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Review

The Role of Autophagy in Lupus Nephritis

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Abstract: Systemic lupus erythematosus (SLE) is a multifactorial autoimmune disease characterized by the generation of immune responses to self-antigens. Lupus nephritis is one of the most common and severe complications in SLE patients. Though the pathogenesis of lupus nephritis has been studied extensively, unresolved questions are still left and new therapeutic methods are needed for disease control. Autophagy is a conserved catabolic process through which cytoplasmic constituents can be degraded in lysosome and reused. Autophagy plays vital roles in maintaining cell homeostasis and is involved in the pathogenesis of many diseases. In particular, autophagy can affect almost all parts of the immune system and is involved in autoimmune diseases. Based on genetic analysis, cell biology, and mechanism studies of the classic and innovative therapeutic drugs, there are growing lines of evidence suggesting the relationship between autophagy and lupus nephritis. In the present review, we summarize the recent publications investigating the relationship between autophagy and lupus nephritis and provide a new perspective towards the pathogenesis of lupus nephritis.

Keywords: autophagy; lupus nephritis; systemic lupus erythematosus

1. Introduction

Systemic lupus erythematosus (lupus; SLE) is a prototype of chronic autoimmune inflammatory disease characterized by loss of tolerance against self-antigens, polyclonal autoantibody production, immune complex formation, and deposition in different part of the body, leading to detrimental

inflammation and multi-organ injuries [1]. Among the wide spectrum of complications of SLE, one of the most common and severe is lupus nephritis. Lupus nephritis occurs in 50%–70% of SLE patients in the first five years of diagnosis [2]. It may lead to permanent renal damage and chronic kidney disease. Renal involvement early in the course of SLE becomes a major predictor of poor prognosis [2]. Although the disease has been known for centuries, the pathogenesis remains unclear. The current treatments for lupus nephritis rely mainly on glucocorticoids and immunosuppressants. They are partially effective and have considerable adverse effects, especially for long-time use. New therapeutic strategies and drugs are eagerly demanded.

Autophagy is an evolutionarily conserved catabolic process to degrade cytoplasmic contents through lysosomes [3]. Depending on the ways delivering cytoplasmic materials to the lysosomes, at least three major types of autophagy have been identified: macroautophagy, microautophagy, and chaperone-mediated autophagy (CMA), with macroautophagy being the best understood [4]. Macroautophagy (hereafter called autophagy) describes a degradation process, through which unique double-membrane vesicles (autophagosomes) are formed to sequester a targeted portion of cytoplasm, then fuse with lysosomes to form autolysosomes and degrade the internal materials. Microautophagy and microautophagy have the ability to engulf structures through non-selective and selective mechanisms [6]. CMA involves direct translocation of unfolded proteins across the lysosome membrane with the help of chaperones. Particularly, cytosolic proteins containing a pentapeptide motif will be recognized by chaperone heat shock cognate 70 (Hsc70) and delivered to the surface of lysosomal membrane through the interaction between Hsc70 and the receptor lysosome-associated membrane protein 2A (LAMP-2A) [6,7].

Autophagy exists at basal level in all cell types and will be activated in response to different stresses [8]. Autophagy is being regulated precisely and plays a vital role in maintaining cell homeostasis. It is a typical protective, pro-survival response at the initiation of damages. However, if hyperactivated, it will ultimately kill the cells, known as type II programmed cell death. Autophagy has many physiological roles and is involved in the pathogenesis of diverse diseases, including cardiovascular diseases, neuro-degeneration, obesity, aging, cancers, infections, autoinflammatory diseases, and autoimmune diseases (reviewed in [9–11]).

Especially, autophagy also affects diverse aspects of immune system (reviewed in [12,13]). Autophagy can directly eliminate intracellular pathogens (xenophagy [14]). Autophagy participates in inflammation. It can regulate type I interferon production by virally-infected cells, however some reports are controversial [15–18]. Inflammasomes are molecular platforms activated under infection or stresses which trigger the maturation of proinflammatory cytokines and play critical roles in host defence and inflammation. Evidence shows that autophagy can negatively regulate inflammasome activation [19]. Assembled inflammasomes would recruit autophagic adaptor p62 to assist their delivery to autophagosomes and the degradation within them [20]. Autophagy can promote MHC class II presentation of endogenous antigens [21]. Autophagy is vital for T cell and B cell homeostasis [22]. Besides, autophagy is involved in autoimmunity. Genome-wide association studies (GWAS) have linked polymorphisms in autophagy-related genes to some autoinflammatory and autoimmune diseases, specifically in inflammatory bowel disease (IBD) and SLE [23]. As lupus nephritis results from immune system disorders, we hypothesize that autophagy may be involved in lupus development and renal injuries. In this review, we discuss recent publications on the relationship between autophagy and lupus

nephritis, with a view of providing new insights and research directions into the pathogenesis and treatment of lupus nephritis.

2. Genes

Lupus is a complex disease and many factors interplay and contribute to the disease, including the genetic, environmental, hormonal and immunoregulatory factors. The rapid development of GWAS has revealed more than 40 genes associated with SLE (based on the data from the National Human Genome Research Institute's Catalog of Published Genome-Wide Association Studies, http://www.ebi.ac.uk/gwas, [24]). Among them, some genes have shown to be highly related to autophagy, including *ATG5* [25,26], *ATG7* [27], *IRGM* (immunity-related GTPase family M) [27], *DRAM1* (DNA damage regulated autophagy modulator) [28], *CDKN1B* (cyclin-dependent kinase inhibitor 1 B) [28], *APOL1* (apolipoprotein L1) [29], and *MTMR3* (Myotubularin-related phosphatase 3) [30]. We will point out some genes which are highly related to lupus nephritis.

