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5 **Effect of polysaccharide chain conformation on ultrasonic degradation of curdlan in**
6 **alkaline solution**

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22 **Abstract:** This study was to investigate the effects of polysaccharide chain conformation on
23 ultrasonic degradation of curdlan, a high MW β -glucan with wide applications. The ultrasonic
24 degradation was performed in alkaline solution at 0.1 M and 0.3 M NaOH, in which the curdlan
25 chain was mainly in triple helical conformation and random coil form, respectively. The
26 degradation rate was represented by the kinetic model, $1/M_t - 1/M_o = kt$, with the rate constant
27 k increasing with the ultrasonic power. The degradation rate was much higher in 0.3 M NaOH
28 than in 0.1 M NaOH, suggesting that curdlan in random coil conformation was more liable to
29 degradation than in helical conformation. Curdlan in 0.1 M NaOH was changed from triple
30 helices to single helices and eventually to random coils with a higher solubility. In summary,
31 ultrasonic degradation of curdlan in alkaline solution had a close and complex relationship to
32 the chain conformation changes.

33

34 **Keywords:** Curdlan; Ultrasonic degradation; Kinetics; Chain conformation; Intrinsic viscosity;
35 Molecular weight

36

37 **1. Introduction**

38 Curdlan is an exopolysaccharide produced by *Agrobacterium biovar*, *Alcaligenes faecalis*
39 and some other bacteria (McIntosh & Stanisich, 2005; Zhang & Edgar, 2014; Liu et al., 2015).
40 It is a water-insoluble linear β -1,3-glucan and soluble in alkaline condition. Curdlan has a high
41 molecular weight with an average degree of polymerization about 450 and up to 12,000
42 monosaccharide units (Futatsuyama et al., 1999). It has a major application in the food industry
43 because of the favorable rheological properties and various bioactivities (Zhang & Edgar,
44 2014). A useful and unique property of curdlan is its ability to form two distinct types of

45 thermo-gels in aqueous media, high-set gel and low-set gel network (Cai & Zhang, 2017). In
46 high-set gel, the polymer chain is predominately in triple helical conformation, while single
47 helix is predominant in low-set gel network. In an alkaline solution, the triple helix of curdlan
48 chain unwinds and conformational transition from helical structure to random coil occurs with
49 increasing NaOH concentration from 0.19 M to 0.24 M (Nakata et al., 1998). Chemical
50 modifications such as carboxymethylation, sulfation, phosphorylation, oxidation and
51 esterification have been applied to prepare curdlan derivatives with good water solubility as
52 well as bioactivity to widen its application (Cai & Zhang, 2017).

53 Power ultrasound (US) has been exploited as a simple and direct means for acceleration or
54 activation of a wide range of chemical and physical processes (Mason & Lorimer, 2002; Tao
55 & Sun, 2015). Depolymerization of high molecular weight polysaccharides is a useful approach
56 for generation of lower molecular weight fractions with lower viscosity, higher water solubility
57 and better functionality. Compared with other techniques for polysaccharide degradation,
58 including thermal, mechanical, oxidative, and hydrolytic and high-energy radiation, US is a
59 clean, non-hazardous, non-thermal mechanical means, which is most favorable for processing
60 food grade products (Gogate & Prajapat, 2015). Because ultrasonic degradation of
61 polysaccharide is mainly caused by the fluid shear forces from collapsing cavitation, it may
62 reduce the molecular weight by splitting the most susceptible chemical bonds, but not cause
63 significant change in chemical structure (Huang et al., 2015; Prajapat et al., 2016; Čížová et al.,
64 2015; Zhi et al., 2017).

65 The US degradation process and rate of polysaccharides can be affected by the structure
66 and properties of polysaccharides to be degraded. Although many previous studies have been

67 done on ultrasonic degradation of polysaccharides, few have addressed the effects of chain
68 conformation on the degradation process (Gogate & Prajapat, 2015; Tao & Sun, 2015). There
69 is still no reported study on the application of power US for modification or degradation of
70 natural curdlan. This study aims to investigate the effect of chain conformation on the US
71 degradation of curdlan in alkaline solution with different NaOH concentrations and to analyze
72 the degradation kinetics and mechanism.

