

1 **Antibiotics in the agricultural soils from the Yangtze River Delta**

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19 **ABSTRACT**

20 This study focused on the occurrence and spatial distribution of 13 common antibiotics in the  
21 agricultural soils of the Yangtze River Delta (YRD), China. Antibiotics were detected in all the  
22 241 soil samples (i.e., 100% detection rate) with the total concentrations ranging from 4.55 to  
23 2,010 ng/g dry weight. The concentrations of three antibiotic classes decreased in the order:  
24 quinolones (mean 48.8 ng/g) > tetracyclines (mean 34.9 ng/g) > sulfonamides (mean 2.35 ng/g).  
25 Ciprofloxacin was the prevalent compound with a mean concentration of 27.7 ng/g, followed by  
26 oxytetracycline (mean of 18.9 ng/g). A distinct spatial distribution was observed, where high  
27 concentrations of antibiotics were detected in the sites adjacent to the livestock and poultry farms.  
28 The potential sources of antibiotics in the agricultural soils were the application of manure and  
29 wastewater irrigation in this region. Risk assessment for single antibiotic compound indicated that  
30 tetracyclines and quinolones could pose a potential risk, in which doxycycline and ciprofloxacin  
31 had the most severe ecological effect in the agricultural soils. Antibiotic resistance genes (ARGs),  
32 such as tetA, sulI, and qnrS, were detected in 15 analyzed soil samples, and sulI showed significant  
33 correlations with quinolones, tetracyclines, copper, and zinc. Further studies on the distribution of  
34 other ARGs in agricultural soil at a region-scale are needed for the risk management of extensively  
35 used antibiotics and major ARGs.

36

37 **INTRODUCTION**

38 Antibiotics have become the most popular antimicrobial drug for human disease control and  
39 livestock growth promotion for a long time. China is one of the largest producers and consumers  
40 of anti-biotics in the world. The total usage of antibiotics was 92,700 tons in 2013, and an

41 estimated 54,000 tons of the antibiotics was discharged after human and animals use (Zhang et al.,  
42 2015). A large number of these antibiotics cannot be metabolized in vivo, 30 - 90% of which will  
43 be excreted into the environment (Hu et al., 2010; Zhao et al., 2010). Due to the use of organic  
44 manure/sludge and irrigation of wastewater or reclaimed water in the agricultural land, a  
45 significant amount of antibiotics, such as tetracyclines and quinolones, are found in soils (Wang  
46 et al., 2014a, 2014b; Hou et al., 2015).

47 Many antibiotics are biologically active in the environment (Zhou et al., 2006, 2013; Xu et al.,  
48 2013), which may pose a pressure on antibiotic resistance genes (ARGs) and bacterial  
49 communities (Ding and He, 2010; Tello et al., 2012; Xiong et al., 2015). The occurrence of ARGs  
50 in sediments, soil, and water has been observed in different countries (Pruden et al., 2006; Knapp  
51 et al., 2010; Chen et al., 2016a), which may be enriched by antibiotic residues (Cheng et al., 2016;  
52 Yang et al., 2017) and transferred into different environmental media (Chen et al., 2016a,b; Xie et  
53 al., 2016). Therefore, antibiotic may cause significant impacts on the environment and human  
54 (Tello et al., 2012).

55 It has been reported that soil is an important reservoir for antibiotics. For example, the levels of  
56 several antibiotics in soils in farmland around feedlots in Shanghai were 3.27e33.4 mg/g (Hu et  
57 al., 2010). The detected concentrations of antibiotics in organic farms ranged from 0.1 to 2,683  
58 ng/g in soils (Xiang et al., 2016). Antibiotics with high concentration were detected in vegetable  
59 farmlands adjacent to livestock farms, and the detected concentration of quinolones can reach up  
60 to 1,540 ng/g (Li et al., 2011). This indicated that livestock manure is an important source of  
61 antibiotic residues in soil. After the application of manure/sludge contaminated with antibiotics to  
62 agricultural soil, the residues of antibiotics in soil can be accumulated by vegetables that are  
63 consumed by human (Hu et al., 2010).

64 The Yangtze River Delta (YRD) is a developed and populated region in China. The YRD region  
65 include Shanghai Municipality, Zhejiang province and Jiangsu province. As a result, the increasing  
66 need for agricultural produce from urban population led to booming agriculture in the region (Sun  
67 et al., 2016b). Our previous studies showed that organochlorine pesticides, phthalate esters,  
68 polybrominated diphenyl ethers (Sun et al., 2016b), and poly- chlorinated biphenyls (Sun et al.,  
69 2016a) were widely detected, which may pose ecological risks in the YRD region. Yet, little in-  
70 formation is known on the distribution of commonly-used antibiotics in agricultural soils of the  
71 YRD region. It is important to evaluate the regional-scale distribution and the potential ecological  
72 risks of antibiotics to facilitate sustainable economic development for this rapidly developing  
73 region.