2.1. APOL1

Apolipoprotein L1 (APOL1) is a BH3-only protein that can induce autophagic cell death in a variety of cells when it is overexpressed [31]. In 2013, Larsen and colleagues examined 546 renal biopsies from African American SLE patients and 26 cases of collapsing glomerulopathy (CG) were identified. DNA was extracted from the biopsy tissues and genotyped for APOL1 risk alleles. Their results showed a strong association between APOL1 and SLE-associated CG (p < 0.001). To explain the role of APOL1 in the pathogenesis of SLE-associated CG, they described a "two-hit hypothesis", in which presence of APOL1 risk alleles together with altered inflammatory milieu helped to disease development [32]. In a subsequent study, Freedman and colleagues genotyped the SNPs in APOL1 (rs73885319 and rs60910145 in G1 allele, and rs71785313 in G2 allele) in 855 African American SLE patients with end-stage renal disease (ESRD) and 534 African American SLE patients without renal involvement. Their results also revealed a strong association between APOL1 G1/G2 alleles and the risk of SLE-ESRD in African Americans (odds ratio 2.57, recessive model $p = 1.49 \times 10^{-9}$) [29]. In a more recent focal segmental glomerulosclerosis clinical trial, Kopp et al. [33] studied the APOL1-asosociated nephropathy and APOL1 renal risk allele frequency in 94 cases of children and young adults. It was demonstrated that APOL1 risk alleles is associated with more severe kidney fibrosis at diagnosis and more rapid progression into ESRD. As elaborated in the Editorial comments by Larsen and Freedman [34], the pathologic mechanisms and the primary kidney cell types related to APOL1 remain unknown. Combining the biochemistry and clinical data, it is probable that the expression of APOL1 is associated with autophagy in renal tissues of SLE patients. Further investigation is needed to substantiate this hypothesis.

2.2. MTMR3

Myotubularin-related phosphatase 3 (MTMR3) is one of the phosphatidylinositol 3-phosphate (PI3P) phosphatases [35]. These phosphatases together with PI kinases reciprocally regulate the cellular level of PI3P and are involved in constitutive autophagy initiation and autophagosome size [36]. Zhou and colleagues [30] identified a significant association between a genetic variant of *MTMR3* (rs9983A) and

the risk of lupus nephritis in northern Han Chinese populations. Expression analysis revealed lower *MTMR3* transcriptions in the blood samples with this variant and in renal biopsy samples from lupus nephritis patients.

3. Environmental Risk Factors

Epidemiologic studies reveal the importance of environmental factors in lupus and lupus nephritis [37]. A genetically susceptible individual exposed to risk environments, such as ultraviolet (UV) light [38], Epstein-Barr virus (EBV) infections [39,40], and smoking [41], will facilitate and enhance the disorders of immune systems and aggravate diseases.

3.1. UV Light

UV light exposure is a risk factor for lupus development. Photosensitivity is a well-known characteristic of lupus and there are reports that sunlight can lead to cutaneous damage and life-threatening lupus nephritis [42]. Kemp and colleagues [43] exposed keratinocytes, monocytes and T cells to UV light and demonstrated that UV-induced DNA damage led to lower protein expressions of AMBRA1 and ULK1, which negatively regulated STING (stimulator of interferon genes) and further led to activation of transcription factor IRF3 (interferon regulatory factor 3). Although the authors did not evaluate the autophagy process in their cells after UV exposure, AMBRA1 and ULK1 are important machineries in autophagy and these results warrant for further investigations on the relationship among UV exposure, dysregulation of autophagy, and organ damage in lupus patients.

3.2. Epstein-Barr Virus (EBV)

The association between EBV infection and lupus development has been investigated for decades [44]. Nearly 100% of lupus patients are positive for EBV-VCA (EBV viral capsid antigen) IgG response [45,46]. Some antibodies against EBV antigens such as EBNA-1 (EB nuclear antigen 1) and EBNA-2 (EB nuclear antigen 2), can cross-react with autoantigenic proteins such as Sm and Ro and support the hypothesis that some humoral autoimmunity in lupus arises through molecular mimicry [47]. Besides, increased EBV load, altered cellular and humoral immune response to EBV, and increased viral gene expressions in peripheral blood mononuclear cells in lupus patients, implicated the important roles of EBV in lupus pathogenesis [44].

The roles of EBV infections in lupus nephritis have also been investigated. Yu and colleagues [48] collected 58 renal tissue samples from patients with lupus nephritis and compared them with samples from patients with non-glomerular hematuria and patients with minimal change nephropathy. They detected expression of EBV-latent membrane protein 1 (LMP1) in renal tissues using immunohistochemistry while they detected the expression of EBV-encoded RNA 1 (EBER-1) using *in situ* hybridization. Results showed that positive rates of renal viral proteins EBER-1 and LMP-1 were significantly higher in lupus patients than those in control groups. And the percentage of anti-Sm antibody positive cases in lupus patients was also higher in renal EBV-positive groups than EBV-negative groups. Ding and colleagues [49] examined 51 renal samples from young patients with lupus nephritis (aged from 6 to 16 years old) and found similar results. Positive rate of LMP1 in renal tissues was higher in young

patients with lupus nephritis than that in control group and the proportion of young patients positive for anti-Sm antibody was higher in LMP1 positive group. These results suggested that renal EBV infection might be relevant to renal injuries in patients with lupus nephritis.

Autophagy is involved in EBV infections. LMP1 could induce autophagy in B cells in a dose-dependent manner. Inhibiting autophagy promoted accumulation of LMP1 in EBV-positive cells [50]. Autophagy is also involved in MHC class II mediated presentation of endogenous antigen EBNA-1 to CD4⁺ T cells and participated in antiviral responses [51]. Thus, autophagy may affect disease course partially though dysregulated immunity linked to EBV.