73

74 **2. Materials and methods**

75 **2.1 Materials**

76 Curdlan powder from *Alcaligenes faecalis* was purchased from Sigma-Aldrich, USA. The
77 powder was stored at 4°C before use. Curdlan solution was prepared by dissolving the sample
78 powder in 0.1 M and 0.3 M sodium hydroxide (NaOH) solution for 2 hours at room temperature
79 with constant stirring. Aqueous solutions were prepared with ultrapure water from a Milli-Q
80 water purification system (Millipore, Bedford, MA, USA). Congo red was purchased from
81 Shanghai Yuan Ye Biotechnology, Ltd. (Shanghai, China). All other reagents were of analytical
82 grade unless otherwise specified.

83

84 **2.2 Ultrasonic treatment conditions of curdlan**

85 Ultrasonic treatment (degradation) of curdlan was performed with a Model VCX-130
86 ultrasonic processor of 20 kHz frequency and 130 W maximum power and a probe horn of 6
87 mm tip diameter (Soncis & Materials Inc., Newton, USA). The actual power at a set amplitude
88 was determined by the calorimetric method as described previously (Cheung & Wu, 2013). The

89 curdlan sample solution (25 mL and 2 g/L) was contained in a 50 ml plastic centrifuge bottle
90 and the ultrasound probe was immersed in the solution at a fixed depth of 2 cm. The sample
91 bottle was placed in an ice bath to maintain a relatively low temperature of $40 \pm 5^\circ\text{C}$ throughout
92 the US treatment. The US processor was operated at three different amplitudes of 20%, 50%
93 and 70%, corresponding to power intensities of 10.6, 34.3 and 48.4 W/cm^2 tip surface,
94 respectively. The treatment period was varied from 2 to 60 minutes, after which the US-treated
95 curdlan solution was taken for measurement of various properties.

96

97 **2.3 Molecular weight determination by intrinsic viscosity measurement**

98 Intrinsic viscosity $[\eta]$ of curdlan solution in NaOH was determined by the dilution method
99 (Yang & Zhang, 2009) and the viscosity was measured using an Ubbelohde viscometer at
100 $25.0 \pm 0.1^\circ\text{C}$ in a water bath. The viscosity-average molecular weight (M_v) of curdlan was
101 determined according to Mark-Houwink equation,

$$102 \quad [\eta] = KM_v^a, \text{ (Eq. 1)}$$

103 where $K = 0.0079$ and $a = 0.78$ for curdlan dissolved in 0.3 M NaOH at 25°C (Nakata et al.,
104 1998), or $K = 0.0032$ and $a = 0.85$ for curdlan dissolved in 0.1 M NaOH at 25°C (Futatsuyama
105 et al., 1999).

106

107 **2.4 Ultrasonic degradation kinetic model**

108 The reduction of curdlan molecular weight (M_v) with time during the US was fitted to the
109 following kinetic model for ultrasonic degradation process of polymer originally derived by
110 Malhotra (1986),

111 $\frac{1}{M_t} - \frac{1}{M_0} = kt$ (Eq. 2)

112 where t is the sonication time (min) and k is the rate constant (mol/g·min) of the ultrasound
113 degradation process, and M_0 , M_t are the viscosity-average molecular weights (mol/g) at time 0
114 and time t , respectively. The value of k at a given US power intensity (W/cm²) was determined
115 by linear regression fit of experimental data according to Eq.2.

116

117 **2.5 Gel formation by acidification**

118 A weak gel is usually formed when curdlan in alkaline solution is neutralized with the
119 addition of acid, due mainly to the single and triple-helical conformation of curdlan polymer
120 chain (Saito et al., 1977; Kanzawa et al., 1987). For evaluation of the US effect on the solubility
121 of curdlan molecules in a neutral solution, 36% HCl was added at an equal mole to NaOH to
122 the US-treated curdlan solution immediately after the US treatment for neutralization. The gel
123 formed after the neutralization was separated from the liquid solution by centrifugation at 6000
124 rpm for 15 min, and then dried at 70°C for 2 days for weight measurement.