74 This study characterizes the concentrations and distribution patterns of tetracycline, sulfonamide  
75 and quinolone residues in agricultural soils from the YRD region of China and their potential  
76 ecological risks, as well as typical ARGs in a number of soil samples. The regional-scale  
77 evaluation of multiple antibiotics pollution in agricultural soils will provide useful information for  
78 the management of antibiotics contamination in China.

79

## 80 **MATERIALS AND METHODS**

### 81 **Chemicals and reagents**

82 The targeted antibiotics in this study were chosen according to their application in livestock  
83 industry in China. There are three classes of antibiotics, i.e., tetracyclines (TCs), quinolones (QNs)  
84 and sulfonamides (SAs). Three TCs included tetracycline (TC), doxycy- cline (DC), and  
85 oxytetracycline (OTC). Four QNs included ofloxacin (OFL), ciprofloxacin (CIP), norfloxacin

86 (NOR), and enrofloxacin (ENR). Six SAs included sulfadimethoxine (SDM), sulfamethazine  
87 (SMZ), sulfamethizole (SMTZ), sulfamerazine (SMR), sulfamethoxazole (SMX), and sulfadiazine  
88 (SDZ). Simeton was used as the internal standard. The standards were obtained from J & K  
89 Chemical Ltd., USA. Acetonitrile and methanol (HPLC grade) were obtained from Sigma-Aldrich.  
90 Oasis HLB Cartridge used for purification were purchased from Waters. Deionized water (18.2  
91 MΩ) from a Milli-Q system was used.

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### 93 **Sample collection**

94 The YRD region is located in east China. In June 2014, 241 agricultural soils were collected in  
95 this region. The studied region covered approximately 45,800 km<sup>2</sup>. The detailed information about  
96 sample collection were presented in our previous report (Sun et al., 2016b). The types of land-use  
97 of the sampling sites included paddy fields, vegetable fields, forests, uncultivated lands, and other  
98 agricultural uses.

99

### 100 **Sample extraction and clean-up**

101 The extraction and cleanup procedures for antibiotics were conducted according to the methods  
102 reported previously (Uslu. et al., 2007), with small modifications. Briefly, all the soil samples were  
103 purified using solid phase extraction (SPE) method after freeze drying, homogenizing and sieving  
104 through a 75 mesh sieve.

105 Two grams of soil were extracted by using acetonitrile (3 mL) and potassium phosphate buffer (3  
106 mL) (pH = 4) containing 0.2 g of EDTA in a 15-mL polypropylene centrifuge tube. After being

107 vortexed for 1 min, the sample was treated ultrasonically for 20 min, followed by centrifugation  
108 at 2600 rpm for 10 min. Afterwards, the supernatants were moved carefully to a new tube. This  
109 extraction procedure was repeated three more times with additions of 5, 5 and 4 mL of reagents.  
110 The extracts were combined and transferred to new glass bottles. Then, these extracts were  
111 concentrated with a rotary evaporator, and diluted to 500 mL to lower the concentration of organic  
112 solvents to below 5%.

113 The Oasis HLB Cartridge was used for the purification of each soil sample. The cartridges were  
114 preconditioned sequentially with 6 mL of methanol and 6 mL of deionized water. Then, the dilute  
115 supernatants were loaded on the cartridges at a rate of about 5 mL/min. The cartridges were then  
116 vacuum-dried after being rinsed with 12 mL of deionized water. The elution on the cartridges were  
117 performed with 0.1% acidified methanol (9 mL) at a rate of 1.0 mL/min. The analytes were then  
118 concentrated to near dryness under a gentle flow of nitrogen gas. Simeton was added and methanol  
119 was used to dissolve the final extract to 1 mL. The final extract was filtered through syringe filters  
120 (0.22  $\mu\text{m}$ ) before further instrumental analysis.

121

## 122 **Instrumental analysis**

123 The determination of antibiotics in this study was performed by using a liquid chromatography-  
124 tandem mass spectrometry system (Agilent 1260 - 6460), according to a previous study (Wu et al.,  
125 2014) with some modifications. A C18 column (5  $\mu\text{m}$ , 2.1 x 150 mm) was used for the  
126 chromatographic separation. The mobile phase was methanoleacetonitrile (1:1, v/v) and 0.3%  
127 formic acid/water (containing 0.1% ammonium formate, v/v). The column temperature was 40°C,  
128 and the injection volume was 20  $\mu\text{L}$ . The detection was conducted with ESI source in the mode of

129 multiple- reaction monitoring (MRM). The detailed information on instrumental parameters is  
130 listed in Supporting Information (Table S1).