4. Cells

Lupus is a systemic autoimmune disorder. Many cells in innate and adaptive immunity are affected and dysregulated, especially including the B cells, T cells, dendritic cells (DCs) and phagocytes. For example, Clarke et al. [52] demonstrated that autophagy in splenic CD19⁺ B cells from NZB/W lupus mice was increased compared with C57BL/6 control mice. Increased autophagy was also observed in B cells in human lupus patients. Differentiation from B cells to plasma cells also required autophagy. Inhibiting autophagy by using Atg7 deficient B cells reduced the number and viability of CD19⁺CD138⁺ plasma cells and decreased secretion of IgM. Chemical autophagy inhibitor 3-MA can also dramatically reduced plasmablast formation and cell proliferation. Previous studies have investigated the role of autophagy in plasma cells differentiation and homeostasis [53]. By using CD19-cre×Atg5 flox/flox mice in which Atg5 was conditionally deleted in CD19-expression cells, Pengo and colleagues [53] revealed that autophagy showed a negative control on endoplasmic reticulum (ER) expansion and immunoglobulin synthesis in plasma cells, via a crucial transcriptional repressor Blimp-1, and demonstrated a protective role for plasma cell maintenance by sustaining energy metabolism and cell viability. Conway and colleagues [54] also demonstrated the importance of autophagy related protein Atg5 in late B cell activation and plasma cell differentiation. Gros et al. [55] reported that central and peripheral T cells from lupus-prone mice (MRL/lpr and NZB/W mice) exhibited increased autophagic flux and autophagosomes compared with CBA/J and BALB/c control mice. Peripheral T cell from lupus patients also exhibited increased number of autophagic vacuoles.

Besides, lupus can lead to multiple end-organ injuries, among which renal involvement is associated with more severe symptoms and poorer prognosis. Thus renal parenchyma cell injuries should also be considered. Here we will discuss different types of cells which are involved in renal damage and present dysregulated autophagy.

4.1. Neutrophils and Neutrophil Extracellular Trap (NET)

Neutrophils are involved in the pathogenesis of lupus. Genomic studies showed that neutrophil-specific genes are the second most prevalent peripheral-blood mononuclear cell transcriptional signature in children with SLE [24,56]. Infections can initiate flares and are major causes of mortality in SLE patients. When facing infections, neutrophils are recruited to infection sites and release NETs to play the antimicrobial role. The NETs are composed of decondensed chromatin fibers and neutrophil proteins, which are released into the extracellular matrix as the neutrophil undergoes a unique form of cell death—NETosis, to trap and kill invading pathogens. The NETs can activate plasmacytoid dendritic

cells through Toll Like Receptors (TLRs) to produce massive type I interferon. If the released NETs cannot be removed timely, they may become a source to be presented as self-antigens and stimulate more autoantibody production. Hakkim *et al.* [57] demonstrated that impairments of NET degradation existed in a subset of SLE patients (36.1% of the 61 SLE patients) and the impairments are correlated with lupus nephritis. Serum DNase 1 is essential for degrading NETs. Existence of DNase 1 inhibitors in serum or anti-NET antibody that prevented DNase 1 accessing to NETs may explain the inhibitory mechanisms of NETs degradation. SLE patients who have impaired NETs degradation were more likely to develop lupus nephritis. They also had higher titers of anti-ds DNA antibodies (which are correlated with renal disease) than the degraders. Martinez-Valle *et al.* reported that DNase 1 activities in SLE patients were significantly lower than that in control group (13.69 ± 8.52 vs. 24.75 ± 12.32 µg/mL, *p* < 0.005) [58]. Leffler *et al.* found that NETs could activate complement *in vitro* and deposited C1q could inhibit NET degradation by direct inhibition of DNase 1 [59].

Evidence shows that autophagy affects the formation and release of NET. Itakura *et al.* [60] analysed the kinetics of NET release by monitoring the extracellular release of DNA and one well-known NET protein neutrophil elastase (NE) from activated neutrophils, using the cell-impermeable DNA dye Sytox green and immunofluorescence staining of NE protein. Bacteria-derived peptide formyl-Met-Leu-Phe (fMLP) can activate and stimulate neutrophils to lease NET, proved by increased percentage of Sytox-positive cells, in a time-dependent manner. After pretreatment with mTOR inhibitors rapamycin or WYE-354 (which can induce autophagy), more Sytox-positive neutrophils and increased NE expression were detected under fMLP stimulation, compared with treatment with vehicle or fMLP alone. Meanwhile, mTOR inhibitors rapamycin and WYE-354 can promote autophagosome formation and induce autophagy. These results indicated that suppressed mTOR activity and increased autophagy may lead to increased NET release, and thus may affect lupus development.

4.2. Macrophages

The macrophage is an important component of the innate immune system, participating in antigen presentation, removal of dying cells and their released components, cytokines production and many other immunological responses. In lupus patients, deficient phagocytic capacity of macrophages was observed and led to accumulation of cell materials, which may provide a source of autoantibodies [61]. Evidences showed that autophagy could regulate macrophage functions and cell homeostasis and participated in the pathogenesis of lupus nephritis. Li and colleagues [62] detected significantly increased LC3-II expression in spleen and kidney macrophages from activated lymphocytes-derived DNA (ALD-DNA) induced lupus mice when compared with those in control mice. For in vitro experiments, ALD-DNA stimulation can induce autophagy in cultured RAW264.7 macrophages, proved by increased expressions of LC3-II, beclin 1 and Atg5. The authors further generated Beclin 1 knockdown macrophages by using a lentiviral shRNA. Then they adoptively transferred these autophagy-suppressed macrophages into lupus mice which were previously depleted of the original macrophages with dichloromethylenediphosphonic acid disodium salts (DMDP). Results showed alleviated symptoms in lupus mice receiving these autophagy-suppressed macrophages, demonstrated by relieved renal pathological severity, decreased anti-dsDNA titer, and declined urine protein. These results suggested that increased autophagy in macrophage may play an important role in the pathogenesis of murine lupus and kidney damage.

