

32 becoming a competitive soil remediation technology because of its cost-effective and
33 environmentally sound characteristics compared with other conventional technologies
34 (Huang *et al.*, 1997; Garbisu and Alkorta, 2001). Plant biomass and metal
35 concentrations in the harvestable parts of plants are the two determining factors for
36 the success of this technique. To achieve higher biomass, some fast-growing crops
37 such as corn, mustard and vetiver grass have been proposed for this process (Chen *et*
38 *al.*, 2004a; Luo *et al.*, 2005). Metal solubility in soils can be improved by adding
39 some chelating chemicals such as EDTA (ethylenediaminetetraacetic acid), EGTA
40 [ethyleneglycol -bis (β -aminoethyl ether), N, N, N', N-tetraacetic acid], CDTA (trans
41 -1, 2 -diaminocyclohexane -N, N, N', N'-tetraacetic acid) and HEDTA
42 (N-hydroxyethylenediaminetriacetic acid) (Blaylock *et al.*, 1997; Cooper *et al.*, 1999;
43 Wu *et al.*, 1999; Shen *et al.*, 2002.).

44 Although EDTA has shown high efficiency in enhancing metal solubility and
45 facilitating metal uptake in plants, the toxicity to plants and microorganisms, and the
46 potential risk of leached metals associated with the application of EDTA because of its
47 low biodegradability has excluded it as a good choice for use in practical applications
48 (Bucheli-Witschel and Egli, 2001; Grčman *et al.*, 2003). Some easily biodegradable
49 chelates such as NTA (nitrilotriacetate), EDDS (S,S-ethylenediaminedisuccinic acid),
50 and organic acids such as citric acid have been tested for this purpose (Luo *et al.*,
51 2005; Meer *et al.*, 2005; Quartacci *et al.*, 2005; Luo *et al.*, 2006a).

52 It was reported that NTA can be degraded as fast as glucose and citric acid in soils,
53 and can also rapidly be biodegraded under anaerobic conditions (Tiedje and Mason,

54 1974). Bucheli-Witschel and Egli (2001) observed a half-life of 2 to 7 d for NTA in
55 sediments. In the last few years, the use of NTA has been proposed to enhance the
56 uptake of heavy metals in soil phytoextraction (Kulli *et al.*, 1999; Kayser *et al.*, 2000).

57 Metal chelate complexes may enter plant tissues through breaks in the root
58 endodermis and Casparian strips, and be rapidly transported to the shoots (Römheld
59 and Marschner, 1981; Bell *et al.*, 1991). Our previous study showed some physical
60 damage to the roots caused by hot water treatment, or the addition of acid can
61 facilitate the uptake of metals by plants (Luo *et al.*, 2006b). The aims of this present
62 study were: (i) to investigate whether soil amendments with biodegradable NTA and
63 citric acid, in comparison to EDTA, added in hot solutions can further enhance the
64 uptake of heavy metals by plants from metal-contaminated soils; and (ii) to further
65 study, using hydroponic experiments, the possible mechanisms involved in
66 NTA-induced metal accumulation in plants.

67

68 **2. Materials and Methods**

69 **2.1. SOIL PREPARATION**

70

71 Soil samples were collected from a disused agricultural field in the Yuen Long area of
72 Hong Kong. The samples were sieved to pass through a 2 mm sieve and air-dried for
73 one week. The soils were artificially contaminated with Cu (400 mg kg⁻¹ of soil) in
74 the form of CuCO₃ (copper carbonate); Pb (500 mg kg⁻¹ of soil) as Pb₃(OH)₂(CO₃)₂
75 (lead hydroxide carbonate) and PbS (lead sulfide – galena, a common lead mineral in

76 mining areas) at a Pb concentration ratio of 1:1; Zn (500 mg kg⁻¹ of soil) in the form
77 of ZnCO₃ (zinc carbonate) and ZnS (zinc sulfide) at a Zn concentration ratio of 1:1;
78 and Cd (15 mg kg⁻¹ of soil) in the form of Cd(NO₃)₂·4H₂O (cadmium nitrate). The
79 basal fertilizers applied to the soil were 80 mg P kg⁻¹ of dry soil, and 100 mg K kg⁻¹ of
80 dry soil as KH₂PO₄ (Shen *et al.*, 2002). After the addition of heavy metals, the soils
81 were equilibrated for two months, undergoing seven cycles of saturation with
82 deionized water and air-drying processes.

83 The electrical conductivity (EC) of the soil was measured using a conductivity
84 meter on the soil extract, obtained by shaking soil with double-distilled water at a
85 water-to-soil ration of 1:2 (w/v). The soil pH was measured by 0.01 mol of CaCl₂ at a
86 1:5 ratio (w/v) using a pH meter. The cation exchangeable capacity (CEC) of the soil
87 was determined using the ammonium acetate saturation method. The soil texture,
88 organic matter content, total N and field water capacity were measured by the
89 procedures described by Avery and Bascomb (1982). The total metal concentrations
90 were determined by ICP-AES (Perkin-Elmer Optima 3300 DV) after strong acid
91 digestion (4:1 concentrated HNO₃ and HClO₄ (v/v)) (Li *et al.*, 2001). The selected
92 physical and chemical properties of the soil used in the present study are presented in
93 Table 1.