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### 132 **DNA extraction and real-time qPCR**

133 The current study analyzed three common ARGs (tetA, sulI, and qnrS) in the selected 15 sampling  
134 sites from the northeast of the YRD region, where high levels of antibiotics were found. The  
135 methods of DNA extraction and real-time qPCR were adopted from the previous literature (Chen  
136 and Zhang, 2013). Briefly, DNA extraction from soil samples was conducted with the PowerSoil  
137 DNA Isolation Kit following the instructions. Three ARGs (tetA, sulI, and qnrS) were analyzed  
138 by PCR and agarose electrophoresis, quantified by real-time qPCR. The reaction system of real-  
139 time qPCR comprised 7.5  $\mu$ L of SYBR Premix Ex Taq™ (TaKaRa), 0.3  $\mu$ L of ROX reference dye  
140 for correction, 0.3  $\mu$ L of each 10  $\mu$ M primer, 2  $\mu$ L of template DNA and 4.6  $\mu$ L of ddH<sub>2</sub>O. The  
141 temperature programming of qPCR contained a hold for 30 s at 95°C, followed by 40 cycles of 5  
142 s at 95°C and 30s at 55°C for annealing, then 30 s at 72°C for extension and a fluorescence  
143 acquisition step at 72°C, a final melt curve stage conducted from 60 to 95°C. The information of  
144 the qPCR primers and standard curves are shown in Table S2.

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### 146 **Quality control and quality assurance**

147 A spiked blank, a procedural blank, and a duplicate of soil were processed in parallel in each batch  
148 of ten soils. The relative standard deviation (RSD) in the measured concentrations of duplicate  
149 samples was less than 15%. The average recoveries of 13 target com- pounds in soil samples  
150 ranged from 73 to 108%. The coefficients of determination of the working calibration curve

151 (0.1e100 ng/g) were all >0.999. The limit of detection (LOD) for antibiotics were calculated and  
152 fell in the range of 0.10e0.80 ng/g (Table S3). Statistical analysis with Microsoft European  
153 Commission, 2003 and SPSS (IBM SPSS Statistics 20) was conducted. Spatial distributions of the  
154 antibiotics were simulated by using universal Kriging in ArcGIS 10.2 (ESRI, Redlands, CA, USA).  
155 The measured concentra- tions of antibiotics were interpolated to the whole study area using  
156 ordinary kriging with normal score transformation (Deutsch and Journel, 1998), which assumed  
157 the normal distributions. Back- transformation was applied to the interpolated results for obtaining  
158 the spatial maps of the antibiotics concentrations.

159

## 160 **RESULTS AND DISCUSSION**

### 161 **Concentrations and profiles of antibiotics**

162 The concentrations of the total antibiotics in agricultural soils were in the range from 4.55 to 2,010  
163 ng/g, with a mean value of 86.1 ng/g (n = 241), indicating widely-spread antibiotic contamination  
164 in the agricultural soils of the region. The concentrations of 13 antibiotics in 241 agricultural soil  
165 samples of the YRD region are summarized in Table 1. Three antibiotic classes, including  
166 quinolones (OFL, ENR, CIP and NOR), tetracyclines (DC, OTC and TC), and sulfonamides (SDZ,  
167 SMZ, SMTZ, SDM, SMX and SMR) were detected, of which the detection rates ranged from 0.8%  
168 to 95% across the study area. The concentrations of total quinolones ( $\sum$  QNs), total tetracyclines  
169 ( $\sum$  TCs) and total sulfonamides ( $\sum$  SAs) ranged from <LOD to 1410 ng/g, from <LOD to 809 ng/g,  
170 and from <LOD to 111 ng/g, respectively. The mean values of three antibiotic classes decreased  
171 in the following order: QNs (48.8 ng/g) > TCs (34.9 ng/ g) > SAs (2.35 ng/g) in the soils.



172 Significant differences in the concentrations of three classes of antibiotics were observed in the  
173 YRD agricultural soils. The TCs and QNs were prevalent antibiotics with higher frequencies (99.0%  
174 and 99.6%, respectively) than SAs (66.8%). This may be caused by the chemical properties of  
175 diverse antibiotic compounds. Compared with most SAs, TCs and QNs can be more strongly  
176 absorbed by soil particles in the absence of biodegradation (Moreno-Bondi et al., 2009), and as a  
177 result they are more persistent in the environment (Hu et al., 2010).

178 The detection frequencies ranged from 72% to 83% for the four kinds of QNs. The CIP was the  
179 dominant compound in the class of QNs, followed by NOR, OFL and ENR. The mean  
180 concentration of CIP, NOR, OFL and ENR was 27.7 ng/g 11.2 ng/g, 5.86 ng/g and 3.98 ng/g,  
181 respectively. The maximum concentration levels of four QNs were 1030 ng/g for CIP, 505 ng/g  
182 for NOR, 498 ng/g for OFL, and 237 ng/g for ENR. Several previous studies reported the  
183 occurrence of QNs in agricultural soils. The level of QNs in this study was higher than the  
184 concentration detected in soils from organic vegetable farms in Guangdong, in which the average  
185 concentration of QNs was 14.0 ng/g (Wu et al., 2014). The mean concentration of QNs in the YRD  
186 region was lower than that from some other regions of China. For example, In the Pearl River  
187 Delta region in south China, the levels of four QNs in soils collected from vegetable farmlands fell  
188 in the range of 27.8 - 1530 ng/g (Li et al., 2011). The detected levels of QNs ranged from <LOD  
189 to 290 ng/g and from <LOD to 649 ng/g in Nanjing and Beijing greenhouse soils, respectively  
190 (Fang et al.,2015; Li et al., 2015).