4.3. Podocytes

Podocyte, also called visceral glomerular epithelial cells, is one of the three components forming the glomerular filtration barrier. They are injured in many forms of glomerular diseases, including lupus nephritis. Cumulative evidence has revealed protective roles of autophagy in podocytes injured by chemicals [63], aging [64] or diseases [65]. Especially, Hartleben *et al.* [64] generated podocyte-specific Atg5 knockout mice (Atg5^{Δ podocyte}) to indicate the protective roles of autophagy in podocyte homeostasis and glomerular diseases susceptibility. Atg5^{Δ podocyte} mice developed proteinuria and glomerulosclerosis after 20 to 24 month follow-up compared with controls. Enhanced ER stress and accumulated aggregates of ubiquitinated proteins led to loss of podocytes. Besides, they also demonstrated that glomerular diseases also led to upregulation of autophagy in podocytes. When inducing glomerular disease models in asymptomatic young Atg5^{Δ podocyte} mice, they observed tremendous increase of albuminuria in autophagy-deficient mice compared to the controls. These results implied the important roles of autophagy in podocyte survival.

However, there are less direct evidences showing the role of autophagy in podocyte injuries in lupus nephritis. A German research group observed that podocytes could uptake SLE autoantibodies. When stimulated by anti-dsDNA autoantibodies which were isolated from lupus patients, podocytes took up and aggregated these antibodies in cytosolic speckles. These aggregates harmed podocytes survivals and could be degraded by autophagy. After inhibition of autophagy with Bafilomycin A or MG132, the number of cytoplasmic aggregates significantly increased [66]. These results indicate that autophagy participates in autoantibodies degradation within podocytes and is involved in regulating podocytes injuries. Maintaining podocyte health may be new therapeutic strategies for lupus nephritis.

5. Drugs

The current treatments for lupus nephritis are mainly dependent on glucocorticoids and immunosuppressants [67]. They affect disease process through different mechanisms and the specific mechanisms within individual diseases are still unrevealed. These medicines are partially effective and have considerable adverse effects, especially for long-time use. Biologic agents with more specific targets represent a promising direction of treatments but clinical trials and extensive follow-ups are needed [67].

Many drugs for lupus can act as autophagy regulators, however their effects appear paradoxical on autophagy process. Glucocorticoid is a classical drug used for lupus for decades and has wide suppressant effects of the whole immune system. Recent evidences show that corticosteroid can inhibit cytoplasmic calcium signaling and induce autophagy in WEHI 7.2 cells (a CD4/CD8 double positive T cell lines) [68]. Chloroquine (CQ) or hydroxychloroquine (HCQ), which are first regarded as antimalarial agents, also present numerous immunomodulatory effects and belong to the first-line therapies for lupus. However, they belong to autophagy inhibitors, as they can raise the lysosomal pH value and impaired the last step of autophagy and autophagic protein degradation. Besides acting as autophagy inhibitor, CQ/HCQ can reduce proinflammatory cytokines production, inhibit BCR/TCR-mediated calcium signaling, absorb and block UV reaction, and inhibit TLR signalings, all of which may contribute to their beneficial effects in lupus patients [69].

5.1. Inhibition of Activated mTOR Pathway in Lupus Nephritis

Activated mTOR pathway has been detected in lupus nephritis and has emerged as a central pathway for the pathogenesis [70]. Treatments with rapamycin and other mTOR inhibitors, have displayed protective and benefit effects on murine lupus nephritis [71,72] and patients with lupus nephritis [73]. A Greek research group studied 32 female NZBW/F1 mice divided into four groups: (1) healthy control group (HG), in which mice were sacrificed before proteinuria development; (2) untreated group (UG), in which the mice were left untreated and developed into lupus nephritis; (3) preventive group (PG), in which mice were treated with rapamycin before they developed proteinuria; and (4) therapeutic group (TG), in which rapamycin was given after the mice were presented with severe proteinuria. Results showed that total and phosphorylated form of Akt and mTOR were increased in the kidney cortex in mice with lupus nephritis when compared with HG mice. Rapamycin intervention (in PG and TG) prolonged mice survival, reduced proteinuria, serum creatinine and anti-ds DNA titer, and alleviated renal histological lesions. They also proved that rapamycin intervention reduced the Akt and mTOR expression in TG and PG mice. These results indicated abnormal activated PI3K/Akt/mTOR pathway in glomerulus in murine lupus nephritis [74]. As this pathway is one of the most important strategies to regulate autophagy, this result implies a relationship between autophagy and lupus nephritis.

Moreover, rapamycin could act as immunosuppressant, which will also benefit patients with lupus nephritis. The effects of rapamycin on lupus nephritis and the underlying mechanisms are complex. Besides its benefits, rapamycin nephrotoxicity is observed to develop in clinical practice [75]. Different dose of rapamycin also exhibit different effects. Zhang and his colleague observed that low dose of rapamycin increased the frequency of antigen-active lymphocytes and decreased activation-induced cell death in T cells, resulting in exacerbated autoimmune experimental uveitis in B10.RIII mice [76]. Thus, we need to carefully differentiate its effects in specific situations.