125

126 **2.6 Congo red-curdlan complex test**

127 Congo red test was a simple and useful measurement of the high-ordered conformation of
128 polysaccharide chain in alkaline solution (Saito et al., 1977; Ogawa & Hatano 1978). The test
129 was performed of the curdlan solution before and after sonication at various NaOH
130 concentrations. Congo red was mixed with curdlan of 0.2% in 25 mL NaOH solution from 0.05
131 to 0.5 M NaOH before and after various sonication periods at a power intensity of 48.6 W/cm².
132 The solution was maintained at room temperature for 1 h and the absorbance spectrum from

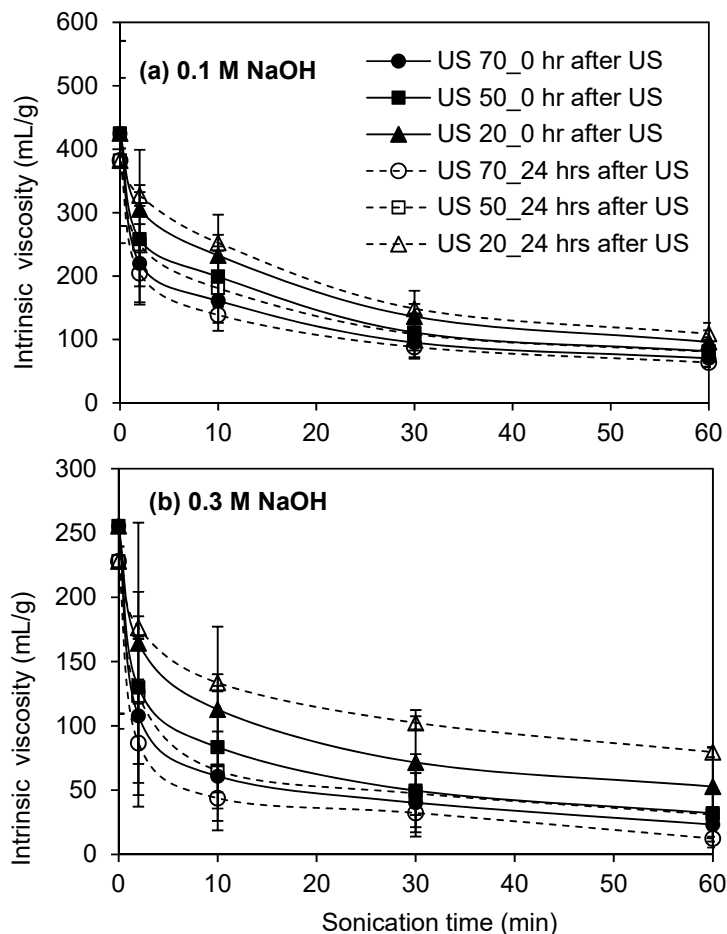
133 400-700 nm was recorded on a UV-Vis spectrometer for detection of the maximum absorbance
134 wavelength (λ_{\max}).

135

136 **3. Results and discussion**

137 **3.1 Effect of ultrasonic power on intrinsic viscosity of curdlan**

138 Fig.1 shows the decrease of intrinsic viscosity of curdlan with sonication time in 0.1 M and
139 0.3 M NaOH. The initial intrinsic viscosity of curdlan on 0.1 M NaOH (4.25 dL/g) was higher
140 than that in 0.3 M NaOH (2.50 dL/g), due probably to the change from helical conformation of
141 curdlan chain in 0.1 M NaOH to random coil in 0.3 M NaOH. With the relatively low alkaline
142 concentration, short degradation period and mild temperature applied in ultrasonic degradation,
143 significant changes in the curdlan structure should be minimum other than conformation
144 changes. According to the literature, extensive degradation of polysaccharides such as cellulose
145 in low alkaline concentrations can only take place at relatively high temperature (over 100°C)
146 or over prolonged periods of days, months or years (VanLoon and Glaus, 1997; Pavasars,
147 Hagberg, Boren, & Allard, 2003).



