal depolymerization, the mechanical process can 514significantly reduce the reaction time, which in turn is conducive to energy and cost savings. 515For the wet ball milling process, an oxidative approach to lignin depolymerization was used 516 with potassium hydroxide and toluene (Yao et al., 2018). IR and HSQC NMR spectroscopies 517suggested two-step oxidative depolymerization of Kraft lignin. Meanwhile, 518mechanochemistry can provide solvent-free condition and unique chemical processes for 519various reactions. A mechanochemical oxidation and cleavage reaction in lignin β-O-4 model 520compounds and lignin catalyzed by HO-TEMPO/KBr/Oxone has been developed under 521 milling conditions (Dabral et al., 2018b). A base-assisted ball milling process was also 522developed for the degradation of lignin (Kleine et al., 2013).

523

5245. Upgrading towards desired chemicals

525 A variety of useful products can be produced from lignin depolymerization, which includes 526aromatic monomers, bio-oil, biochar, syngas, heat, and energy. Syngas can be further 527processed using the Fischer-Tropsch process into methanol, dimethyl ether, and other 528chemical feedstocks (Sikarwar et al., 2016; Zhang et al., 2018b). Biochar as a secondary 529product of bioenergy production can be applied in the environmental maintenance, 530electrochemical or catalytic reactions as the functional support due to its unique chemical 531structure and properties (Xiong et al., 2017; Zhang et al., 2017; Shaheen et al., 2018; Vikrant 532et al., 2018). A few of the monomeric compounds obtained from various depolymerization 533approaches can be applied as such in end-use applications. For instance, vanillin is the most 534available monoaromatic phenol currently produced at an industrial scale from lignin. Around 53520000 tons of vanillin is produced per year, 15% of which coming from lignin (Fache et al., 5362015). Thus, vanillin has the potential to be a key renewable aromatic building block. 537Nonetheless, many depolymerization methods give rise to substituted phenolic compounds 538that require additional transformation route to marketable chemicals. Low-cost separation and 539purification of pure compounds from these complex streams are technically challenging, 540while the obtained quantities of a single compound are generally low. The intensive approach

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5436. Future prospect

544Biorefineries of cellulose and hemicellulose fractions from lignocellulose biomass are more 545mature than lignin biorefinery, while the latter is often regarded as a waste fraction from 546biorefinery processes. Therefore, a cost-effective integration of lignin biorefinery into the 547overall lignocellulose biorefinery can significantly enhance value and resource recovery and 548contribute to economic sustainability in the future. To achieve sustainable and cost-549competitive lignin biorefineries, the following prospects for future research needs are 550underlined within the framework of this review:

(i) Identify the authentic structure of lignin upon various treatment methods. The structure 552 and reactivity of the extracted lignin are probably governed by the biomass types and 553 extraction methods. Further understanding of structure and development of structural 554 characteristics is the key issue for efficient transformation and separation downstream. In 555 addition to 2D-NMR, GPC, FTIR, GC-MS, and LC-MS analytic techniques, density-556 functional theory, and reaction mechanisms should be further explored.

557 (ii) Most existing industrial processing conditions, such as Kraft pulping and 5580rganosolvolysis methods for the production of bio-ethanol, significantly modify or damage 559the lignin and lead to a substantial loss of β -O-4 structural units. The main target is to 560maximally retain the native β -O-4 linkages in lignin and prevent the formation of inactive C-561C bonds. Therefore, new fractionation methods that can deliver both high-purity sugars and 562native lignin streams in biomass separation should be developed.

563 (iii) The structure-activity relationship between lignin and its valorization products is still 564inconclusive. Many studies have been performed on pure and simple lignin model 565compounds such as phenol, guaiacol, and selected compounds with specific β-O-4 linkage, 566which urges to broaden the applicability of chemocatalytic conversions to more complex 567compounds and field-related lignin recovered from various industrial applications.

568

5697. Conclusions

570Lignin valorization technologies to produce value-added chemicals have attracted a great deal 571of attention. Lignin is a widely abundant, renewable feedstock comprising aromatic rings

572from nature. This contribution aims to critically review the state-of-the-art chemical and 573technical methodologies as well as the latest improvement of advanced analysis techniques 574and critical factors for efficient lignin depolymerization. This work also highlights the 575importance of future fractionation process that delivers native lignin, which can provide a 576competitive advantage for sustainable lignin valorization.

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582

583Appendix A. Supplementary data

584Supplementary data associated with this article can be found, in the online version.

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586References

5871. Anderson, E.M., Katahira, R., Reed, M., Resch, M.G., Karp, E.M., Beckham, G.T.,
Román-Leshkov, Y., 2016. Reductive Catalytic Fractionation of Corn Stover Lignin.
ACS Sustain. Chem. Eng. 4(12), 6940-6950.

5902. Anderson, E.M., Stone, M.L., Katahira, R., Reed, M., Beckham, G.T., Román-Leshkov, Y.,

591 2017. Flowthrough Reductive Catalytic Fractionation of Biomass. Joule 1(3), 613-592 622.

5933. Anderson, E.M., Stone, M.L., Katahira, R., Reed, M., Muchero, W., Ramirez, K.J.,
Beckham, G.T., Roman-Leshkov, Y., 2019. Differences in S/G ratio in natural poplar
variants do not predict catalytic depolymerization monomer yields. Nat. Commun.
10(1), 2033.