5.2. P140

P140 is a splicesomal peptide (sequence 131–151 of the U1-70K small nuclear ribonucleoprotein) phosphorylated at Ser140. In a Phase IIb clinical trial, P140 (Lupuzor) treatment improved Systemic Lupus Erythematosus Disease Activity Index (SLEDAI) score in lupus patients [77]. Administration of P140 in saline to MRL/lpr lupus-prone mice before they presented symptoms could prolong mice survival, reduce proteinuria, and decrease titer of anti-dsDNA antibodies [78]. P140 treatment could also alleviate renal vasculitis, glomerulonephritis and dermatitis in MRL/lpr mice. Evidences showed that P140 could bind with chaperone HSC70 protein (which played a vital role in CMA), decrease its expression in MRL/lpr splenic B cells and impaired the refolding properties of this chaperone. P140 treatment could suppress the autophagic flux, with increased accumulation of p62 and LC3-II in B cells from MRL/lpr mice [79]. The effects of P140 on lupus nephritis indicated the potential role of autophagy on this disease.

6. Conclusions

There are emerging evidences indicating the relationship between autophagy and lupus nephritis. GWAS in human studies reveals autophagy-relative genes associated with lupus nephritis. Environmental risk factors for lupus nephritis can regulate the autophagy process. Autophagy can affect functions of diverse immune cells and the immune responses. The widely-used first-line drugs and innovative drugs for treatment of lupus nephritis can function as autophagy regulators. The current roles of autophagy in lupus nephritis are summarized in Figure 1. Further understanding the roles of autophagy in lupus nephritis and other autoimmune diseases, may develop new views to interpret the mechanisms of these complex diseases and autophagy process may provide new targets for diagnosis and therapy of autoimmune diseases.



Figure 1. The roles of autophagy in lupus nephritis. Abbreviations: SLE: systemic lupus erythematosus; LN: lupus nephritis; EBV: Epstein–Barr virus; NETs: neutrophil extracellular traps.

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Author Contributions

Linlin Wang and Helen Ka Wai Law conceived and wrote the paper.

Conflicts of Interest

The authors declare no conflict of interest.

References

- 1. Lech, M.; Anders, H.J. The pathogenesis of lupus nephritis. J. Am. Soc. Nephrol. 2013, 24, 1357–1366.
- 2. Maroz, N.; Segal, M.S. Lupus nephritis and end-stage kidney disease. *Am. J. Med. Sci.* **2013**, *346*, 319–323.
- 3. Levine, B.; Klionsky, D.J. Development by self-digestion: Molecular mechanisms and biological functions of autophagy. *Dev. Cell* **2004**, *6*, 463–477.
- 4. Mizushima, N. Autophagy: Process and function. Genes Dev. 2007, 21, 2861–2873.
- 5. Mijaljica, D.; Prescott, M.; Devenish, R.J. Microautophagy in mammalian cells: Revisiting a 40-year-old conundrum. *Autophagy* **2011**, *7*, 673–682.
- 6. Mizushima, N.; Levine, B.; Cuervo, A.M.; Klionsky, D.J. Autophagy fights disease through cellular self-digestion. *Nature* **2008**, *451*, 1069–1075.
- 7. Bejarano, E.; Cuervo, A.M. Chaperone-mediated autophagy. Proc. Am. Thorac. Soc. 2010, 7, 29–39.
- He, C.; Klionsky, D.J. Regulation mechanisms and signaling pathways of autophagy. *Annu. Rev. Genet.* 2009, *43*, 67–93.
- 9. Levine, B.; Kroemer, G. Autophagy in the pathogenesis of disease. Cell 2008, 132, 27-42.
- 10. Shintani, T.; Klionsky, D.J. Autophagy in health and disease: A double-edged sword. *Science* **2004**, *306*, 990–995.
- 11. Nixon, R.A. The role of autophagy in neurodegenerative disease. Nat. Med. 2013, 19, 983–997.
- 12. Levine, B.; Deretic, V. Unveiling the roles of autophagy in innate and adaptive immunity. *Nat. Rev. Immunol.* **2007**, *7*, 767–777.
- 13. Schmid, D.; Munz, C. Innate and adaptive immunity through autophagy. Immunity 2007, 27, 11–21.
- 14. Gomes, L.C.; Dikic, I. Autophagy in antimicrobial immunity. Mol. Cell 2014, 54, 224-233.
- 15. Harris, J. Autophagy and cytokines. Cytokine 2011, 56, 140-144.
- 16. Lee, H.K.; Lund, J.M.; Ramanathan, B.; Mizushima, N.; Iwasaki, A. Autophagy-dependent viral recognition by plasmacytoid dendritic cells. *Science* **2007**, *315*, 1398–1401.
- Jounai, N.; Takeshita, F.; Kobiyama, K.; Sawano, A.; Miyawaki, A.; Xin, K.Q.; Ishii, K.J.; Kawai, T.; Akira, S.; Suzuki, K.; *et al.* The Atg5 Atg12 conjugate associates with innate antiviral immune responses. *Proc. Natl. Acad. Sci. USA* 2007, *104*, 14050–14055.
- Shrivastava, S.; Raychoudhuri, A.; Steele, R.; Ray, R.; Ray, R.B. Knockdown of autophagy enhances the innate immune response in hepatitis C virus-infected hepatocytes. *Hepatology* 2011, 53, 406–414.
- Saitoh, T.; Fujita, N.; Jang, M.H.; Uematsu, S.; Yang, B.G.; Satoh, T.; Omori, H.; Noda, T.; Yamamoto, N.; Komatsu, M.; *et al.* Loss of the autophagy protein atg1611 enhances endotoxin-induced il-1β production. *Nature* 2008, 456, 264–268.
- Shi, C.S.; Shenderov, K.; Huang, N.N.; Kabat, J.; Abu-Asab, M.; Fitzgerald, K.A.; Sher, A.; Kehrl, J.H. Activation of autophagy by inflammatory signals limits II-1β production by targeting ubiquitinated inflammasomes for destruction. *Nat. Immunol.* 2012, *13*, 255–263.