148

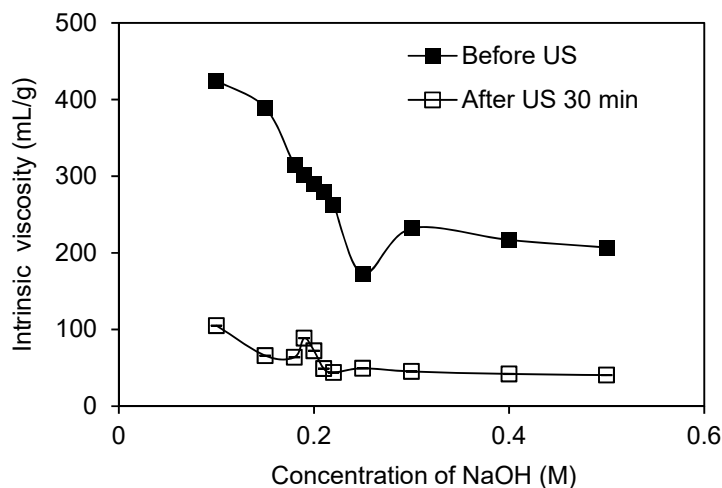
149 **Fig. 1.** Intrinsic viscosity change of curdlan solution with sonication time in (a) 0.1 M and (b)
 150 0.3 M NaOH caused by ultrasound treatment at various power intensity (solid line: measured
 151 immediately after US treatment; dash line: measured 24 hours after US treatment).

152

153 The intrinsic viscosity $[\eta]$ decreased rapidly within the initial 5-10 min and slowly in the
 154 later period of sonication. At 70% amplitude for instance, $[\eta]$ was reduced by about 70% in the
 155 first 10 min and by about 90% in 60 min. The reduction of intrinsic viscosity was more rapid
 156 and significant at a higher US power intensity. The intrinsic viscosity value was not much
 157 different when the measurement was taken immediately after sonication or 24 h later,
 158 suggesting a stable change by the US treatment. However, at a low US power intensity of 10.6

159 W/cm² corresponding to 20% amplitude the intrinsic viscosity measured 24 h later was slightly
160 higher than that measured immediately after US treatment.

161 Fig. 2 shows the variation of intrinsic viscosity with NaOH concentration in the curdland
162 solution. The intrinsic viscosity of native curdland decreased sharply with the increase in NaOH
163 concentration from 0.1 to 0.25 M and increased noticeably as the NaOH concentration
164 increased from 0.25 to 0.3 M. This intrinsic viscosity trend reflects the transition of
165 conformation from helical to random coil form (Nakata et al., 1998). In contrast, the curdland
166 solution after 30 min US treatment had a much lower intrinsic viscosity and only changed
167 slightly with the increase in NaOH from 0.1 to 0.5 M, due mainly to the fact that the US treated
168 curdland was mainly present in random coil form. The conformational changes of curdland with
169 US treatment and NaOH concentration were further examined in section 3.4.



170

171 **Fig. 2.** Intrinsic viscosity change of native and US-treated curdland with NaOH concentration
172 (curdland 2 g/L; US intensity 48.4 W/cm² for 30 min).

173

174 **3.2 Ultrasonic degradation kinetics of curdland**

175 Table 1 shows the viscosity-average molecular weight M_v of curdlan before and after
176 sonication in various conditions estimated from the intrinsic viscosity shown above. Table 2
177 shows the degradation rate constants and correlation coefficients derived by linear regression
178 fitting of the data of $1/M_v$ versus sonication time t to the kinetic model Eq.2. The rate constant
179 k increased with the US power intensity. In addition, k was much larger in 0.3 M NaOH than
180 in 0.1 M NaOH and it also increased with US power intensity more rapidly at 0.3 M NaOH.
181 This suggests that the random coil conformation of curdlan chain in 0.3 M NaOH is more
182 susceptible to degradation than the rigid triple helical conformation in 0.1 M NaOH.