94

95 2.2. HOT EDTA, CITRIC ACID AND NTA TREATMENTS

96

97 Air-dried soils (500 g) were placed in plastic pots (12 cm i.d. x 12 cm in height). The

98 moisture of the soil was maintained to near field water capacity by adding deionized
99 water (DIW) on a daily basis. Seeds of beans (*P vulgaris* L. white bean) were sown
100 directly in the soils. After germination, the seedlings were thinned to four plants per
101 pot. On the 21st day after the beans were sown, EDTA, citric acid and NTA were
102 applied in the form of 100 ml Na₂EDTA, citric acid and Na₃NTA solutions to the
103 surface of the soils in two different ways (as heated and non-heated solutions) at rates
104 of 0 (control), 1.0, 3.0 and 5.0 mmol kg⁻¹ of soil. The three chemicals were all from
105 BDH Laboratory Supplies (Poole U.K.), with a minimum assay of above 99.5%. The
106 hot solution treatments were conducted by adding boiled solution to the soil in the
107 pots, with the final temperature of the soils being about 40 °C. Three replicates were
108 conducted for each treatment. All of the experiments were operated in a greenhouse
109 under natural light. Air temperatures ranged from 16 to 21 °C. All of the plants were
110 harvested 7 d after the treatment by cutting the shoots 0.5 cm above the surface of the
111 soil. The shoots were washed with tap water, rinsed with DIW, and dried at 70 °C in a
112 drying oven to a constant weight for dry weight measurements. The dried plant
113 materials were ground using an agate mill.

114

115 2.3. EXTRACTING METALS WITH DIFFERENT NTA SOLUTIONS

116

117 About 4.0 g of soil (based on dry weight) were placed in a 50-mL polypropylene
118 centrifuge tube. NTA solutions at rates of 0, 1, 3 and 5 mmol kg⁻¹ of soil were added
119 to the soil (at a soil-to-water ratio of 1:5 (w/v)) at different temperatures of 25 °C

120 (control), 40 °C, 60 °C and 80 °C, respectively. Every treatment was replicated three
121 times. The suspension was shaken for 30 min. After the centrifugation, the
122 supernatant was filtered through a 0.45 µm paper filter (Whatman [Maidstone, UK]
123 42), acidified with HNO₃ and analyzed for metal concentrations by ICP-AES (Perkin
124 Elmer 3000DV).

125

126 2.4. ROOT PRETREATMENT WITH HOT WATER

127

128 Seeds of beans (*P vulgaris* L. white bean) were sterilized in 0.1% (w / v) HgCl₂ for 10
129 min, and rinsed four times in deionized water before being placed on filter paper for
130 germination. After germination, the plants of the same size were selected and
131 transferred to 2-L polyethylene vessels containing a modified 0.2-strength Rorison's
132 nutrient solution (Hewitt, 1966) with the following composition (in µmol L⁻¹): 400
133 Ca(NO₃)₂, 200 Mg(SO₄)₂, 50 K₂HPO₄, 300 KCl, 9.2 H₃BO₃, 1.8 MnSO₄·4H₂O, 0.21
134 Na₂MoO₄·2H₂O, 0.31 CuSO₄·5H₂O, 10 ZnSO₄·7H₂O and 10.8 Fe-EDTA at pH 6.0.
135 Nutrient solutions were aerated continuously and renewed every two days. The plants
136 were grown in a greenhouse where the temperature ranged from 17 °C to 22 °C.

137 After 7 d of the transplanting, different pretreatments were conducted to assess the
138 effects of roots damaged by hot water on the accumulation of Cu in shoots. Five
139 pretreatments were included: the roots were exposed in hot water at 30 °C, 40 °C, 50
140 °C, 60 °C and 80 °C for 15 min. Plants that were not subjected to pretreatment in hot
141 water (that were placed in a room where the temperature was about 20 °C) were used

142 as the control. After pretreatments, eighteen plants from every treatment were used to
143 assess the membrane permeability of the roots (the relative electrolytic leakage) by
144 measuring the electrical conductivity (Zhu *et al.*, 1990; Zhou and Leul, 1998). The
145 root samples (0.5 g) were placed in a test tube containing 15 ml of deionized water,
146 and the root tissue was immersed and vibrated at room temperature for 2 h. The
147 conductivity of the solution was measured using a conductivity meter (DDS - 11A).
148 After boiling the samples for 10 min, the conductivity was measured again when the
149 solution had cooled to room temperature. The relative electrical conductivity (REC)
150 was calculated as follows: $REC = C_1 / C_2 \times 100$, where C_1 and C_2 were the
151 electrolyte conductivities measured before and after boiling, respectively. The
152 remaining eighteen plants from every treatment group were exposed to $500 \mu\text{mol L}^{-1}$
153 of Cu + $500 \mu\text{mol L}^{-1}$ of NTA solutions for 2 d (pH = 6.0). Cu and NTA were added in
154 the forms of $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$, Na_3NTA solutions, respectively. Every treatment was
155 replicated three times. At the end of these experiments, the shoots and roots were
156 harvested for further chemical analysis. The effects of root damage on the
157 accumulations of Pb, Zn and Cd were studied in the same way, whereby Pb, Zn and Cd
158 were applied in the forms of $\text{Pb}(\text{NO}_3)_2$, $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$ and $\text{CdNO}_3 \cdot 4\text{H}_2\text{O}$ solutions,
159 respectively.