- Dengjel, J.; Schoor, O.; Fischer, R.; Reich, M.; Kraus, M.; Muller, M.; Kreymborg, K.; Altenberend, F.; Brandenburg, J.; Kalbacher, H.; *et al.* Autophagy promotes MHC class II presentation of peptides from intracellular source proteins. *Proc. Natl. Acad. Sci. USA* 2005, *102*, 7922–7927.
- 22. Kuballa, P.; Nolte, W.M.; Castoreno, A.B.; Xavier, R.J. Autophagy and the immune system. *Annu. Rev. Immunol.* **2012**, *30*, 611–646.
- Zhou, X.J.; Zhang, H. Autophagy in immunity: Implications in etiology of autoimmune/autoinflammatory diseases. *Autophagy* 2012, *8*, 1286–1299.
- 24. Zhou, X.J.; Cheng, F.J.; Zhang, H. Emerging view of autophagy in systemic lupus erythematosus. *Int. Rev. Immunol.* **2015**, *34*, 280–292.
- Gateva, V.; Sandling, J.K.; Hom, G.; Taylor, K.E.; Chung, S.A.; Sun, X.; Ortmann, W.; Kosoy, R.; Ferreira, R.C.; Nordmark, G.; *et al.* A large-scale replication study identifies TNIP1, PRDM1, JAZF1, UHRF1BP1 and IL10 as risk loci for systemic lupus erythematosus. *Nat. Genet.* 2009, *41*, 1228–1233.
- 26. International Consortium for Systemic Lupus Erythematosus Group; Harley, J.B.; Alarcon-Riquelme, M.E.; Criswell, L.A.; Jacob, C.O.; Kimberly, R.P.; Moser, K.L.; Tsao, B.P.; Vyse, T.J.; Langefeld, C.D.; *et al.* Genome-wide association scan in women with systemic lupus erythematosus identifies susceptibility variants in ITGAM, PXK, KIAA1542 and other loci. *Nat. Genet.* 2008, *40*, 204–210.
- Zhou, X.J.; Lu, X.L.; Lv, J.C.; Yang, H.Z.; Qin, L.X.; Zhao, M.H.; Su, Y.; Li, Z.G.; Zhang, H. Genetic association of PRDM1-Atg5 intergenic region and autophagy with systemic lupus erythematosus in a chinese population. *Ann. Rheum. Dis.* 2011, *70*, 1330–1337.
- Yang, W.; Tang, H.; Zhang, Y.; Tang, X.; Zhang, J.; Sun, L.; Yang, J.; Cui, Y.; Zhang, L.; Hirankarn, N.; *et al.* Meta-analysis followed by replication identifies loci in or near CDKN1B, TET3, CD80, DRAM1, and ARID5B as associated with systemic lupus erythematosus in asians. *Am. J. Hum. Genet.* 2013, *92*, 41–51.
- Freedman, B.I.; Langefeld, C.D.; Andringa, K.K.; Croker, J.A.; Williams, A.H.; Garner, N.E.; Birmingham, D.J.; Hebert, L.A.; Hicks, P.J.; Segal, M.S.; *et al.* End-stage renal disease in african americans with lupus nephritis is associated with APOL1. *Arthritis Rheumatol.* 2014, *66*, 390–396.
- Zhou, X.J.; Nath, S.K.; Qi, Y.Y.; Cheng, F.J.; Yang, H.Z.; Zhang, Y.; Yang, W.; Ma, J.Y.; Zhao, M.H.; Shen, N.; *et al.* Brief report: Identification of *MTMR3* as a novel susceptibility gene for lupus nephritis in northern han chinese by shared-gene analysis with IgA nephropathy. *Arthritis Rheumatol.* 2014, 66, 2842–2848.
- Wan, G.; Zhaorigetu, S.; Liu, Z.; Kaini, R.; Jiang, Z.; Hu, C.A. Apolipoprotein L1, a novel Bcl-2 homology domain 3-only lipid-binding protein, induces autophagic cell death. *J. Biol. Chem.* 2008, 283, 21540–21549.
- Larsen, C.P.; Beggs, M.L.; Saeed, M.; Walker, P.D. Apolipoprotein 11 risk variants associate with systemic lupus erythematosus-associated collapsing glomerulopathy. J. Am. Soc. Nephrol. 2013, 24, 722–725.
- Kopp, J.B.; Winkler, C.A.; Zhao, X.; Radeva, M.K.; Gassman, J.J.; D'Agati, V.D.; Nast, C.C.; Wei, C.; Reiser, J.; Guay-Woodford, L.M.; *et al.* Clinical features and histology of apolipoprotein L1-associated nephropathy in the FSGS clinical trial. *J. Am. Soc. Nephrol.* 2015, *26*, 1443–1448.