191 The detection frequencies ranged from 37% to 95% for TCs. The mean values of individual TCs  
192 decreased in the following order: OTC (18.9 ng/g) > DC (13.9 ng/g) > TC (2.07 ng/g). The  
193 maximum concentrations were 530 ng/g for OTC, 105 ng/g for DC, and 197 ng/g for TC. It was  
194 reported that the mean values of TCs, including DC, TC and OTC, ranged from 2.38 to 21.9 ng/g

195 in agricultural soils of organic farms in south China (Xiang et al., 2016). Besides, in Fuyang,  
196 Zhejiang province, the mean concentration of TCs was 23 ng/g, with OTC as the dominant  
197 compound in different soil samples (Wu et al.,

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200 2013). In this research, the mean value of TCs was higher than these reported values in agricultural  
201 soils. As for SAs, the detection rate ranged from 0.8% to 44%, and six SAs were detected at very  
202 low levels (<LOD to 21.5 ng/g), which was consistent with the survey on the agricultural soils of  
203 Beijing with the mean concentration of 1.1 ng/g for SAs (Li et al., 2015). However, the detected  
204 levels of SDZ, SMX and SMZ in the current investigation were much lower than the concentrations  
205 in agricultural soils nearby feedlots (5.85 - 33.37  $\mu\text{g/g}$ ) (Ji et al., 2012). In our previous studies,  
206 various organic pollutants in these soils were detected (Sun et al., 2016a, b). There were significant  
207 differences between the levels of total antibiotics and phthalate esters (PAEs) ( $p < 0.001$ ),  
208 polybrominated diphenyl ethers (PBDEs) ( $p < 0.001$ ), and polychlorinated biphenyls (PCBs) ( $p <$   
209  $0.001$ ). There was no significant difference between the concentrations of antibiotics and  
210 organochlorine pesticides (OCPs) ( $p = 0.211$ ). These results reflect the extensive contamination of  
211 antibiotics and historical agricultural use of OCPs in this area. Significantly positive correlations  
212 were found between the levels of antibiotic and PCBs ( $r = 0.207$ ,  $p < 0.05$ ). The combined organic  
213 pollution in soil was widely detected, however, antibiotics used in China might originate from  
214 different sources.

215 The Veterinary International Committee on Harmonization sets the trigger value (100  $\mu\text{g/kg}$ ) that  
216 can cause ecotoxicity effects of soil antibiotics on a range of organisms (Huang et al., 2013). In

217 this study, it should be noted that the concentrations of 15.4%  $\Sigma$  antibiotics and 3.7% of individual  
218 antibiotics in soil samples exceeded the trigger value. The proportion of sites in which the  
219 concentrations of antibiotics exceeded the trigger value was 0.4%, 0.4%, 1.2%, 0.4%, 0.4%, 1.2%,  
220 1.7%, and 0.8% for SMTZ, DC, OTC, TC, OFL, ENR, CIP, and NOR, respectively.

221

## 222 **Spatial distribution and potential sources**

223 The spatial distribution pattern of three classes of antibiotics and total antibiotics are shown in Fig.  
224 1. Antibiotics are used for disease control and growth promotion for livestock and fishing farms  
225 (Wu et al., 2014; Liu et al., 2017), and they can be continuously excreted into the environment.  
226 Distinct spatial distribution was observed that the total concentrations of antibiotics were much  
227 higher in Jiangsu and Shanghai than in Zhejiang (Fig. 1A). This indicated the different  
228 contamination status of antibiotics in agricultural soil of different areas. The soil antibiotic  
229 concentrations were strongly influenced by proximity to the sources. The levels of QNs in  
230 agricultural soils were higher in Shanghai Municipality and Jiangsu province than in Zhejiang  
231 province (Fig. 1B). As for TCs, the levels in soil were higher in the border of Shanghai and Jiangsu  
232 (Fig. 1C). High levels of both TCs and QNs were detected in north- east of the YRD region,  
233 including Taicang, Jiading, Qingpu and Baoshan, where the livestock and fishing farming industry  
234 were well-developed (Hu et al., 2010; Li et al., 2015). Thus, the high levels of antibiotics might be  
235 originated from the extensive use of organic fertilizer and wastewater containing animal excreta  
236 in agricultural soils. There was a negative correlation between antibiotics and population density  
237 in the YRD region ( $r = 0.187$ ,  $p < 0.01$ ). This indicated the consumption of antibiotics by human  
238 beings may not be the main source of antibiotic contamination in agricultural soils. On the contrary,

239 the excessive use of antibiotics in the livestock and fishing farming industry in rural areas may  
240 contribute to the widespread contamination of antibiotics in these soils.

241 Moreover, the exacerbation of agricultural land use had intensified the application of fertilizers as  
242 well as the wastewater and reclaimed water irrigation, which could result in severe antibiotic  
243 contamination. It has been reported that there were high concentrations of antibiotics in wastewater  
244 and reclaimed water (Wang et al., 2014a; Wu et al., 2015; Ostman et al., 2017). In the coastal water  
245 of the South Yellow Sea in China, 25 antibiotics were widely present in the region with the total  
246 concentration up to 1350 ng/L (Du et al., 2017). In the YRD region, the SAs concentrations in  
247 soils were higher in Jiangsu province than in Shanghai and Zhejiang, which might result from farm  
248 irrigation with water containing SAs (Luo et al., 2011). In aquatic environments of Indochina, the  
249 concentrations of SMX and SMZ ranged from <LOD to 6.06  $\mu\text{g/g}$  (Suzuki and Hoa, 2012).