160

161 2.5. PLANT AND SOIL ANALYSIS

162

163 Subsamples of ground shoot samples (200 mg) were digested in a mixture of

164 concentrated HNO₃ and HClO₄ (4:1, by volume), and the major and trace elements in
165 the solutions were determined with ICP-AES (Chen *et al.*, 2004b). Certified standard
166 reference material (SRM 1515, apple leaves) from the National Institute of Standards
167 and Technology, U.S.A., was used in the digestion and analysis as part of the QA/QC
168 protocol. Reagent blank and analytical duplicates were also used where appropriate to
169 ensure accuracy and precision in the analysis. The recovery rates were around 90 ±
170 5% for all of the metals in the plant reference material. The data reported in this paper
171 were the mean values based on the results of the three replicated experiments.
172 Statistical analyses of the experimental data, such as correlation and significant
173 differences, were performed using SPSS® 11.0 statistical software.

174

175 **3. Results**

176

177 **3.1. PLANT GROWTH**

178

179 The dry mass yields of beans are shown in Fig. 1. When the three chelates were added
180 in normal solutions, EDTA prohibited plant growth the most, followed by the NTA
181 application. The depressed effects on plant growth increased with the dosage of
182 chelate. The application of citric acid did not have any significant effect on the growth
183 of the plants ($P < 0.05$). When the chelates were added in hot solutions, a significant
184 decrease in the dry biomass yields was observed in all the plants, in comparison with
185 the non-heated treatments ($P < 0.05$). On average, on the 7th day after the application

186 of the chelates, the dry biomass of the shoots decreased by 18%, 19% and 15% in the
187 hot EDTA, citric acid and NTA applications, respectively, in comparison with the
188 results observed in the treatments using a normal chelate solution.

189

190 3.2. METAL CONCENTRATIONS AND PHYTOEXTRACTIONS IN HOT EDTA, 191 CITRIC ACID AND NTA TREATMENTS

192

193 Compared with the control group, there was no significant change in the
194 concentrations of heavy metals in the shoots after the addition of citric acid (Fig. 2).

195 In contrast to this, the application of EDTA and NTA to the soil significantly increased
196 the concentrations of Cu, Pb, Zn and Cd in the shoots. Generally, NTA was
197 comparable to EDTA in increasing the concentrations of Cu, Zn and Cd in shoots. For
198 Pb, NTA was significantly less effective than EDTA, although an enhanced uptake
199 was also observed.

200 When the chelates were applied as hot solutions to the soils, the concentrations of
201 metals in the shoots of beans increased greatly in comparison with the chelates
202 applied in normal solutions at the same dosage (Fig. 2). The concentrations of Cu
203 ranged from 878 to 1460, 60 to 128, and 540 to 1250 mg kg⁻¹ in the shoots of beans
204 treated with hot EDTA, citric acid and NTA, respectively. These were 5.1 - 8.3, 3.2 -
205 7.2 and 4.7 - 7.3 times the concentrations of those with the normal chelates treatments
206 without heating, and 26.5 - 44, 1.8 - 3.9 and 16.3 - 37.8 times that in the control group
207 (with the application of hot water only), respectively. The highest Pb concentration of

208 998 mg kg⁻¹ was found in the shoots of beans treated with hot EDTA at the rate of 5
209 mmol kg⁻¹. The average enhanced effects of hot EDTA, citric acid and NTA on the Pb
210 shoot uptake were 12.9, 3.5 and 10.1 times greater than those in the corresponding
211 chelate treatments without heating. With regard to Zn and Cd, when EDTA, citric acid
212 and NTA were applied at rates of 1 - 5 mmol kg⁻¹, the concentrations of Zn and Cd in
213 the shoots of the beans were about 8.2 and 21 times those of the controls.

214 Results on the total metal phytoextraction by the shoots of beans are shown in
215 Table 2. The maximum phytoextraction of Cu, Zn and Cd was found in the heated
216 NTA treatment, which increased 47-, 6.5- and 18- fold, respectively, compared with
217 the control group (to which normal water was added). With regard to Pb, the plants
218 treated with 5 mmol kg⁻¹ of hot EDTA attained the maximum level of phytoextraction
219 of approximately 136-fold that of the corresponding control group.

220

221 3.3. METAL DISSOLUTION STUDY WITH THE ADDITION OF HOT NTA

222

223 In order to examine the effects of temperature on metal solubility, soil was extracted
224 with NTA solutions at different temperatures (see Fig. 3). The concentrations of
225 water-soluble metals in soil were mainly dependent upon the chelate application
226 dosage. The soluble metal concentrations increased as levels of NTA applied to the
227 soil increased. At the same application dosage, no significant differences were
228 observed in the concentrations of soluble metals between the treatments with hot NTA
229 solutions and those with normal NTA solutions (Fig. 3).