- 5974. Belkheiri, T., Andersson, S.-I., Mattsson, C., Olausson, L., Theliander, H., Vamling, L.,
 2018. Hydrothermal Liquefaction of Kraft Lignin in Subcritical Water: Influence of
 Phenol as Capping Agent. Energy Fuels 32(5), 5923-5932.
- 6005. Beneroso, D., Monti, T., Kostas, E.T., Robinson, J., 2017. Microwave pyrolysis of biomass601 for bio-oil production: Scalable processing concepts. Chem. Eng. J. 316, 481-498.
- 6026. Cao, L., Zhang, C., Hao, S., Luo, G., Zhang, S., Chen, J., 2016. Effect of glycerol as co-
- solvent on yields of bio-oil from rice straw through hydrothermal liquefaction.Bioresour. Technol., 220, 471-478.
- 6057. Cao, L., Zhang, C., Chen, H., Tsang, D.C.W., Luo, G., Zhang, S., Chen, J., 2017.
 606 Hydrothermal liquefaction of agricultural and forestry wastes: State-of-the-art review
 607 and future prospects. Bioresour. Technol. 245, 1184-1193.
- 6088. Cao, L., Yu, I.K.M., Chen, S.S., Tsang, D.C.W., Wang, L., Xiong, X., Zhang, S., Ok, Y.S.,
 609 Kwon, E.E., Song, H., Poon, C.S., 2018a. Production of 5-hydroxymethylfurfural
 610 from starch-rich food waste catalyzed by sulfonated biochar. Bioresour. Technol. 252,
 611 76-82.
- 6129. Cao, L., Yu, I.K.M., Liu, Y., Ruan, X., Tsang, D.C.W., Hunt, A.J., Ok, Y.S., Song, H.,
 613 Zhang, S., 2018b. Lignin valorization for the production of renewable chemicals:
 614 State-of-the-art review and future prospects. Bioresour. Technol. 269, 465-475.
- 61510. Cao, Z., Dierks, M., Clough, M.T., Daltro de Castro, I.B., Rinaldi, R., 2018c. A
 616 Convergent Approach for a Deep Converting Lignin-First Biorefinery Rendering
 617 High-Energy-Density Drop-in Fuels. Joule 2(6), 1118-1133.
- 61811. Chaudhary, R., Dhepe, P.L., 2017. Solid base catalyzed depolymerization of lignin intolow molecular weight products. Green Chem. 19(3), 778-788.
- 62012. Chen, S.S., Maneerung, T., Tsang, D.C.W., Ok, Y.S., Wang, C.H., 2017a. Valorization of
- biomass to hydroxymethylfurfural, levulinic acid, and fatty acid methyl ester byheterogeneous catalysts. Chem. Eng. J. 328, 246-273.
- 62313. Chen, S.S., Yu, I.K.M., Tsang, D.C.W., Yip, A.C.K., Khan, E., Wang, L., Ok, Y.S., Poon,
 624 C.S., 2017b. Valorization of cellulosic food waste into levulinic acid catalyzed by
 625 heterogeneous Brønsted acids: Temperature and solvent effects. Chem. Eng. J. 327,
 626 328-335.

- 62714. Chen, S.S., Yu, I.K.M., Cho, D.W., Song, H., Tsang, D.C.W., Tessonnier, J.P., Ok, Y.S.,
 Poon, C.S., 2018. Selective glucose isomerization to fructose via a nitrogen-doped
 solid base catalyst derived from spent coffee grounds. ACS Sustain. Chem. Eng. 6,
 16113-16120.
- 63115. Dabral, S., Engel, J., Mottweiler, J., Spoehrle, S.S.M., Lahive, C.W., Bolm, C., 2018a.632 Mechanistic studies of base-catalysed lignin depolymerisation in dimethyl carbonate.
- 632 Mechanistic studies of base-catalysed lignin depolymerisation in dimethyl carbonate.633 Green Chem. 20(1), 170-182.
- 63416. Dabral, S., Wotruba, H., Hernández, J.G., Bolm, C., 2018b. Mechanochemical Oxidation
 and Cleavage of Lignin β-O-4 Model Compounds and Lignin. ACS Sustain. Chem.
 Eng. 6(3), 3242-3254.
- 63717. Dai, L., Fan, L., Liu, Y., Ruan, R., Wang, Y., Zhou, Y., Zhao, Y., Yu, Z., 2017. Production
 of bio-oil and biochar from soapstock via microwave-assisted co-catalytic fast
 pyrolysis. Bioresour. Technol. 225, 1-8.
- 64018. Dai, J., Styles, G.N., Patti, A.F., Saito, K., 2018. CuSO₄/H₂O₂-Catalyzed Lignin
 641 Depolymerization under the Irradiation of Microwaves. ACS Omega 3(9), 10433642 10441.
- 64319. Deepa, A.K., Dhepe, P.L., 2014. Lignin Depolymerization into Aromatic Monomers over644 Solid Acid Catalysts. ACS Catal. 5(1), 365-379.
- 64520. Deng, W., Zhang, H., Wu, X., Li, R., Zhang, Q., Wang, Y., 2015. Oxidative conversion of
 lignin and lignin model compounds catalyzed by CeO₂-supported Pd nanoparticles.
 647 Green Chem. 17(11), 5009-5018.
- 64821. Dong, C., Feng, C., Liu, Q., Shen, D., Xiao, R., 2014. Mechanism on microwave-assistedacidic solvolysis of black-liquor lignin. Bioresour. Technol. 162, 136-41.
- 65022. Duan, D., Wang, Y., Dai, L., Ruan, R., Zhao, Y., Fan, L., Tayier, M., Liu, Y., 2017. Ex-situ
 catalytic co-pyrolysis of lignin and polypropylene to upgrade bio-oil quality by
 microwave heating. Bioresour. Technol. 241, 207-213.
- 65323. Duan, D., Ruan, R., Lei, H., Liu, Y., Wang, Y., Zhang, Y., Zhao, Y., Dai, L., Wu, Q.,
 Chang, S., 2018. Microwave-assisted co-pyrolysis of pretreated lignin and soapstock
 for upgrading liquid oil: Effect of pretreatment parameters on pyrolysis behavior.
 Bioresour. Technol. 258, 98-104.

65724. Fache, M., Boutevin, B., Caillol, S., 2015. Vanillin Production from Lignin and Its Use asa Renewable Chemical. ACS Sustain. Chem. Eng. 4(1), 35-46.

65925. Fan, J., De bruyn, M., Budarin, V.L., Gronnow, M.J., Shuttleworth, P.S., Breeden, S.,
Macquarrie, D.J., Clark, J.H., 2013. Direct microwave-assisted hydrothermal
depolymerization of cellulose. J. Am. Chem. Soc. 135(32), 11728-31.

66226. Fan, L., Chen, P., Zhou, N., Liu, S., Zhang, Y., Liu, Y., Wang, Y., Omar, M.M., Peng, P.,663 Addy, M., Cheng, Y., Ruan, R., 2018. In-situ and ex-situ catalytic upgrading of vapors

from microwave-assisted pyrolysis of lignin. Bioresour. Technol. 247, 851-858.