250 However, the levels of SAs in all the soils of the studied region were relatively low (Fig. 1D). The  
251 distribution pattern of a total of 13 antibiotics was consistent with that of QNs (Fig. 1A). The soil  
252 physicochemical properties, including total organic carbon (TOC), total phosphorous (TP), total  
253 nitrogen (TN), soil moisture content, and pH (Table S4) were reported in our previous studies (Sun  
254 et al., 2016a, b). However, no significant correlations were found between the total antibiotics and  
255 soil physicochemical properties ( $0.09 < p < 0.85$ ). These results indicated that other factors, such  
256 as human activity, might have greater influence on the residues of antibiotics. The microbial  
257 communities of the studied soils were previously reported (Sun et al., 2016a). In this study, the  
258 impacts of antibiotics on soil microbial communities were assessed, and no clear correlation was  
259 found between anti-biotic concentrations and the abundance of bacteria, fungi, and actinomycin  
260 in soil.

261 In this study, the cluster analysis was conducted to determine the combined characteristics of all  
262 the target antibiotics in the region (Fig. 2). The 13 antibiotics were divided to two groups. The first  
263 major group contained seven antibiotics including OTC, ENR, CIP, NOR, SDM, SMTZ, and SMX.  
264 The second groups contained six antibiotics including DC, OFL, SMZ, SMR, SDZ, and TC. In  
265 general, the chemicals in the same branch of the same group have a closer correlation, such as CIP  
266 and NOR, indicating their positive correlation in soils. It may be deduced for coexisting  
267 compounds based on the cluster analysis that the compounds share common source or have  
268 analogical sorption affinity even though they are in different classes of pollutants. On the contrary,  
269 a class of chemicals may not be divided into the same group. For instance, ENR, CIP and NOR  
270 were in one group while OFL was in another group. This indicated that ENR, CIP and NOR  
271 showed similar environmental source and fate in soils, whereas there might be distinct input  
272 sources of OFL. The results of cluster analysis can provide reference for combined antibiotic  
273 pollution in the regional-scale assessment.

274

### 275 **Potential ecological risk and occurrence of ARGs**

276 The potential ecological risk of contaminants such as antibiotics in the environment could be  
277 assessed by using the mean values of risk quotient (RQs). In the present study, the RQ value was  
278 equal to the quotient between the measured environmental concentration (MEC) and the predicted  
279 no effect concentration (PNEC) (European Commission, 2003). High levels of antibiotics, such as  
280 the TCs and QNs, were frequently detected in the agricultural soils, and their ecological risk should  
281 be determined. The PNEC values were obtained from toxicity assessment (Li et al., 2013; Zhang  
282 et al., 2013). The values of PNEC of OTC and TC were obtained directly from the published  
283 literature (Vaclavik et al., 2004; Thiele-Bruhn and Beck, 2005). As for QNs and DC, only a small

284 percentage of the literature reported the ecological risks to soils (Gao et al., 2008). Thus, it is  
285 impossible to obtain the values of PNEC in soils to calculate the RQs. In our study, PNEC values  
286 in soils were calculated from the PNEC values in water ( $PNEC_{water}$ ) as shown in Table S5.

287 For the assessment of ecological risk of antibiotics in agricultural soils, RQ values were divided  
288 into three risk levels: high ( $RQ \geq 1$ ), medium ( $0.1 < RQ < 1$ ), and low ( $0.01 < RQ < 0.1$ ) (Martin et  
289 al., 2012; Zhang et al., 2013). In this study, DC, OTC, TC, OFL, ENR, CIP, and NOR may cause  
290 high ecological risk, in 64.3%, 5.4%, 1.2%, 4.6%, 2.1%, 30.7%, and 7.5% of the studied  
291 agricultural soils, respectively (Fig. 3). The potential ecological risk of TC is the lowest, with 93.4%  
292 of soil samples pose a low or no risk. In the YRD region, DC and CIP showed the most potential  
293 ecological effects. In greenhouse soil samples from Beijing, OTC, NOR, CIP and ENR might pose  
294 high ecological risk with the proportions of 39.3%, 10.7%, 17.9%, and 41.1%, respectively (Li et  
295 al., 2015). It was reported that DC may cause high risk in 44.3% of the soil samples collected in  
296 organic farms of south China (Xiang et al., 2016), which was similar to the condition in this study.

297 According to these calculations, it was considered that the measured levels and distribution of  
298 antibiotics can result in different degrees of ecological risks the region, and the risks should receive  
299 more attention especially for antibiotic resistance genes in the environment. Nevertheless, the risk  
300 was calculated through the concentration of individual analytes without the consideration of  
301 coexisting pollutants (Zhu et al., 2013), such as PCBs and OCPs in studied area (Sun et al., 2016a,  
302 2016b; Cai et al., 2017). The actual risk may be higher than expected (Chen and Zhou, 2014).