230

231 3.4. EFFECTS OF PRETREATMENT WITH HOT WATER ON THE
232 ACCUMULATION OF CU IN BEANS

233

234 The roots of beans were pretreated with hot water at different temperatures before they
235 were exposed in solutions containing $500 \mu\text{mol L}^{-1}$ of Cu + $500 \mu\text{mol L}^{-1}$ of NTA. Two
236 days after the exposure, the concentrations of Cu in shoots were measured (Fig. 4).

237 The results showed that there was a significantly positive correlation between the Cu
238 concentration in shoots and the relative electrolyte leakage rate of root cells ($R^2 = 0.93$,
239 $n = 18$). Similar significantly positive correlation results with R^2 0.90, 0.84 and 0.89
240 were also obtained for Pb, Zn and Cd, respectively ($n = 18$).

241

242 **4. Discussion**

243

244 Chelate-enhanced phytoextraction has been proposed as an effective technology for
245 the cleaning up of metal-contaminated soils (Huang *et al.*, 1997; Wu *et al.*, 1999, Luo
246 *et al.*, 2005). The present study showed that by adding an easily biodegradable chelate
247 of NTA in a hot solution to the soil, the concentrations of metals, specifically Cu, Pb,
248 Zn and Cd, in the shoots of beans increased by 55, 45, 7.4 and 19-fold respectively
249 compared to those of the control group (to which normal water had been applied). The
250 total levels of metal phytoextraction in the shoots of the plant were also enhanced to
251 25, 20, 5.4 and 10 times those seen in the control group, despite a drop in the

252 production of biomass. The enhancement in metal uptake after the application of hot
253 NTA solution achieved in the present study was significantly higher than those
254 previous studies on NTA applications (Kulli *et al.*, 1999; Kayser *et al.*, 2000;
255 Robinson *et al.*, 2000; Wenger *et al.*, 2002; Meers *et al.*, 2004; Quartacci *et al.*, 2005).
256 In addition, the concentrations and metal phytoextraction of Cu, Pb, Zn and Cd in the
257 shoots of beans treated with 1 mmol kg⁻¹ of hot NTA were significantly higher than
258 those achieved in the treatment with normal NTA solution at a rate of 5 mmol kg⁻¹ soil.
259 The results indicate that for a given phytoremediation efficiency, the application
260 dosage of NTA can be reduced to 1/5 (20%) if the NTA is applied in a hot solution
261 instead of a normal solution. Thus, the application cost of this strategy can be greatly
262 reduced and the potential risk associated with the application of NTA of metals
263 leaching to the surrounding area can also be reduced accordingly. It should be noted
264 that in the current study, the plants were grown in the pots for only 28 days, which
265 differed greatly from the field conditions. First, the pot was a closed container, which
266 limited the added chelate to a small space. Second, the metal uptake capacity of plant
267 varied a lot at different growth stages. The young seedlings were usually more
268 sensitive to the chelate application than those at the mature age. Therefore, further
269 field experiments are essential to test this result before this technology can be adopted
270 on a large scale.

271 In soils, most heavy metals have low phytoavailability because they are usually
272 strongly associated with organic matter, Fe-Mn oxides, clays and precipitation as
273 carbonates, hydroxides and phosphates (McBride 1994). Once the chelate is applied

274 into soils, it will solubilize metals from the soils and transfer them to the roots, which
275 is the so-called stage I process (Ensley *et al.*, 1999). The stage II process involving the
276 enhanced transfer of the mobilized metals to the shoots will take place afterwards. As
277 shown in Fig. 3, compared with the addition of a normal NTA solution, the
278 application of NTA in a hot solution does not further improve the solubility of metals
279 from soils, although the application of NTA in a hot or normal solution caused the
280 metals to be much more soluble than was seen with the control group (to which NTA
281 had not been applied). The less pronounced effect of temperature on the extraction of
282 metals can be ascribed to the fact that the soils had been artificially contaminated. The
283 absence of the effects of “aging” minimized mass transfer limitations and therefore
284 the potential effect of temperature. Thus, the significantly higher metal uptake
285 achieved in the hot NTA treatment than in the normal NTA treatment may be
286 attributed to the possible enhanced transport of metal chelate complexes from soil
287 solution to root xylems, and then to their translocation from the roots to the shoots of
288 the plant with the transpiration stream.

289 A significantly positive correlation was found between the metal concentrations in
290 the shoots of the beans and the relative electrolyte leakage rate of the root cells (Fig. 4),
291 which meant that root damage could be helpful in the accumulation of metals in plant
292 shoots. This result was consistent with our previous studies, where pretreatments on
293 the roots of Indian mustard with MC (methanol-trichloromethane) solution, HCl and
294 hot water before a combined treatment of Pb and EDTA dramatically increased the
295 concentration of Pb in shoots compared with shoots that had not been pretreated (Luo

296 *et al.*, 2006b). The enhanced translocation of metals from roots to shoots because of
297 root damage can be explained by the breakdown of the root exclusion mechanism.
298 Bell *et al.* (1991) suggested that the plant uptake of metal chelate complexes occurs at
299 breaks in the root endodermis and Casparian strip. It is hypothesized that the hot
300 solution firstly destroyed the physiological barrier(s) of plant roots that normally
301 function to control the uptake and translocation of solutes. Then, the rapid
302 equilibration of the soil solution with the sap of the xylem was achieved. After
303 entering the xylem, metals would be translocated with the transpiration stream from
304 the roots to shoots of the plant, leading to a high concentration of metals in the shoots.
305 It has been reported that Pb can be absorbed and transferred as a Pb-EDTA complex in
306 the presence of high concentrations of EDTA (Vassil *et al.*, 1998; Epstein *et al.*, 1999;
307 Sarret *et al.*, 2001). In the process of hot NTA facilitated metal uptake, the metal might
308 be transported in the form of a metal-NTA complex through the apoplastic route, as
309 suggested by Wenger *et al.* (2003).