66527. Gao, F., Webb, J.D., Sorek, H., Wemmer, D.E., Hartwig, J.F., 2016. Fragmentation of
Lignin Samples with Commercial Pd/C under Ambient Pressure of Hydrogen. ACS
667 Catalysis 6(11), 7385-7392.

66828. Gioia, C., Lo Re, G., Lawoko, M., Berglund, L., 2018. Tunable Thermosetting Epoxies
Based on Fractionated and Well-Characterized Lignins. J. Am. Chem. Soc. 140(11),
4054-4061.

67129. Jongerius, A.L., Copeland, J.R., Foo, G.S., Hofmann, J.P., Bruijnincx, P.C.A., Sievers, C.,
Weckhuysen, B.M., 2013. Stability of Pt/γ-Al₂O₃ Catalysts in Lignin and Lignin
Model Compound Solutions under Liquid Phase Reforming Reaction Conditions.
ACS Catal. 3(3), 464-473.

67530. Hdidou, L., Khallouk, K., Solhy, A., Manoun, B., Oukarroum, A., Barakat, A., 2018.
676 Synthesis of CoFeO mixed oxides via an alginate gelation process as efficient
677 heterogeneous catalysts for lignin depolymerization in water. Catal. Sci. Technol.
678 8(21), 5445-5453.

67931. Hu, J., Zhang, Q., Lee, D.J., 2018. Kraft lignin biorefinery: A perspective. Bioresour.680 Technol. 247, 1181-1183.

68132. Huang, X., Atay, C., Korányi, T.I., Boot, M.D., Hensen, E.J.M. 2015. Role of Cu–Mg–Al

682 Mixed Oxide Catalysts in Lignin Depolymerization in Supercritical Ethanol. ACS683 Catalysis, 5(12), 7359-7370.

68433. Kärkäs, M.D., Bosque, I., Matsuura, B.S., Stephenson, C.R.J., 2016. Photocatalytic
685 Oxidation of Lignin Model Systems by Merging Visible-Light Photoredox and
686 Palladium Catalysis. Org. Lett. 18(19), 5166-5169.

68734. Kasakov, S., Shi, H., Camaioni, D.M., Zhao, C., Baráth, E., Jentys, A., Lercher, J.A.,
2015. Reductive deconstruction of organosolv lignin catalyzed by zeolite supported
nickel nanoparticles. Green Chem. 17(11), 5079-5090.

69035. Kawamoto, H., 2017. Lignin pyrolysis reactions. J. Wood Sci. 63(2), 117-132.

69136. Kim, S.M., Abdala, P.M., Margossian, T., Hosseini, D., Foppa, L., Armutlulu, A., van

Beek, W., Comas-Vives, A., Coperet, C., Muller, C., 2017. Cooperativity and

693 Dynamics Increase the Performance of NiFe Dry Reforming Catalysts. J. Am. Chem.694 Soc. 139(5), 1937-1949.

69537. Kleine, T., Buendia, J., Bolm, C., 2013. Mechanochemical degradation of lignin and 696 wood by solvent-free grinding in a reactive medium. Green Chem. 15(1), 160-166.

69738. Koranyi, T.I., Huang, X., Coumans, A.E., Hensen, E.J., 2017. Synergy in Lignin
Upgrading by a Combination of Cu-Based Mixed Oxide and Ni-Phosphide Catalysts
in Supercritical Ethanol. ACS Sustain. Chem. Eng. 5(4), 3535-3543.

Zubive, C.W., Deuss, P.J., Lancefield, C.S., Sun, Z., Cordes, D.B., Young, C.M., Tran, F.,
Slawin, A.M., de Vries, J.G., Kamer, P.C., Westwood, N.J., Barta, K., 2016. Advanced
Model Compounds for Understanding Acid-Catalyzed Lignin Depolymerization:
Identification of Renewable Aromatics and a Lignin-Derived Solvent. J. Am. Chem.
Soc. 138(28), 8900-11.

70540. Lancefield, C.S., Panovic, I., Deuss, P.J., Barta, K., Westwood, N.J., 2017. Pre-treatment
of lignocellulosic feedstocks using biorenewable alcohols: towards complete biomass
valorisation. Green Chem. 19(1), 202-214.

Zonsteind, C.S., Wienk, H.L.J., Boelens, R., Weckhuysen, B.M., Bruijnincx, P.C.A.,
2018. Identification of a diagnostic structural motif reveals a new reaction
intermediate and condensation pathway in kraft lignin formation. Chem. Sci. 9(30),
6348-6360.

71242. Lee, J., Kim, K.-H., Kwon, E.E., 2017a. Biochar as a Catalyst. Renew. Sustain. EnergyRev. 77, 70-79.

71443. Lee, J., Yang, X., Cho, S.-H., Kim, J.-K., Lee, S.S., Tsang, D.C.W., Ok, Y.S., Kwon, E.E.,

715 2017b. Pyrolysis process of agricultural waste using CO₂ for waste management,

energy recovery, and biochar fabrication. Appl. Energy 185, 214-222.

71744. Li, C., Zhao, X., Wang, A., Huber, G.W., Zhang, T., 2015. Catalytic Transformation of718 Lignin for the Production of Chemicals and Fuels. Chem. Rev. 115(21), 11559-624.

71945. Liu, F., Liu, Q., Wang, A., Zhang, T., 2016. Direct Catalytic Hydrogenolysis of Kraft
Lignin to Phenols in Choline-Derived Ionic Liquids. ACS Sustain. Chem. Eng. 4(7),
3850-3856.

72246. Lu, J., Liu, Z., Zhang, Y., Savage, P.E., 2018. Synergistic and Antagonistic Interactions
during Hydrothermal Liquefaction of Soybean Oil, Soy Protein, Cellulose, Xylose,
and Lignin. ACS Sustain. Chem. Eng. 6(11), 14501-14509.

72547. Luo, N., Wang, M., Li, H., Zhang, J., Liu, H., Wang, F., 2016. Photocatalytic Oxidation–
Hydrogenolysis of Lignin β-O-4 Models via a Dual Light Wavelength Switching
Strategy. ACS Catal. 6(11), 7716-7721.