183 **Table 1.** Viscosity-average molecular weight of curdlan in alkaline solution before and after
 184 ultrasonication (M_v estimated from measured $[\eta]$ as described in section 2.3).

US intensity (W/cm ²)	US time (min)	M_v (0.1 M NaOH)	M_v (0.3 M NaOH)
(Original)	0	165,453	86,196
48.4 (US 70)	2	71,278	28,515
	10	47,697	13,692
	30	24,248	8,066
	60	16,500	3,984
34.3 (US 50)	2	87,309	36,220
	10	62,617	20,523
	30	29,590	10,547
	60	19,912	6,022
10.6 (US 20)	2	108,314	48,908
	10	76,345	30,186
	30	38,422	16,836
	60	24,596	11,415

185

186 **Table 2.** Degradation rate constant k (mol/g·min) and correlation coefficient (R^2) of curdlan
 187 under ultrasound treatment at different power intensity (W/cm²).

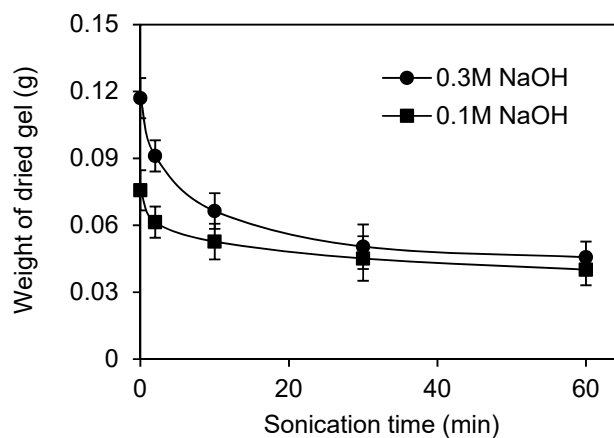
US intensity (W/cm ²)	Rate constant, k (mol/g·min)		Correlation coefficient (R^2)	
	0.3 M NaOH	0.1 M NaOH	0.3 M NaOH	0.1 M NaOH
48.4	0.0040	0.0010	0.932	0.982
34.3	0.0026	0.0008	0.977	0.952
10.6	0.0014	0.0006	0.971	0.914

188

189 3.3 Effect of US on gel formation and solubility of curdlan in neutral condition

190 After sonication, the amount of gel formed by neutralization of the alkaline curdlan solution

191 was decreased (Fig.3). In other words, solubility of curdlan molecules in neutral solution was
192 increased by US treatment. The decrease in the gel formed in 0.3 M NaOH was more significant
193 than that in 0.1 M NaOH. In the alkaline solution of curdlan, gel forms by curdlan chains
194 through interaction with the sodium chloride salts (Saito *et al.*, 1990) so that more gels could
195 be formed at a higher NaOH concentration in which more sodium salts were formed during
196 acidification.



197
198 **Fig. 3.** Variation of the amount of gel formed by acidification for the curdlan solution (2 g/L)
199 after US treatment at 48.4 W/cm².