310 EDTA has been one of the most efficient chelating agents in increasing the uptake
311 of metals, especially Pb (Blaylock *et al.*, 1997; Huang *et al.*, 1997, Cooper *et al.*,
312 1999). The chelate of NTA is usually shown to have a lower efficiency in solubilizing
313 metals from soils and to be less effective in enhancing the uptake of metals by plants
314 than EDTA (Shen *et al.*, 2002; Meers *et al.*, 2004; Meers *et al.*, 2005). In our present
315 study, however, when NTA was applied in a normal solution, the concentrations and
316 total phytoextraction of Cu, Zn and Cd in the shoots of beans were comparable to that
317 achieved in the normal EDTA treatments at the same application dosage, although the

318 data for Pb was far lower than that in the treatment of EDTA (Fig. 2 and Table 2). The
319 higher metal uptake efficiency after the application of NTA may be attributed to
320 different experimental conditions such as soil properties, metal concentrations and
321 components in soil, and to chelate application dosages and methods. Chiu *et al.* (2005)
322 found that NTA was more effective than EDTA in extracting Zn and Cu within the
323 tested concentration of 20 mmol kg⁻¹ of soil. Tandy *et al.* (2004) also reported that at
324 pH 7, NTA showed higher extraction efficiency for Cu and Zn than EDTA.

325 Besides the screening of plant species, the selection of chelates and the
326 optimization of the chelate application strategy will be very useful for increasing the
327 uptake of metals by plants and reducing the potential risk to the surrounding
328 environment, such as the leaching of metals, in the process of chemical-induced
329 phytoremediation. It was reported that a split application of chelates is more effective
330 than the application of single dosages in increasing the phytoextraction of metals from
331 soils (Grčman *et al.*, 2001; Puschenreiter *et al.*, 2001; Shen *et al.*, 2002). Combining
332 EDTA / NTA and glyphosate increased the concentration of Pb in plant tissues when
333 glyphosate was added shortly before the plants were harvested (Ensley *et al.*, 1999
334 Kayser *et al.*, 1999). The combined application of EDTA and EDDS dramatically
335 improved the uptake of Pb by corn (Luo *et al.*, 2006c). Applying an electric field
336 around the plants in combination with the application of EDTA can also enhance the
337 uptake of Pb by Indian mustard compared with the addition of EDTA only (Lim *et al.*,
338 2004). Recently, a new slow-release chelating agent application was reported, where
339 solid EDTA was coated with a layer of silicate to slow down the mobilization of

340 metals in the soil in order to match their uptake by the plant, and thus prevent
341 excessive mobilization (Li *et al.*, 2005). The present study showed when NTA was
342 applied in hot solutions at the rate of 1 mmol kg⁻¹ of soil, the total metal uptakes of Cu,
343 Zn and Cd were higher than those achieved in the normal EDTA application at a
344 dosage of 5 mmol kg⁻¹ of soil (see Table 2). For Pb, although the total phytoextraction
345 observed at the treatment of 1 mmol kg⁻¹ of hot NTA was lower than that achieved by
346 the application of 5 mmol kg⁻¹ of normal EDTA, it was still higher than that of 1
347 mmol kg⁻¹ of normal EDTA (Table 2). This result indicates that if NTA were to be
348 applied in a hot solution, the efficiency in enhancing metal uptake could exceed that
349 of a normal EDTA treatment. In addition, the characteristic of easy biodegradability
350 makes NTA more suitable than EDTA for metal phytoremediation. Another
351 biodegradable chelate, EDDS, also showed high efficiency in metal phytoextraction,
352 particularly when it was added in heated solutions (Luo *et al.*, 2007). However, EDDS
353 is far more costly than NTA. Taking these factors into account, the application of a hot
354 NTA solution might be a better alternative for chelate-enhanced metal
355 phytoextraction.