72848. Luo, Y., Fan, J., Budarin, V.L., Hu, C., Clark, J.H., 2017. Microwave-assisted
hydrothermal selective dissolution and utilisation of hemicellulose in Phyllostachys
heterocycla cv. pubescens. Green Chem. 19(20), 4889-4899.

73149. Luo, H., Wang, L., Li, G., Shang, S., Lv, Y., Niu, J., Gao, S., 2018. Nitrogen-Doped
732 Carbon-Modified Cobalt-Nanoparticle-Catalyzed Oxidative Cleavage of Lignin β-O-4
733 Model Compounds under Mild Conditions. ACS Sustain. Chem. Eng. 6(11), 14188734 14196.

73550. Maki-Arvela, P., Simakova, I.L., Salmi, T., Murzin, D.Y., 2014. Production of lactic
acid/lactates from biomass and their catalytic transformations to commodities. Chem.
Rev. 114(3), 1909-71.

73851. Margossian, T., Larmier, K., Kim, S.M., Krumeich, F., Müller, C., Copéret, C., 2017.
Supported Bimetallic NiFe Nanoparticles through Colloid Synthesis for Improved
Dry Reforming Performance. ACS Catal. 7(10), 6942-6948.

74152. Matsagar, B.M., Hossain, S.A., Islam, T., Alamri, H.R., Alothman, Z.A., Yamauchi, Y.,
742 Dhepe, P.L., Wu, K.C.W., 2017. Direct Production of Furfural in One-pot Fashion
743 from Raw Biomass Using Brønsted Acidic Ionic Liquids. Sci. Rep. 7(1), 13508.

74453. Matsagar, B.M., Hossain, S.A., Islam, T., Yamauchi, Y., Wu, K.C.W., 2018a. A novel
745 method for the pentosan analysis present in jute biomass and its conversion into sugar
746 monomers using acidic ionic liquid. J. Visualized Exp. (136), e57613.

74754. Matsagar, B. M., Van Nguyen, C., Hossain, M.S.A., Islam, M.T., Yamauchi, Y., Dhepe,
P.L., Wu, K.C.W., 2018b. Glucose isomerization catalyzed by bone char and the
selective production of 5-hydroxymethylfurfural in aqueous media. Sustainable
Energy Fuels. 2, 2148–2153

75155. Matsagar, B.M., Kang, T.-C., Wang, Z.-Y., Yoshikawa, T., Nakasaka, Y., Masuda, T.,
752 Chuang, L.-C., Wu, K.C.W., 2019. Efficient liquid-phase hydrogenolysis of a lignin
753 model compound (benzyl phenyl ether) using a Ni/carbon catalyst. Rea. Chem. Eng.
754 4(3), 618-626.

75556. Mei, Q., Liu, H., Shen, X., Meng, Q., Liu, H., Xiang, J., Han, B., 2017. Selective
756 Utilization of the Methoxy Group in Lignin to Produce Acetic Acid. Angew. Chem.
757 Int. Ed. 56(47), 14868-14872.

75857. Muñoz-Batista, M.J., Rodriguez-Padron, D., Puente-Santiago, A.R., Luque, R., 2018.
759 Mechanochemistry: Toward Sustainable Design of Advanced Nanomaterials for
760 Electrochemical Energy Storage and Catalytic Applications. ACS Sustain. Chem.
761 Eng. 6(8), 9530-9544.

76258. Nandiwale, K.Y., Danby, A.M., Ramanathan, A., Chaudhari, R.V., Subramaniam, B.,
2018. Dual Function Lewis Acid Catalyzed Depolymerization of Industrial Corn
Stover Lignin into Stable Monomeric Phenols. ACS Sustain. Chem. Eng. 7(1), 13621371.

76659. Nguyen, J.D., Matsuura, B.S., Stephenson, C.R., 2014. A photochemical strategy for767 lignin degradation at room temperature. J. Am. Chem. Soc. 136(4), 1218-21.

76860. Opris, C., Cojocaru, B., Gheorghe, N., Tudorache, M., Coman, S.M., Parvulescu, V.I.,Duraki, B., Krumeich, F., van Bokhoven, J.A., 2017. Lignin Fragmentation onto

770 Multifunctional Fe₃O₄@Nb₂O₅@Co@Re Catalysts: The Role of the Composition and

771 Deposition Route of Rhenium. ACS Catal. 7(5), 3257-3267.

77261. Ouyang, X., Huang, X., Hendriks, B.M.S., Boot, M.D., Hensen, E.J.M., 2018. Coupling
organosolv fractionation and reductive depolymerization of woody biomass in a twostep catalytic process. Green Chem. 20(10), 2308-2319.

77562. Pan, J., Fu, J., Lu, X., 2015. Microwave-Assisted Oxidative Degradation of Lignin Model

Compounds with Metal Salts. Energy Fuels 29(7), 4503-4509.

Panyadee, R., Posoknistakul, P., Jonglertjunya, W., Kim-Lohsoontorn, P., Laosiripojana,
N., Matsagar, B.M., Wu, K.C.W., Sakdaronnarong, C., 2018. Sequential Fractionation
of Palm Empty Fruit Bunch and Microwave-Assisted Depolymerization of Lignin for
Producing Monophenolic Compounds. ACS Sustain. Chem. Eng. 6(12), 1689616906.

78264. Petridis, L., Smith, J.C., 2018. Molecular-level driving forces in lignocellulosic biomassdeconstruction for bioenergy. Nat. Rev. Chem. 2(11), 382-389.

78465. Prado, R., Erdocia, X., De Gregorio, G.F., Labidi, J., Welton, T., 2016. Willow Lignin
785 Oxidation and Depolymerization under Low Cost Ionic Liquid. ACS Sustain. Chem.
786 Eng. 4(10), 5277-5288.

78766. Pu, Y., Cao, S., Ragauskas, A.J., 2011. Application of quantitative 31P NMR in biomass
788 lignin and biofuel precursors characterization. Energy Env. Sci. 4(9), 3154.

78967. Qi, S.-C., Hayashi, J.-i., Kudo, S., Zhang, L., 2017. Catalytic hydrogenolysis of kraft
lignin to monomers at high yield in alkaline water. Green Chem. 19(11), 2636-2645.

79168. Qi, W., Su, D., 2014. Metal-Free Carbon Catalysts for Oxidative Dehydrogenation792 Reactions. ACS Catal. 4(9), 3212-3218.