303 The analysis of ARGs in a portion of these soils was conducted in this study to provide a  
304 preliminary result of the occurrence of ARGs in the region. The three ARGs (i.e., tetA, sulI, and  
305 qnrS) were all detected in 15 soil samples with a concentration of

306  $3.5 \times 10^5 - 2.1 \times 10^8$  copies per dry soil for *sulI*,  $3.6 \times 10^6 - 8.1 \times 10^7$  copies per dry soil for *tetA*, and  
307  $2.5 \times 10^5 - 4.2 \times 10^7$  copies per dry soil for *qnrS*, respectively. The relative abundance of *tetA* and *qnrS*  
308 were more than that of *sulI* gene in most soils, which might be due to the comparatively higher  
309 concentration of TCs and QNs. The correlation analysis between antibiotics and ARGs indicated  
310 that total antibiotics, TCs, and QNs showed significant positive correlation with *sulI* in 15 soil  
311 samples ( $p < 0.01$ ). However, SAs and QNs showed no significant correlation with *sulI* and *qnrS*,  
312 respectively. These results may be caused by cross-selection of the studied antibiotics to the  
313 related ARGs (McKinney et al., 2010; Shen et al., 2008). The environmental behavior of  
314 antibiotics and ARGs may be influenced by many factors. Here, we only assessed several  
315 important parameters which may cause impact on the target antibiotics and ARGs. Redundancy  
316 analysis of the relative abundance of ARGs and environmental variables was shown in Fig. 4. The  
317 data were obtained from the correlation analysis of different components included in this study  
318 (Table S6). The measurement of Cu and Zn in these soils were reported in a previous study (Shao  
319 et al., 2016). Both Cu and Zn were positively correlated with *sulI*, which reflect the potential impact  
320 of metals on the abundance of this ARG (Hu et al., 2017). The soil properties such as TOC and  
321 TN were positively correlated with *sulI*, whereas pH showed a negative correlation with *sulI*.  
322 Comparatively, there was no significant correlation between *tetA* and antibiotics. Soil properties,  
323 Cu, and Zn also showed no clear influence on *tetA* and *qnrS* (Fig. 4). ARGs could be affected by  
324 many factors and *qnrS* and *tetA* may be imported directly into soils from other media such as  
325 manure and wastewater. In the selected 15 sampling sites, NE6, NE8, and NE12 are all rice paddy  
326 fields, and other sites are vegetable field, fruit garden or uncultivated field. The abundance of three  
327 ARGs were predominant in paddy soils of sites NE6, NE8 and NE12, among 15 soil samples (Fig.  
328 S1). Moreover, the highest concentration of antibiotics was also found in NE6. These results

329 indicated that water irrigation in paddy fields may intensify the contamination of antibiotics and  
330 ARGs (Chen et al., 2016a,b). The abundance of three ARGs covered from  $2.5 \times 10^5$  to  $2.1 \times 10^8$   
331 copies per dry soil and were lower than that in sewage sludge (Xie et al., 2016) and swine  
332 wastewater (Sui et al., 2016). In agricultural soils, ARGs had many sources and could be influenced  
333 by antibiotics or heavy metals (Luo et al., 2017). In addition, with more application of manure,  
334 sewage sludge and wastewater in agricultural soils, there will be aggravation of ARGs  
335 dissemination from the environment to the agricultural system, such as soils and crops (Zhu et al.,  
336 2017). As a result, the antibiotic pollution in soils and their potential damage to ecosystem and  
337 human health require more systematic research in the future, and further data on the distribution  
338 of ARGs in the studied region will provide more information for the risk management of antibiotics  
339 in the environment.

340

## 341 **CONCLUSION**

342 This study revealed the occurrence and distribution of antibiotics including QNs, TCs and SAs  
343 and their associated ecological risk assessment by analyzing soil samples from the agricultural  
344 fields in the YRD, China. Across the studied area, the levels of total antibiotics fell in the range  
345 4.55 - 2,010 ng/g dry weight with a detection rate of 100%. The concentrations of total antibiotics  
346 were higher in Shanghai and Jiangsu than in Zhejiang. The mean concentrations of three classes  
347 of antibiotics measured in all soil samples were in the following order: QNs (48.8 ng/g) > TCs  
348 (34.9 ng/g) > SAs (2.35 ng/g). Ecological risk assessment based on the risk quotient values  
349 revealed that tetracyclines and quinolones could pose potential ecological risks, and doxycycline  
350 showed the higher risk, followed by ciprofloxacin. These results suggest that regular monitoring  
351 of antibiotics in regional-scale agricultural soils is needed to prevent the risks from the excessive



352 use of antibiotics in the livestock and fishing farming industry, and proper management of their  
353 dis- charges to the environment.