356

357

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358

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488

489 Table 1
 490 The physicochemical properties of the soils used in the study
 491

pH (CaCl ₂)	7.12
Electrical conductivity at 25°C (μS cm ⁻¹)	262
Sand (%) > 0.05 mm	79.5
Silt (%) 0.05 - 0.001 mm	13
Clay (%) < 0.001 mm	7.5
N _{Total} (%)	0.15
Organic matter (%)	2.7
Cation exchange capacity (cmol kg ⁻¹)	4.2
Field water capacity (%)	39.7
Total metal concentration after amendment (mg kg ⁻¹)	
Cu	480
Pb	575
Zn	700
Cd	17

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493

494 Table 2

495 Total phytoextraction (mg kg⁻¹ soil) of Cu, Pb, Zn, and Cd in the shoots of beans 7 d
496 after the application of EDTA, citric acid (CA) and NTA at different concentrations
497 (mmol kg⁻¹ soil)

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Treatments	Cu	Pb	Zn	Cd
Water	70 ± 8.6a	8.6 ± 0.6a	482 ± 52a	6.6 ± 0.9a
Hot-water	110 ± 13a	13 ± 2.1a	468 ± 69a	8.8 ± 1.5a
1mM EDTA	359 ± 49ab	19 ± 2a	551 ± 80a	14.4 ± 2.8a
Hot-1mM EDTA	2600 ± 208cd	327 ± 40b	1910 ± 293c	77 ± 8b
3mMEDTA	622 ± 70b	126 ± 13ab	790 ± 54b	22.2 ± 3.6a
Hot-3mM EDTA	2630 ± 300cd	1140 ± 100	2420 ± 300c	105 ± 20c
5mM EDTA	700 ± 55b	358 ± 20b	926 ± 56b	28 ± 1a
Hot-5mM EDTA	3060 ± 245d	2100 ± 260c	2430 ± 315d	112 ± 19c
1mM CA	79 ± 8.2a	9.7 ± 0.5a	582 ± 70a	7.2 ± 5.5a
Hot-1mM CA	200 ± 24a	26 ± 3a	510 ± 65a	8.6 ± 0.7a
3mMCA	67 ± 7.5a	6.5 ± 0.8a	476 ± 35a	5.7 ± 0.6a
Hot-3mM CA	410 ± 56ab	19 ± 2.1a	586 ± 40a	10.2 ± 2.8a
5mM CA	67 ± 4.5a	8.4 ± 0.6a	493 ± 30a	6.2 ± 1.4a
Hot-5mM CA	385 ± 42ab	24 ± 3.5a	616 ± 75a	12.5 ± 2a
1mM NTA	427 ± 50ab	18.4 ± 2a	510 ± 80a	16 ± 2.5a
Hot-1mM NTA	1730 ± 150c	70 ± 5a	780 ± 64b	32.8 ± 4.8a
3mM NTA	585 ± 65b	29.3 ± 4.5a	586 ± 40a	17.4 ± 3a
Hot-3mM NTA	3310 ± 278d	340 ± 40b	777 ± 80b	117 ± 25c
5mM NTA	578 ± 25b	41 ± 6a	616 ± 50a	18.2 ± 3.8a
Hot-5mM NTA	3130 ± 480d	378 ± 48b	836 ± 86b	117 ± 25c

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500 The values are means ± S.D. (n = 3); in the vertical direction the different small letters
501 stand for statistical significance at the 0.05 level with the LSD test.

502

503 **Figure legends:**

504

505 Fig. 1. Effects of the application of EDTA, citric acid and NTA on the dry matter
506 yields of beans. The values are means \pm S.D. (n = 3).

507

508 Fig. 2. Effects of the application of chelates on the concentrations of Cu (a), Pb (b),
509 Zn (c), and Cd (d) in the shoots of beans. The values are means \pm S.D. (n = 3).

510

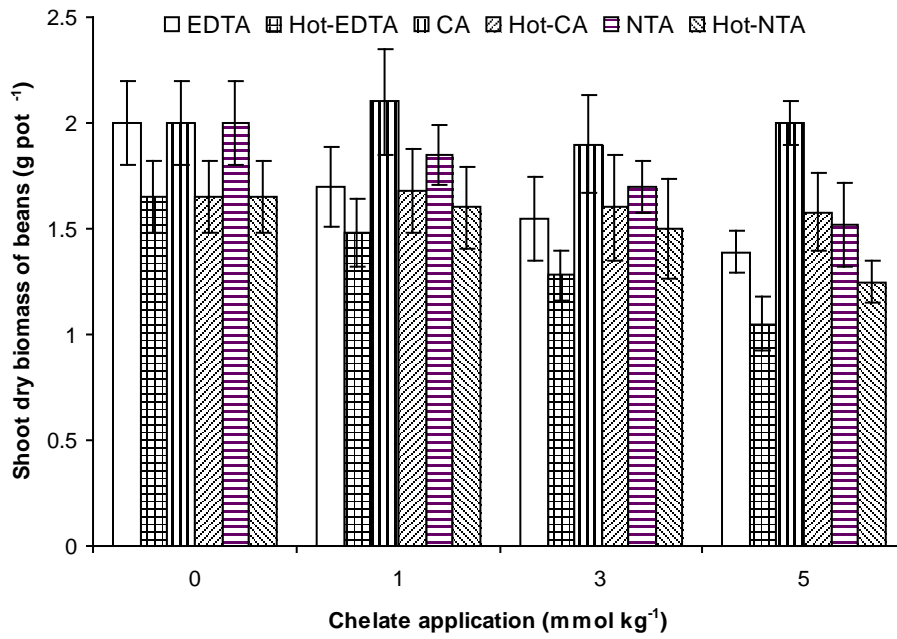
511 Fig. 3. Effects of the application of NTA (mmol kg^{-1} soil) at different temperatures on
512 the solubilization of Cu, Pb, Zn and Cd (mg kg^{-1} soil) in the soil. The values are
513 means \pm S.D. (n = 3).