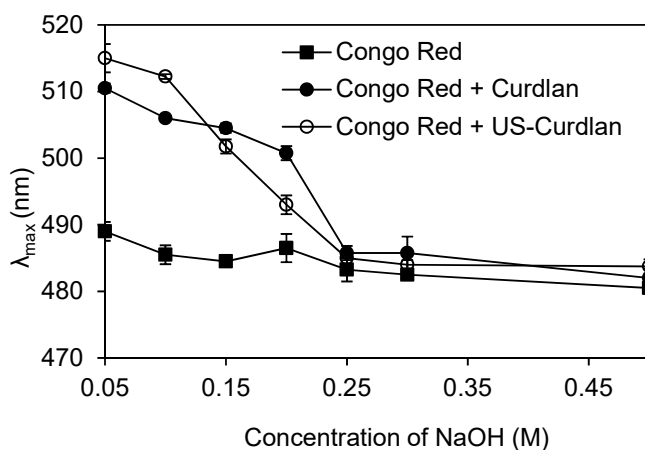
200

201 3.4 Effect of US on curdlan chain conformation

202 Fig.4 shows the results of Congo red test on curdlan before and after sonication. In the
203 NaOH concentration range 0.05-0.2 M, the λ_{\max} of Congo red-curdlan solution was all higher
204 than Congo red solution. The increase in the absorption maximum or the red shift is attributed
205 to the formation of complex between Congo red molecules and the single helical structure of
206 the glucan chain (Ogawa & Hatano, 1978). Therefore, single helices existed in the curdlan
207 solution at NaOH concentrations below 0.2 M. The λ_{\max} value of sonicated curdlan was lower
208 than that of the Congo red mixed with the native curdlan solution at a higher NaOH

209 concentration than 0.15 M, due probably to the disruption of the single helices, which hindered
210 the complex formation between the curdlan and Congo red molecules. In 0.1 M NaOH solution
211 when the curdlan polymer chain was mainly in triple-helix conformation, the corresponding
212 λ_{\max} value of the US-treated curdlan attained a maximum at 30 min of sonication (Fig. 5). The
213 result suggests that the amount of single helices was increased in the curdlan solution after
214 sonication. However, a sharp decrease of the λ_{\max} was detected after much longer period of
215 sonication for 60 min, due probably to the severe destruction and loss of the single helix
216 conformation.

217

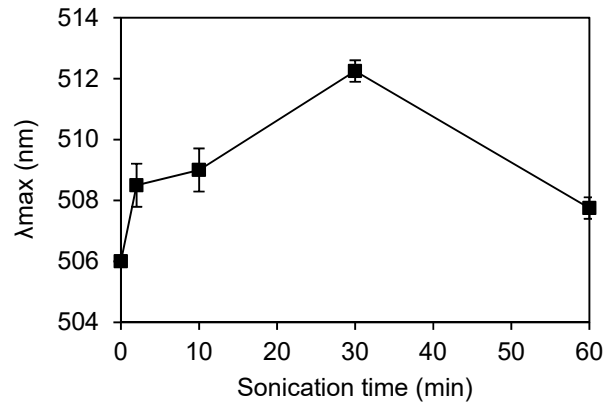


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219

220 **Fig. 4.** Variation of λ_{\max} of Congo red solution alone (Congo red), Congo red with native
221 curdlan sample (Congo red + curdlan), and Congo red with sonicated curdlan at 48.4 W/cm²
222 for 30 min (Congo red + US-Curdlan) at various NaOH concentrations.

223



224

225 **Fig. 5.** Variation of λ_{\max} with sonication time of US-treated curdlan in the Congo red test (US
226 intensity 48.4 W/cm² at 70% amplitude; curdlan in 0.1 M NaOH solution).

227

228 3.5 General discussion

229 Curdlan is a linear β -1,3-glucan and soluble in alkaline solution, and forms gels during
230 acidification of the alkaline solution. The curdlan polymer chain is mainly present in single and
231 triple helix conformation at low concentration of alkaline (0.01-0.19 M NaOH) and the helical
232 structure can be disrupted, and unwounded into random coil structure at high alkaline
233 concentrations (> 0.25 M NaOH) (Nakata et al., 1998; Zhang et al., 2003; Ogawa et al., 1972).
234 High-intensity US can also cause disruption of the curdlan backbone in an alkaline solution,
235 leading to a higher flexibility of the backbone (Gogate & Prajapat, 2015; Yan et al., 2015).
236 Therefore, the triple helix structure is initially degraded into triplex or duplex loops and finally
237 into random coils. The loose loops and random coils of curdlan chains have weaker intra- and
238 inter-molecular interactions such as hydrogen bonds with the interstitial water molecules for
239 the formation of micelles in the solution. Consequently, the amount of micelles decreases and
240 the amount of random coil chains increases with prolonged US treatment. The
241 intra/intermolecular hydrogen bonds between backbones are also weakened by increasing

242 alkaline concentration. Curdlan in random coil conformation is more susceptible to the
243 ultrasonic degradation, as evidenced by the higher degradation rate constant k and more rapid
244 increase with the ultrasonic intensity in 0.3 M NaOH than in 0.1 M NaOH.