354

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360

## 361 Appendix A. Supplementary data

362 Supplementary data related to this article can be found at [http://](http://dx.doi.org/10.1016/j.chemosphere.2017.09.040)  
363 [dx.doi.org/10.1016/j.chemosphere.2017.09.040](http://dx.doi.org/10.1016/j.chemosphere.2017.09.040)

364

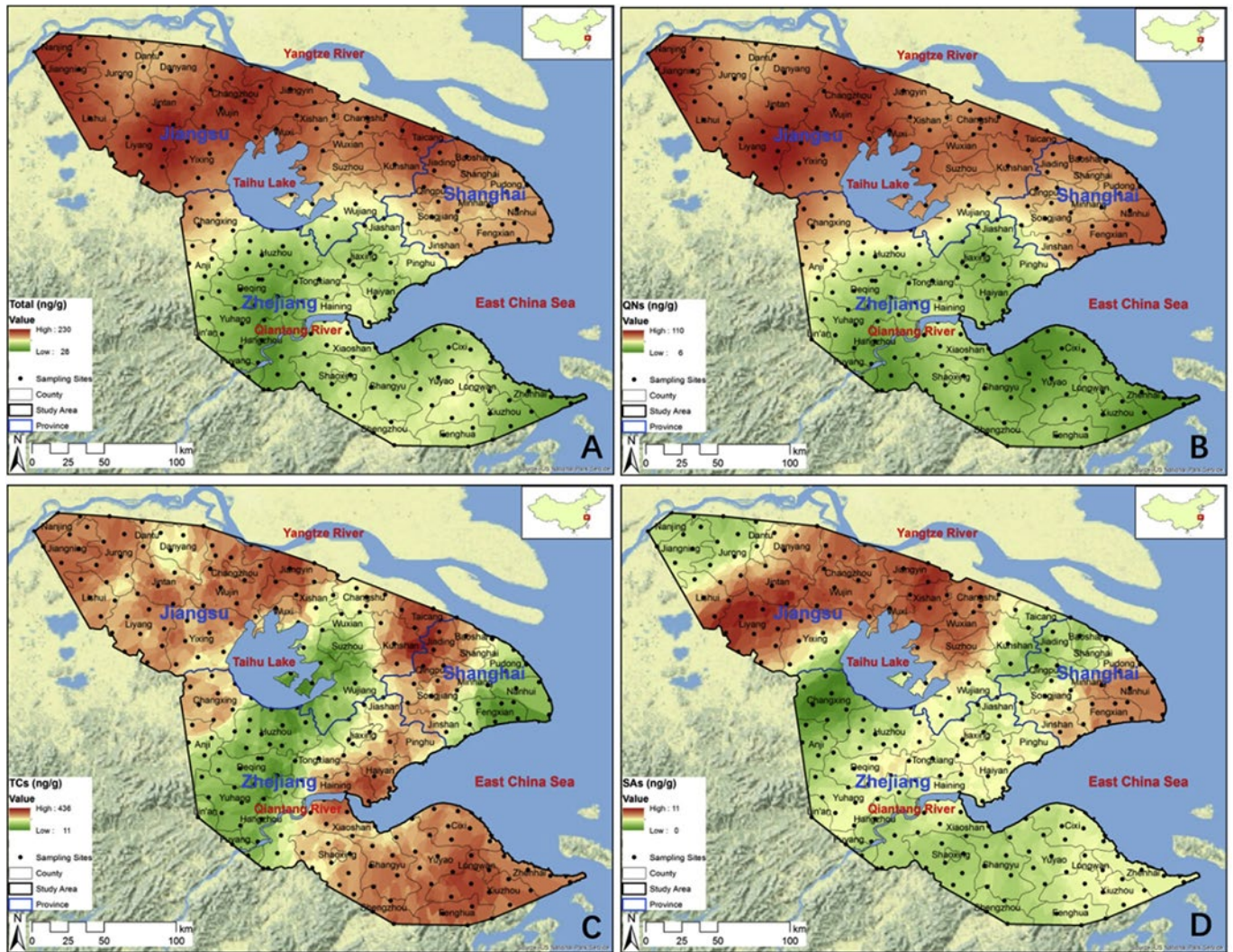
## 365 LIST OF TABLES AND FIGURES

**Table 1**  
Concentrations (ng/g) of antibiotics in the agricultural soil samples in the YRD region, China.

Compounds	Frequency (%)	Mean (ng/g)	Median (ng/g)	Min (ng/g)	Max (ng/g)
SDM	44	1.46	ND	ND	21.5
SMZ	3	0.01	ND	ND	0.63
SMTZ	14	0.55	ND	ND	111
SMR	0.8	0.01	ND	ND	0.71
SDX	41	0.29	ND	ND	2.44
SDZ	5	0.03	ND	ND	0.91
ΣSAs	66.8	2.35	0.92	ND	111
DC	95	13.9	11	ND	105
OTC	68	18.9	13.2	ND	530
TC	37	2.07	ND	ND	197
ΣTCs	99.6	34.9	25.8	ND	809
OFL	72	5.86	0.88	ND	498
ENR	75	3.98	1.02	ND	237
CIP	83	27.7	16.6	ND	1030
NOR	73	11.2	3.98	ND	505
ΣQNs	99.0	48.8	27.1	ND	1410
Σantibiotics	100	85.6	56.2	4.55	2010

ND: not detected.