- 34. Larsen, C.P.; Freedman, B.I. Apolipoprotein L1-associated nephropathy and the future of renal diagnostics. *J. Am. Soc. Nephrol.* **2015**, *26*, 1232–1235.
- 35. Vergne, I.; Deretic, V. The role of PI3P phosphatases in the regulation of autophagy. *FEBS Lett.* **2010**, *584*, 1313–1318.
- Taguchi-Atarashi, N.; Hamasaki, M.; Matsunaga, K.; Omori, H.; Ktistakis, N.T.; Yoshimori, T.; Noda, T. Modulation of local PtdIns3P levels by the PI phosphatase MTMR3 regulates constitutive autophagy. *Traffic* 2010, *11*, 468–478.
- 37. Tsokos, G.C. Systemic lupus erythematosus. N. Engl. J. Med. 2011, 365, 2110-2121.
- 38. Bijl, M.; Kallenberg, C.G. Ultraviolet light and cutaneous lupus. Lupus 2006, 15, 724–727.
- 39. Draborg, A.H.; Duus, K.; Houen, G. Epstein-barr virus and systemic lupus erythematosus. *Clin. Dev. Immunol.* **2012**, *2012*, 370516.
- 40. Evans, A.S.; Rothfield, N.F.; Niederman, J.C. Raised antibody titres to EB virus in systemic lupus erythematosus. *Lancet* **1971**, *1*, 167–168.
- 41. Simard, J.F.; Costenbader, K.H.; Liang, M.H.; Karlson, E.W.; Mittleman, M.A. Exposure to maternal smoking and incident SLE in a prospective cohort study. *Lupus* **2009**, *18*, 431–435.
- 42. Schmidt, E.; Tony, H.P.; Brocker, E.B.; Kneitz, C. Sun-induced life-threatening Lupus Nephritis. *Ann. N. Y. Acad. Sci.* **2007**, *1108*, 35–40.
- Kemp, M.G.; Lindsey-Boltz, L.A.; Sancar, A. UV light potentiates STING (stimulator of interferon genes)-dependent innate immune signaling through deregulation of ULK1 (Unc51-like kinase 1). *J. Biol. Chem.* 2015, 290, 12184–12194.
- 44. Poole, B.D.; Templeton, A.K.; Guthridge, J.M.; Brown, E.J.; Harley, J.B.; James, J.A. Aberrant epstein-barr viral infection in systemic lupus erythematosus. *Autoimmun. Rev.* **2009**, *8*, 337–342.
- 45. James, J.A.; Neas, B.R.; Moser, K.L.; Hall, T.; Bruner, G.R.; Sestak, A.L.; Harley, J.B. Systemic lupus erythematosus in adults is associated with previous epstein-barr virus exposure. *Arthritis Rheum.* **2001**, *44*, 1122–1126.
- 46. Harley, J.B.; James, J.A. Epstein-barr virus infection may be an environmental risk factor for systemic lupus erythematosus in children and teenagers. *Arthritis Rheum.* **1999**, *42*, 1782–1783.
- McClain, M.T.; Heinlen, L.D.; Dennis, G.J.; Roebuck, J.; Harley, J.B.; James, J.A. Early events in lupus humoral autoimmunity suggest initiation through molecular mimicry. *Nat. Med.* 2005, *11*, 85–89.
- 48. Yu, X.X.; Yao, C.W.; Tao, J.L.; Yang, C.; Luo, M.N.; Li, S.M.; Liu, H.F. The expression of renal epstein-barr virus markers in patients with lupus nephritis. *Exp. Ther. Med.* **2014**, *7*, 1135–1140.
- 49. Ding, Y.; He, X.; Liao, W.; Yi, Z.; Yang, H.; Xiang, W. The expression of EBV-encoded LMP1 in young patients with lupus nephritis. *Int. J. Clin. Exp. Med.* **2015**, *8*, 6073–6078.
- 50. Lee, D.Y.; Sugden, B. The latent membrane protein 1 oncogene modifies B-cell physiology by regulating autophagy. *Oncogene* **2008**, *27*, 2833–2842.
- Leung, C.S.; Haigh, T.A.; Mackay, L.K.; Rickinson, A.B.; Taylor, G.S. Nuclear location of an endogenously expressed antigen, EBNA1, restricts access to macroautophagy and the range of CD4 epitope display. *Proc. Natl. Acad. Sci. USA* 2010, *107*, 2165–2170.
- 52. Clarke, A.J.; Ellinghaus, U.; Cortini, A.; Stranks, A.; Simon, A.K.; Botto, M.; Vyse, T.J. Autophagy is activated in systemic lupus erythematosus and required for plasmablast development. *Ann. Rheum. Dis.* **2015**, *74*, 912–920.