79369. Qian, F., Zhu, X., Liu, Y., Shi, Q., Wu, L., Zhang, S., Chen, J., Ren, Z.J., 2018. Influences
of Temperature and Metal on Subcritical Hydrothermal Liquefaction of
Hyperaccumulator: Implications for the Recycling of Hazardous Hyperaccumulators.
Environ. Sci. Technol. 52(4), 2225-2234.

79770. Qu, S., Dang, Y., Song, C., Guo, J., Wang, Z.-X., 2015. Depolymerization of Oxidized
Lignin Catalyzed by Formic Acid Exploits an Unconventional Elimination
Mechanism Involving 3c–4e Bonding: A DFT Mechanistic Study. ACS Catal. 5(11),
6386-6396.

80171. Ragauskas, A.J., Beckham, G.T., Biddy, M.J., Chandra, R., Chen, F., Davis, M.F.,
Bovison, B.H., Dixon, R.A., Gilna, P., Keller, M., Langan, P., Naskar, A.K., Saddler,
J.N., Tschaplinski, T.J., Tuskan, G.A., Wyman, C.E., 2014. Lignin valorization:
improving lignin processing in the biorefinery. Science 344(6185), 1246843.

80572. Rahimi, A., Ulbrich, A., Coon, J.J., Stahl, S.S., 2014. Formic-acid-induced

depolymerization of oxidized lignin to aromatics. Nature 515(7526), 249-52.

80773. Rinaldi, R., Jastrzebski, R., Clough, M.T., Ralph, J., Kennema, M., Bruijnincx, P.C.A.,
Weckhuysen, B.M., 2016. Paving the Way for Lignin Valorisation: Recent Advances
in Bioengineering, Biorefining and Catalysis. Angew. Chem., Int. Ed. 55(29), 8164810 8215.

81174. Renders, T., Schutyser, W., Van den Bosch, S., Koelewijn, S.-F., Vangeel, T., Courtin,
C.M., Sels, B.F., 2016a. Influence of Acidic (H₃PO₄) and Alkaline (NaOH) Additives
on the Catalytic Reductive Fractionation of Lignocellulose. ACS Catal. 6(3), 20552066.

81575. Renders, T., Van den Bosch, S., Vangeel, T., Ennaert, T., Koelewijn, S.-F., Van den
Bossche, G., Courtin, C.M., Schutyser, W., Sels, B.F., 2016b. Synergetic Effects of
Alcohol/Water Mixing on the Catalytic Reductive Fractionation of Poplar Wood. ACS
Sustain. Chem. Eng. 4(12), 6894-6904.

81976. Schutyser, W., Kruger, J.S., Robinson, A.M., Katahira, R., Brandner, D.G., Cleveland,
N.S., Mittal, A., Peterson, D.J., Meilan, R., Román-Leshkov, Y., Beckham, G.T.,
2018a. Revisiting alkaline aerobic lignin oxidation. Green Chem. 20(16), 3828-3844.

82277. Schutyser, W., Renders, T., Van den Bosch, S., Koelewijn, S.F., Beckham, G.T., Sels,823 B.F., 2018b. Chemicals from lignin: an interplay of lignocellulose fractionation,

depolymerization, and upgrading. Chem. Soc. Rev. 47(3), 852-908.

82578. Sedai, B., Díaz-Urrutia, C., Baker, R.T., Wu, R., Silks, L.A.P., Hanson, S.K., 2011.
Comparison of Copper and Vanadium Homogeneous Catalysts for Aerobic Oxidation
of Lignin Models. ACS Catal. 1(7), 794-804.

82879. Shaheen, S.M., Niazi, N.K., Hassan, N.E.E., Bibi, I., Wang, H., Tsang, Daniel C.W., Ok,
Y.S., Bolan, N., Rinklebe, J., 2018. Wood-based biochar for the removal of potentially
toxic elements in water and wastewater: a critical review. Int. Mater. Rev. 64(4), 216247.

83280. Shao, L., Zhang, Q., You, T., Zhang, X., Xu, F., 2018. Microwave-assisted efficient
depolymerization of alkaline lignin in methanol/formic acid media. Bioresour.
834 Technol. 264, 238-243.

83581. Shao, Y., Xia, Q., Dong, L., Liu, X., Han, X., Parker, S.F., Cheng, Y., Daemen, L.L.,

Ramirez-Cuesta, A.J., Yang, S., Wang, Y., 2017. Selective production of arenes via
direct lignin upgrading over a niobium-based catalyst. Nat. Commun. 8, 16104.

83882. Shuai, L., Amiri, M.T., Questell-Santiago, Y.M., Heroguel, F., Li, Y., Kim, H., Meilan, R.,
Chapple, C., Ralph, J., Luterbacher, J.S., 2016. Formaldehyde stabilization facilitates
lignin monomer production during biomass depolymerization. Science 354(6310),
329-333.

84283. Shuai, L., Sitison, J., Sadula, S., Ding, J., Thies, M.C., Saha, B., 2018. Selective C–C
843 Bond Cleavage of Methylene-Linked Lignin Models and Kraft Lignin. ACS Catal.
844 8(7), 6507-6512.

84584. Shylesh, S., Gokhale, A.A., Ho, C.R., Bell, A.T., 2017. Novel Strategies for the
Production of Fuels, Lubricants, and Chemicals from Biomass. Acc. Chem. Res.
50(10), 2589-2597.

84885. Sikarwar, V.S., Zhao, M., Clough, P., Yao, J., Zhong, X., Memon, M.Z., Shah, N.,
Anthony, E.J., Fennell, P.S., 2016. An overview of advances in biomass gasification.
Energy Envi. Sci. 9(10), 2939-2977.

85186. Song, S., Zhang, J., Gozaydin, G., Yan, N., 2019. Production of Terephthalic Acid from852 Corn Stover Lignin. Angew. Chem. Int. Ed. 58(15), 4934-4937.

85387. Stiefel, S., Di Marino, D., Eggert, A., Kühnrich, I.R., Schmidt, M., Grande, P.M., Leitner,

W., Jupke, A., Wessling, M., 2017. Liquid/liquid extraction of biomass-derived lignin
from lignocellulosic pretreatments. Green Chem. 19(1), 93-97.