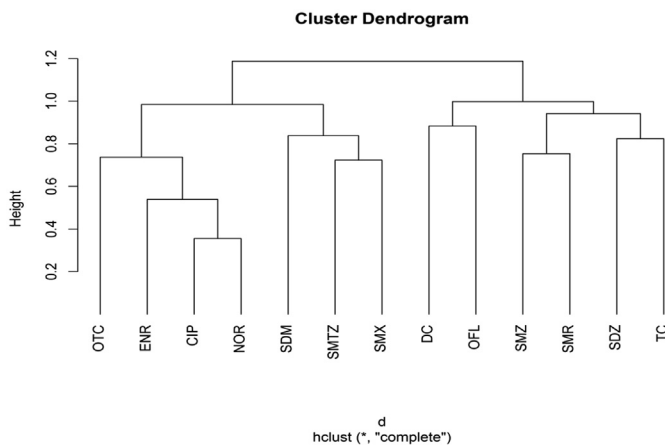
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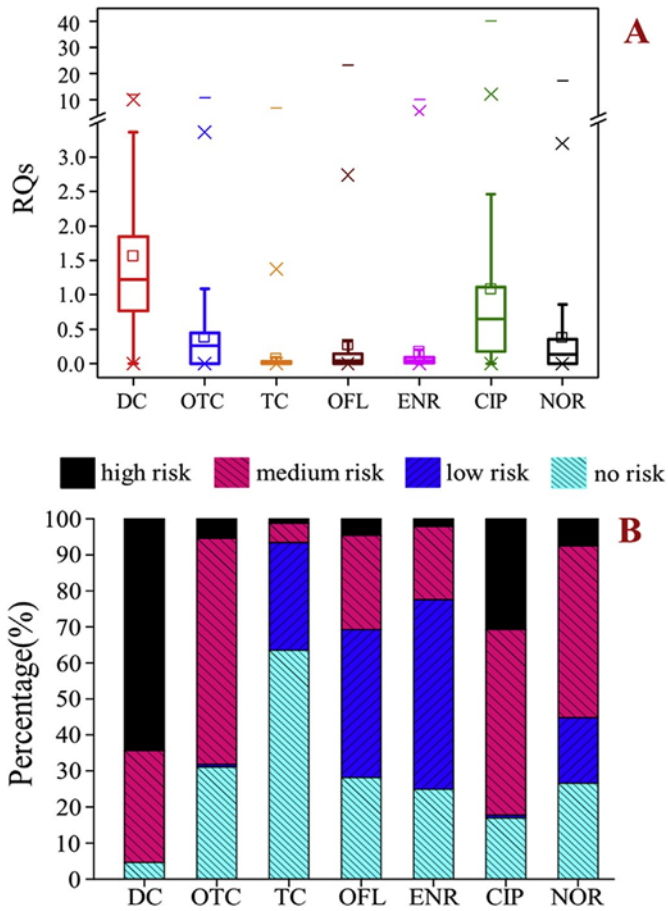
369 Fig. 1. Spatial distributions of (A) total antibiotics, (B) QNs, (C) TCs, and (D) SAs in the  
 370 agricultural soils of the YRD region

371



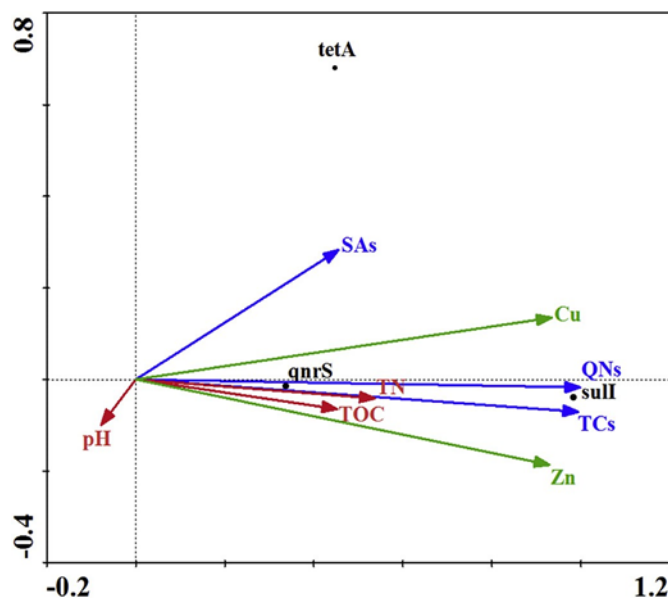
372

373 Fig. 2. Cluster dendrogram for 241 soil samples from the YRD region through cluster analysis.  
 374



375  
 376 Fig. 3. Risk quotients (A) and percentages (B) for QNs and TCs detected in the agricultural soils  
 377 of the YRD region

378



379  
380 Fig. 4. Redundancy analysis of the relative abundance of ARGs and environmental variables  
381 (including various antibiotics, copper, zinc, pH, TOC, and total nitrogen) in 15 soil samples from  
382 northeast of the YRD region.

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