- Pengo, N.; Scolari, M.; Oliva, L.; Milan, E.; Mainoldi, F.; Raimondi, A.; Fagioli, C.; Merlini, A.; Mariani, E.; Pasqualetto, E.; *et al.* Plasma cells require autophagy for sustainable immunoglobulin production. *Nat. Immunol.* 2013, 14, 298–305.
- 54. Conway, K.L.; Kuballa, P.; Khor, B.; Zhang, M.; Shi, H.N.; Virgin, H.W.; Xavier, R.J. Atg5 regulates plasma cell differentiation. *Autophagy* **2013**, *9*, 528–537.
- Gros, F.; Arnold, J.; Page, N.; Decossas, M.; Korganow, A.S.; Martin, T.; Muller, S. Macroautophagy is deregulated in murine and human lupus T lymphocytes. *Autophagy* 2012, *8*, 1113–1123.
- 56. Bosch, X. Systemic lupus erythematosus and the neutrophil. N. Engl. J. Med. 2011, 365, 758-760.
- Hakkim, A.; Furnrohr, B.G.; Amann, K.; Laube, B.; Abed, U.A.; Brinkmann, V.; Herrmann, M.; Voll, R.E.; Zychlinsky, A. Impairment of neutrophil extracellular trap degradation is associated with lupus nephritis. *Proc. Natl. Acad. Sci. USA* 2010, *107*, 9813–9818.
- Martinez-Valle, F.; Balada, E.; Ordi-Ros, J.; Bujan-Rivas, S.; Sellas-Fernandez, A.; Vilardell-Tarres, M. Dnase 1 activity in patients with systemic lupus erythematosus: Relationship with epidemiological, clinical, immunological and therapeutical features. *Lupus* 2009, *18*, 418–423.
- Leffler, J.; Martin, M.; Gullstrand, B.; Tyden, H.; Lood, C.; Truedsson, L.; Bengtsson, A.A.; Blom, A.M. Neutrophil extracellular traps that are not degraded in systemic lupus erythematosus activate complement exacerbating the disease. *J. Immunol.* 2012, *188*, 3522–3531.
- 60. Itakura, A.; McCarty, O.J. Pivotal role for the mtor pathway in the formation of neutrophil extracellular traps via regulation of autophagy. *Am. J. Physiol. Cell Physiol.* **2013**, *305*, C348–C354.
- 61. Li, Y.; Lee, P.Y.; Reeves, W.H. Monocyte and macrophage abnormalities in systemic lupus erythematosus. *Arch. Immunol. Ther. Exp.* **2010**, *58*, 355–364.
- 62. Li, B.; Yue, Y.; Dong, C.; Shi, Y.; Xiong, S. Blockade of macrophage autophagy ameliorates activated lymphocytes-derived DNA induced murine lupus possibly via inhibition of proinflammatory cytokine production. *Clin. Exp. Rheumatol.* **2014**, *32*, 705–714.
- 63. Kang, Y.L.; Saleem, M.A.; Chan, K.W.; Yung, B.Y.; Law, H.K. Trehalose, an mtor independent autophagy inducer, alleviates human podocyte injury after puromycin aminonucleoside treatment. *PLoS ONE* **2014**, *9*, e113520.
- 64. Hartleben, B.; Godel, M.; Meyer-Schwesinger, C.; Liu, S.; Ulrich, T.; Kobler, S.; Wiech, T.; Grahammer, F.; Arnold, S.J.; Lindenmeyer, M.T.; *et al.* Autophagy influences glomerular disease susceptibility and maintains podocyte homeostasis in aging mice. *J. Clin. Investig.* **2010**, *120*, 1084–1096.
- Lenoir, O.; Jasiek, M.; Henique, C.; Guyonnet, L.; Hartleben, B.; Bork, T.; Chipont, A.; Flosseau, K.; Bensaada, I.; Schmitt, A.; *et al.* Endothelial cell and podocyte autophagy synergistically protect from diabetes-induced glomerulosclerosis. *Autophagy* 2015, doi:10.1080/15548627.2015.1049799.
- Anja Hillmann, H.W.; Pap, T.; Jacobi, A. Uptake of sle autoantibodies by podocytes. *Ann. Rheum. Dis.* 2012, *71*, A1–A93.
- 67. Chan, T.M. Treatment of severe lupus nephritis: The new horizon. *Nat. Rev. Nephrol.* 2015, *11*, 46–61.
- Harr, M.W.; McColl, K.S.; Zhong, F.; Molitoris, J.K.; Distelhorst, C.W. Glucocorticoids downregulate Fyn and inhibit IP₃-mediated calcium signaling to promote autophagy in T lymphocytes. *Autophagy* 2010, *6*, 912–921.

- 69. Ben-Zvi, I.; Kivity, S.; Langevitz, P.; Shoenfeld, Y. Hydroxychloroquine: From malaria to autoimmunity. *Clin. Rev. Allergy Immunol.* **2012**, *42*, 145–153.
- 70. Perl, A. mTOR activation is a biomarker and a central pathway to autoimmune disorders, cancer, obesity, and aging. *Ann. N. Y. Acad. Sci.* **2015**, *1346*, 33–44.
- Reddy, P.S.; Legault, H.M.; Sypek, J.P.; Collins, M.J.; Goad, E.; Goldman, S.J.; Liu, W.; Murray, S.; Dorner, A.J.; O'Toole, M. Mapping similarities in mtor pathway perturbations in mouse lupus nephritis models and human lupus nephritis. *Arthritis Res. Ther.* 2008, 10, R127.
- Lui, S.L.; Tsang, R.; Chan, K.W.; Zhang, F.; Tam, S.; Yung, S.; Chan, T.M. Rapamycin attenuates the severity of established nephritis in lupus-prone NZB/W F1 mice. *Nephrol. Dial. Transplant.* 2008, 23, 2768–2776.
- 73. Yap, D.Y.; Ma, M.K.; Tang, C.S.; Chan, T.M. Proliferation signal inhibitors in the treatment of lupus nephritis: Preliminary experience. *Nephrology* **2012**, *17*, 676–680.
- Stylianou, K.; Petrakis, I.; Mavroeidi, V.; Stratakis, S.; Vardaki, E.; Perakis, K.; Stratigis, S.; Passam, A.; Papadogiorgaki, E.; Giannakakis, K.; *et al.* The PI3K/Akt/mTOR pathway is activated in murine lupus nephritis and downregulated by rapamycin. *Nephrol. Dial. Transplant.* 2011, 26, 498–508.
- 75. Marti, H.P.; Frey, F.J. Nephrotoxicity of rapamycin: An emerging problem in clinical medicine. *Nephrol. Dial. Transplant.* **2005**, *20*, 13–15.
- 76. Zhang, Z.; Wu, X.; Duan, J.; Hinrichs, D.; Wegmann, K.; Zhang, G.L.; Hall, M.; Rosenbaum, J.T. Low dose rapamycin exacerbates autoimmune experimental uveitis. *PLoS ONE* **2012**, *7*, e36589.
- 77. Schall, N.; Muller, S. Resetting the autoreactive immune system with a therapeutic peptide in lupus. *Lupus* **2015**, *24*, 412–418.
- Monneaux, F.; Lozano, J.M.; Patarroyo, M.E.; Briand, J.P.; Muller, S. T cell recognition and therapeutic effect of a phosphorylated synthetic peptide of the 70K snRNP protein administered in MRL/Ipr mice. *Eur. J. Immunol.* 2003, *33*, 287–296.
- 79. Page, N.; Gros, F.; Schall, N.; Decossas, M.; Bagnard, D.; Briand, J.P.; Muller, S. Hsc70 blockade by the therapeutic peptide P140 affects autophagic processes and endogenous MHCII presentation in murine lupus. *Ann. Rheum. Dis.* **2011**, *70*, 837–843.

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