85688. Sun, Z., Fridrich, B., de Santi, A., Elangovan, S., Barta, K., 2018. Bright Side of Lignin
Bepolymerization: Toward New Platform Chemicals. Chem. Rev. 118(2), 614-678.

85889. Toledano, A., Serrano, L., Pineda, A., Romero, A.A., Luque, R., Labidi, J., 2014.
Microwave-assisted depolymerization of organosolv lignin via mild hydrogen-free

hydrogenolysis: Catalyst screening. Appl. Catal. B: Envi. 145, 43-55.

86190. Van den Bosch, S., Schutyser, W., Vanholme, R., Driessen, T., Koelewijn, S.F., Renders,

T., De Meester, B., Huijgen, W.J.J., Dehaen, W., Courtin, C.M., Lagrain, B., Boerjan,

863 W., Sels, B.F., 2015. Reductive lignocellulose fractionation into soluble lignin-derived

phenolic monomers and dimers and processable carbohydrate pulps. Energy Envi.

865 Sci. 8(6), 1748-1763.

86691. Vikrant, K., Kim, K.H., Ok, Y.S., Tsang, D.C.W., Tsang, Y.F., Giri, B.S., Singh, R.S.,
2018. Engineered/designer biochar for the removal of phosphate in water and
wastewater. Sci. Total Environ. 616-617, 1242-1260.

86992. Wang, M., Lu, J., Zhang, X., Li, L., Li, H., Luo, N., Wang, F., 2016. Two-Step, Catalytic
870 C–C Bond Oxidative Cleavage Process Converts Lignin Models and Extracts to
871 Aromatic Acids. ACS Catal. 6(9), 6086-6090.

- 87293. Wang, S., Li, Z., Bai, X., Yi, W., Fu, P., 2019a. Catalytic pyrolysis of lignin in a cascade
 dual-catalyst system of modified red mud and HZSM-5 for aromatic hydrocarbon
 production. Bioresour. Technol. 278, 66-72.
- 87594. Wang, W., Wang, M., Huang, J., Zhao, X., Su, Y., Wang, Y., Li, X., 2019b. Formateassisted analytical pyrolysis of kraft lignin to phenols. Bioresour. Technol. 278, 464467.
- 87895. Wu, X., Fan, X., Xie, S., Lin, J., Cheng, J., Zhang, Q., Chen, L., Wang, Y., 2018. Solar
 energy-driven lignin-first approach to full utilization of lignocellulosic biomass under
 mild conditions. Nature Catal. 1(10), 772-780.
- 88196. Wu, Ζ., Zhao, Х., Zhang, J., Li, X., Zhang, Y., Wang, F., 2019. 882 Ethanol/1,4-dioxane/formic acid as synergistic solvents for the conversion of lignin 883 into high-value added phenolic monomers. Bioresour. Technol. 278, 187-194.

88497. Xiao, L.-P., Wang, S., Li, H., Li, Z., Shi, Z.-J., Xiao, L., Sun, R.-C., Fang, Y., Song, G.,

- 2017. Catalytic Hydrogenolysis of Lignins into Phenolic Compounds over Carbon
 Nanotube Supported Molybdenum Oxide. ACS Catal. 7(11), 7535-7542.
- 88798. Xiong, X., Yu, I.K.M., Cao, L., Tsang, D.C.W., Zhang, S., Ok, Y.S., 2017. A review of
 biochar-based catalysts for chemical synthesis, biofuel production, and pollution
 control. Bioresour. Technol. 246, 254-270.
- 89099. Xiong, X.; Yu, I.K.M.; Tsang, D.C.W.; Bolan, N.S.; Ok, Y.S.; Igalavithana, A.D.;
 Kirkham, M.B.; Kim, K.H.; Vikrant, K., 2019. Value-added chemicals from food
 supply chain wastes: A critical review. Chem. Eng. J., 375, 121983.
- 893100. Xu, R., Zhang, K., Liu, P., Han, H., Zhao, S., Kakade, A., Khan, A., Du, D., Li, X.,
 2018. Lignin depolymerization and utilization by bacteria. Bioresour. Technol. 269,
 557-566.

- 896101. Yan, N., Zhao, C., Dyson, P.J., Wang, C., Liu, L.-t., Kou, Y., 2008. Selective
 B97 Degradation of Wood Lignin over Noble-Metal Catalysts in a Two-Step Process.
 898 ChemSusChem 1(7), 626-629.
- 899102. Yang, S., Yuan, T.-Q., Sun, R.-C., 2016. Structural Elucidation of Whole Lignin in Cell
 Walls of Triploid of Populus tomentosa Carr. ACS Sustain. Chem. Eng. 4(3), 10061015.
- 902103. Yang, X., Igalavithana, A.D., Oh, S.E., Nam, H., Zhang, M., Wang, C.H., Kwon, E.E.,
 903 Tsang, D.C.W., Ok, Y.S., 2018. Characterization of bioenergy biochar and its
 904 utilization for metal/metalloid immobilization in contaminated soil. Sci. Total
 905 Environ., 640-641, 704-713.
- 906104. Yang, X., Yu, I.K.M., Cho, D.W., Chen, S.S., Tsang, D.C.W., Shang, J., Yip, A.C.K.,
 907 Wang, L., Ok, Y.S., 2019. Tin-functionalized wood biochar as a sustainable solid
 908 catalyst for glucose isomerization in biorefinery. ACS Sustain. Chem. Eng., 7, 4851909 4860.
- 910105. Yao, S.G., Mobley, J.K., Ralph, J., Crocker, M., Parkin, S., Selegue, J.P., Meier, M.S.,
 2018. Mechanochemical Treatment Facilitates Two-Step Oxidative Depolymerization
 of Kraft Lignin. ACS Sustain. Chem. Eng. 6(5), 5990-5998.

913106. Yong, T.L.-K., Matsumura, Y., 2013. Kinetic Analysis of Lignin Hydrothermal
914 Conversion in Sub- and Supercritical Water. Ind. & Eng. Chem. Res. 52(16), 5626915 5639.