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515

516 Fig. 4. The correlation between the relative electrolyte leakage of roots and the
517 concentration of Cu in the shoots of beans. Plants were pretreated with hot water at
518 different temperatures, then exposed in solutions containing $500 \mu\text{mol L}^{-1}$ of Cu + 500
519 $\mu\text{mol L}^{-1}$ of NTA for 2 d. The root cell electrolytic leakage (relative electrical
520 conductivity) was measured immediately after the pretreatment with hot water.

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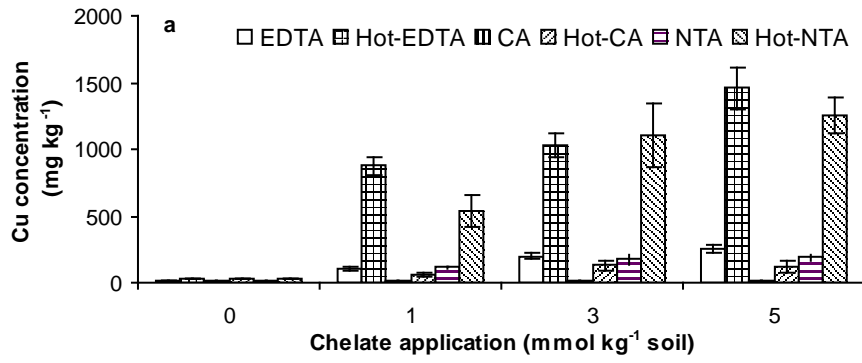


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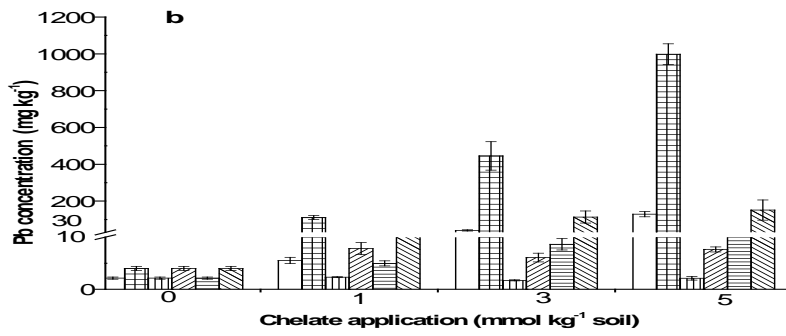
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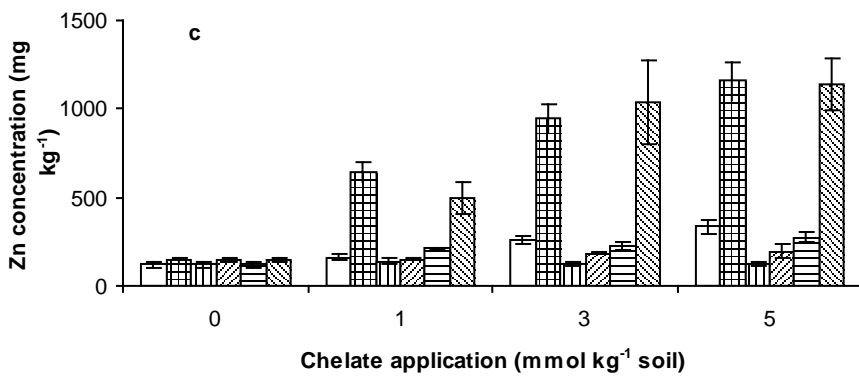
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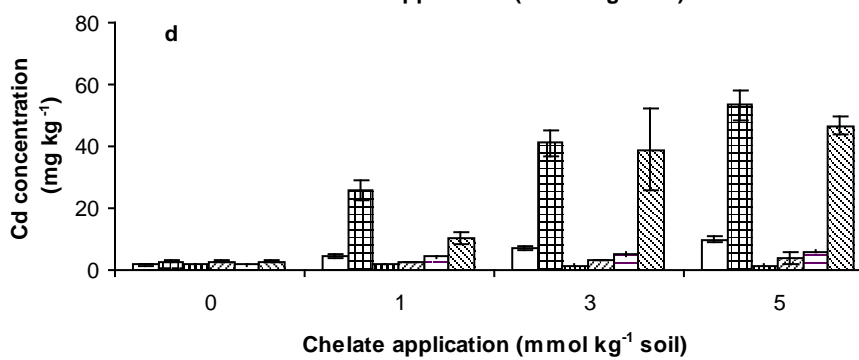
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Fig. 2 Effects of the application of chelates on the concentrations of Cu (a), Pb (b), Zn

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(c), and Cd (d) in the shoots of beans. The values are means \pm S.D. (n = 3).