916107. Yoo, C.G., Dumitrache, A., Muchero, W., Natzke, J., Akinosho, H., Li, M., Sykes, R.W.,

917 Brown, S.D., Davison, B., Tuskan, G.A., Pu, Y., Ragauskas, A.J., 2017a. Significance
918 of Lignin S/G Ratio in Biomass Recalcitrance of Populus trichocarpa Variants for
919 Bioethanol Production. ACS Sustain. Chem. Eng. 6(2), 2162-2168.

920108. Yoo, C.G., Li, M., Meng, X., Pu, Y., Ragauskas, A.J., 2017b. Effects of organosolv and
921 ammonia pretreatments on lignin properties and its inhibition for enzymatic
922 hydrolysis. Green Chem. 19(8), 2006-2016.

923109. Yoo, J.C., Beiyuan, J., Wang, L., Tsang, D.C.W., Baek, K., Bolan, N.S., Ok, Y.S., Li, X.D., 2018. A combination of ferric nitrate/EDDS-enhanced washing and sludge-

925 derived biochar stabilization of metal-contaminated soils. Sci. Total Environ., 616-926 617, 572-582.

927110. You, S., Ok, Y.S., Chen, S.S., Tsang, D.C.W., Kwon, E.E., Lee, J., Wang, C.H., 2017. A
928 critical review on sustainable biochar system through gasification: Energy and
929 environmental applications. Bioresour. Technol. 246, 242-253.

930111. Younas, R., Hao, S., Zhang, L., Zhang, S., 2017. Hydrothermal liquefaction of rice strawwith NiO nanocatalyst for bio-oil production. Renew. Energy 113, 532-545.

932112. Yu, I.K.M., Tsang, D.C.W., 2017a. Conversion of biomass to hydroxymethylfurfural: A
review of catalytic systems and underlying mechanisms. Bioresour. Technol. 238,
716-732.

935113. Yu, I.K.M., Tsang, D.C.W., Yip, A.C.K., Chen, S.S., Wang, L., Ok, Y.S., Poon, C.S.,
2017b. Catalytic valorization of food waste into hydroxymethylfurfural (HMF):
Controlling relative kinetics for high productivity. Bioresour. Technol. 237, 222-230.

938114. Yu, I.K.M., Tsang, D.C.W., Yip, A.C.K., Hunt, A.J., Sherwood, J., Shang, J., Song, H.,

939 Ok, Y.S., Poon, C.S., 2018. Propylene carbonate and γ-valerolactone as green solvents
940 enhance Sn(IV)-catalysed hydroxymethylfurfural (HMF) production from bread
941 waste. Green Chem. 20, 2064-2074.

942115. Yu, I.K.M., Xiong, X., Tsang, D.C.W., Wang, L., Hunt, A.J., Song, H., Shang, J., Ok,

943 Y.S., Poon, C.S., 2019. Aluminium-biochar composite as a sustainable heterogeneous944 catalyst for glucose isomerization in biorefinery. Green Chem. 21, 1267-1281.

945116. Zaheer, M., Kempe, R., 2015. Catalytic Hydrogenolysis of Aryl Ethers: A Key Step in946 Lignin Valorization to Valuable Chemicals. ACS Catal. 5(3), 1675-1684.

947117. Zhai, Y., Li, C., Xu, G., Ma, Y., Liu, X., Zhang, Y., 2017. Depolymerization of lignin via
a non-precious Ni–Fe alloy catalyst supported on activated carbon. Green Chem.
19(8), 1895-1903.

950118. Zhang, Z., Harrison, M.D., Rackemann, D.W., Doherty, W.O.S., O'Hara, I.M., 2016.

951 Organosolv pretreatment of plant biomass for enhanced enzymatic saccharification.952 Green Chem. 18(2), 360-381.

953119. Zhang, X., Rajagopalan, K., Lei, H., Ruan, R., Sharma, B.K., 2017. An overview of anovel concept in biomass pyrolysis: microwave irradiation. Sustain. Energy Fuels

955 1(8), 1664-1699.

- 956120. Zhang, B., Yang, D., Wang, H., Qian, Y., Huang, J., Yu, L., Qiu, X., 2018a. Activation of
 957 Enzymatic Hydrolysis Lignin by NaOH/Urea Aqueous Solution for Enhancing Its
 958 Sulfomethylation Reactivity. ACS Sustain. Chem. Eng. 7(1), 1120-1128.
- 959121. Zhang, P., Guo, Y.-J., Chen, J., Zhao, Y.-R., Chang, J., Junge, H., Beller, M., Li, Y.,
 2018b. Streamlined hydrogen production from biomass. Nature Catalysis 1(5), 332338.
- 962122. Zhang, C., Jia, C., Cao, Y., Yao, Y., Xie, S., Zhang, S., Lin, H., 2019. Water-assisted
 selective hydrodeoxygenation of phenol to benzene over the Ru composite catalyst in
 the biphasic process. Green Chem. 21(7), 1668-1679.
- 965123. Zhang, K., Li, H., Xiao, L.P., Wang, B., Sun, R.C., Song, G. 2019. Sequential utilization
 of bamboo biomass through reductive catalytic fractionation of lignin. Bioresour.
 967 Technol., 285, 121335.
- 968124. Zhou, L., Budarin, V., Fan, J., Sloan, R., Macquarrie, D. 2017. Efficient Method of
 P69 Lignin Isolation Using Microwave-Assisted Acidolysis and Characterization of the
 P70 Residual Lignin. ACS Sustainable Chem. Eng. 5(5), 3768-3774.
- 971125. Zhou, M., Sharma, B.K., Liu, P., Xia, H., Xu, J., Jiang, J.-C., 2018. Microwave Assisted
- 972 Depolymerization of Alkaline Lignin over Hydrotalcite-Based CuNiAl Mixed Oxides.
- 973 ACS Sustain. Chem. Eng. 6(9), 11519-11528.

Fractionation	Depolymerization	Upgrading
Various fractionations	Aid/Base-catalyzed depolymerization	Aromatic chemicals;
result in modification of	Oxidative/Reductive depolymerization	Bio-oils; Bio-char;
native lignin.	Thermocatalytic / Photocatalytic / Mechanochemical_depolymerization	Bio-gases (CO/H ₂ /CH ₄)
Challenges: Identification	meenancenemical acpolymenzation	Challenges: Selective
of lignin structure (building	Challenges: High yield and selective	functionalization and
blocks and linkage types)	production of aromatic chemicals from	de-functionalization of
with various process.	waste lignin.	aromatic products.
	1	
Lignin valorization: Advances	in pristine lignin structure analysis and cor	version technologies can
facilitate recovery and sub	sequent utilization of lignin towards bio-ba	sed chemicals & fuels

Fig. 1. Overview of lignin valorization.





Fig. 2. An overview of depolymerization methodologies of lignin.

T	Caller i		X7'-1.1. 7.4	
Entry	Substrates	Condition: Solvent;	Yield: Monomers	Kei.
		ume; temperature and	and D10-011S	
1	Kraft lignin	Ethanol/1 4-dioxano/	Monomore 25 4%	(Wu ot al. 2019)
T	Kian nghin	formic acid:	Rio oils 55%	(Wu et al., 201 <i>5)</i>
			DI0-0115 5570	
C	Dulping lignin	Sou C, 2 II.	Dio oils E4.0/	(Dong et al. 2014)
2	Pulping nginin	$160 ^{\circ}C ^{2}O ^{\circ}min$	DI0-0115 54 70	(Dolig et al., 2014)
2	Organocoly	Totralin:	Bio oils 17 %	(Tolodano et al
J	lignin	160 °C 20 min:	DI0-0113 17 70	(101euano et al.,
	IIgiiiii	N:100/AISDA cotolyct		2014)
4	Organacalty	NIIU%AISDA Caldiyst.	Monomore 29 ret 0/	(Nandiwala at al
4	lignin		Witholiners 20 wt 70	
	IIgIIII	250° C, 511, N ₂ ; 7r KIT C costalyat		2016)
-	Organacalty		Die eile 52 67 w#0/	(Dahral at al
J		$\Pi_2 O;$	DI0-0115 52-07 W1%	
	ngnin			2018a)
C	Allealing	Cs_2CO_3 catalyst.		(Chaudhaura 9
0	Alkaline	Ethanol/ H_2O ;	BI0-0115 51 W1%	(Chaudhary &
	ngnin	250 °C, 1 II;		Dhepe, 2017)
7	Allealing	NdX (SI/AI = 1.2).	Die eile $24 - \pm 0/$	(Chaudhaura 9
/	Alkaline	Ethanol/ H_2O ;	BI0-0115 34 W1%	(Chaudhary &
	ngnin	250 °C, 1 II;		Dhepe, 2017)
0	Dulaine lieuin		Manager 20 and 9/	(11
ð	Pulping lignin	Ethanol; $240 \times C 4 = N$.	Monomers 36 wt %	(Huang et al., 2015)
		540 C 4 II, N_2 ;		
0	Doplar	NaOH colution	Monomore 20 wt9/	(Schutzcor of al
9	Popiar	175 °C 20 min O / Ho	Monomers 30 wt%	(Schutyser et al.,
10		1/5 C, 50 IIIII, O ₂ / He.	Dia cile 72.0 $ \pm 10^{\prime} $	$\frac{2010d}{(\text{Shap} et al. 2010)}$
10	Alkaline	Methanol/formic aciu;	BI0-0115 /2.0 Wt%	(Shao et al., 2018)
11	ngiini	100 C, 30 IIIII.	Monomore 9 - +0/	(Deng et al. 2015)
11			Monomers 8 wt%	(Deng et al., 2015)
	ngnin	$185^{-1}C$, 24 II, O_2 ;		
10	Organacala		Manamara 17.0 a d	(III): dow of al
12	Urganosolv	$H_2U;$	Monomers 17.8 wt	
	IIgiiiii	200° C, 4 II, O_2 ,	70	2018)
10	Domboo	CofeO mixed oxides.	Manager 22 20/	(7hang at al. 2010)
13	Bamboo		Monomers 32.2%	(Zhang et al., 2019)
	DIOMASS	$260^{-1}C, 4 \Pi, H_2;$		
14	Allealing	FUC Catalyst.	Monomore E2 at 0/	(Koranzi et al
14	AIKaille	Emidiion; $240 \circ C$ $A = 11 \cdot 12$	wonomers 53 wt %	(NOTAIIYI et al.,
	ngiiii	C_{11} U, 4 II, Π_2 ;		2017)
		$Cu+IN1_2P/S1O_2$ catalyst.		

Table 1. Summary of various catalytic depolymerization of lignin/biomass

15	Enzymatic	Methanol;	Monomers 47 wt%	(Xiao et al., 2017)
	lignin	260 °C, 4 h, H ₂ ;		
		MoOx/CNT catalyst.		
16	Corn stover	Methanol;	Monomers 28 wt%	(Anderson et al.,
		200 °C, 24 h;		2016)
		Ni/C catalysts.		
17	Corn Stover	Toluene;	De-methoxylation	(Song et al., 2019)
		320 °C, 2 h, H ₂ ;	monomers 51 wt%	
		MoO _x /AC catalyst		
18	Kraft lignin	HMinBF ₄ ;	Acetic acid 87 wt%	(Mei et al., 2017)
		120 °C, 12 h, CO;		
		RhCl ₃ catalyst.		

Techniques	Advantages	Disadvantages	Ref.
Microwave	Rapid heating rate;	Difficulties of scaling up;	(Shao et al., 2018;
reaction	Energy efficiency;	Uncontrolled heating	Zhang et al.,
	Low temperature and	process;	2017)
	pressure.	Energy consumption.	
Hydrothermal	Mild reaction condition;	Relatively inefficient	(Lu et al., 2018;
liquefaction	Variety of biomass;	convection;	Yong &
	Large-scale production.	Water consumption.	Matsumura, 2013)
Pyrolysis and	Flexibility of feedstock;	Harsh temperature;	(Duan et al.,
Gasification	Fast rates;	Non-selective conversion.	2017; Fan et al.,
	High yield.		2018)
Photocatalytic	Definite mild condition;	Design of efficient catalyst;	(Luo et al., 2016)
reaction	Renewable source of energy.	Technological barriers.	
Mechanochemi	Fast rates;	Non-selective	(Yao et al., 2018;
cal reaction	Solvent-free;	depolymerization;	Dabral et al.,
	Flexibility of feedstock.	Process complexity;	2018b)
		High energy consumption.	

984Table. 2 Advantages and disadvantages of various techniques for lignin conversion