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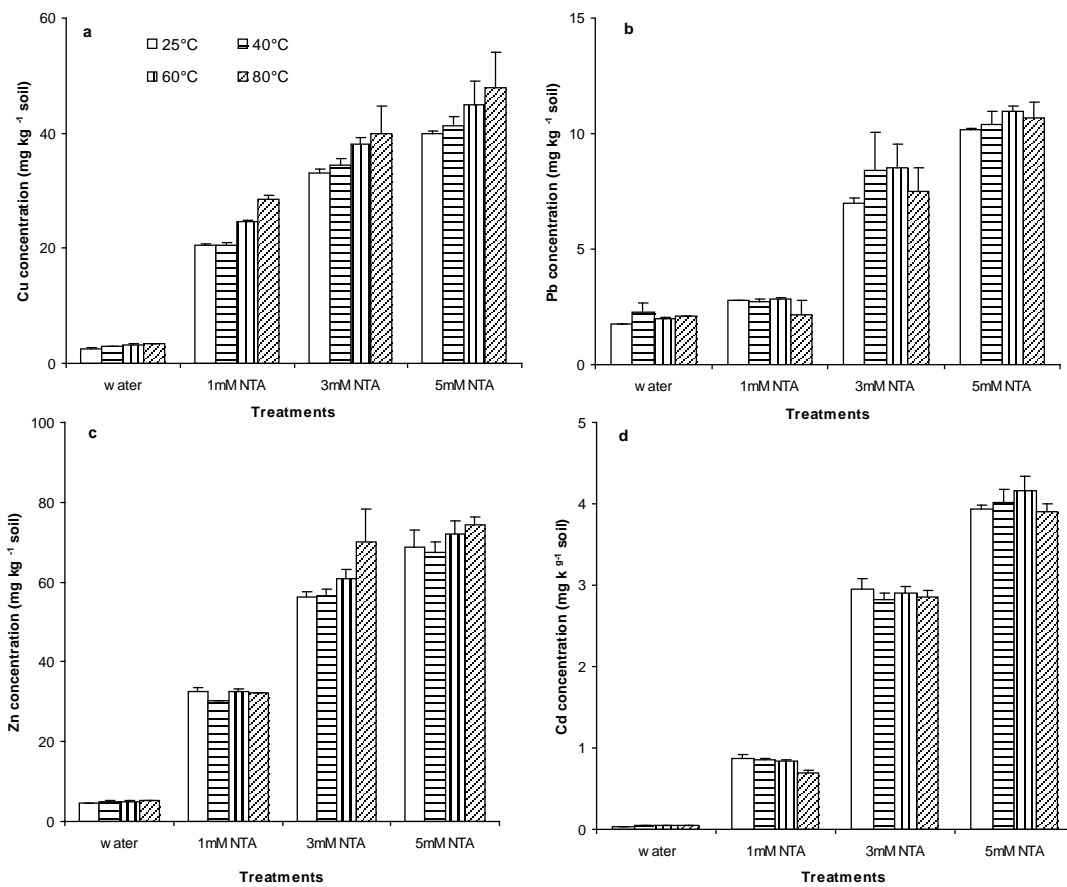
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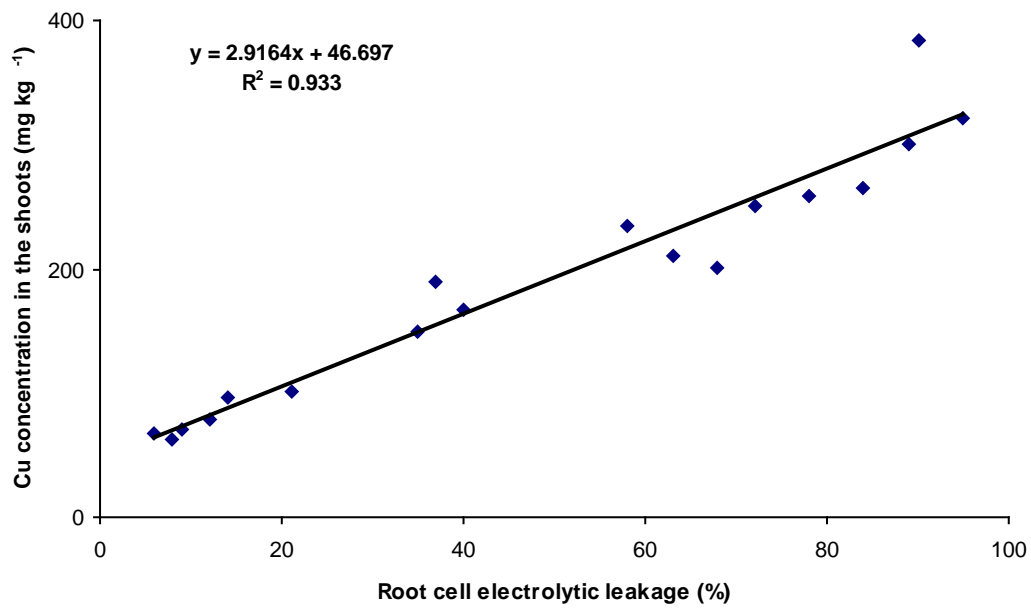
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574 Fig. 3. Effects of the application of NTA (mmol kg⁻¹ soil) at different temperatures on
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576 means ± S.D. (n = 3).

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 582 different temperatures, then exposed in solutions containing $500 \mu\text{mol L}^{-1}$ of Cu + 500
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