245 The notable decrease of viscosity with increasing US power and sonication time is
246 attributable to the conformation change of curdlan chain from relatively rigid triple helix to
247 more flexible single helix and random coil in 0.3 M NaOH solution. The Congo red test
248 provided direct evidence for the conformation changes. With the decrease of molecular weight
249 by ultrasonic degradation, more hydroxyl groups are exposed in solution, resulting in a higher
250 charge-to-mass ratio and stronger electrostatic interaction. The bound water per molecule also
251 increases, leading to higher water solubility and reduced gel formation in neutral condition.

252

253

254 **4. Conclusions**

255 Power US was proven a simple and effective means for partial degradation of curdlan in
256 aqueous alkaline solution. The degradation rate was dependent on the alkaline (NaOH)
257 concentration, being higher at a higher NaOH concentration. Ultrasonic degradation of curdlan
258 caused conformational changes of the polysaccharide chain as well as reduction of intrinsic
259 viscosity and molecular weight. It is of interest to further investigate the sites of chain cleavage
260 occurring in the curdlan chain by ultrasonic degradation and to characterize the specific
261 structure of US-degraded curdlan.

262

263

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271 **References**

272 Cai, Z., Zhang, H. (2017). Recent progress on curdlan provided by functionalization strategies.
273 *Food Hydrocolloid*, 68, 128-135.

274 Cheung, Y. C., & Wu, J. Y. (2013). Kinetic models and process parameters for ultrasound-
275 assisted extraction of water-soluble components and polysaccharides from a medicinal
276 fungus, *Biochemical Engineering Journal*, 79, 214-220.

277 Čížová, A., Bystrický, P., & Bystrický, S. (2015). Ultrasonic and free-radical degradation of
278 mannan from *Candida albicans*. *International Journal of Biological Macromolecules*, 75,
279 32-36.

280 Futatsuyama, H., Yui, T., & Ogawa, K. (1999). Viscometry of curdlan, a linear (1→3)-β-D-
281 glucan, in DMSO or alkaline solutions. *Bioscience, Biotechnology, and Biochemistry*,
282 63(8), 1481-1483.

283 Gogate, P. R., & Prajapat, A. L. (2015). Depolymerization using sonochemical reactors: A
284 critical review. *Ultrasonics Sonochemistry*, 27, 480-494.

285 Huang C., Miao, M. B., Jiang, S. W., Cui, Jia X., & Zhang T. (2015). Polysaccharides
286 modification through green technology: Role of ultrasonication towards improving
287 physicochemical properties of (1-3)(1-6)-α-D-glucans. *Food Hydrocolloid*, 50 166-173.

288 Kanzawa, Y., Harada, T., Koreeda, A., & Harada, A. (1987). Curdlan gel formed by neutralizing
289 its alkaline solution. *Agricultural and Biological Chemistry*, 51(7), 1839-1843.

290 Liu, Y., Gu, Q., Ofosu, F. K., & Yu, X. (2015). Isolation and characterization of curdlan
291 produced by *Agrobacterium* HX1126 using α-lactose as substrate. *International Journal*
292 *of Biological Macromolecules*, 81, 498-503.

293 Malhotra, S. (1986). Ultrasonic solution degradations of poly (alkyl methacrylates), *Journal of*
294 *Macromolecular Science, Part A*: 23(6), 729-748.

295 Mason, T. J., & Lorimer, J. P. (2002). *Applied sonochemistry: the uses of power ultrasound in*
296 *chemistry and processing*, Wiley-VCH, UK.

297 McIntosh, M., Stone, B., & Stanisich, V. (2005). Curdlan and other bacterial (1→ 3)-β-D-

298 glucans. *Applied Microbiology and Biotechnology*, 68(2), 163-173.

299 Nakata, M., Kawaguchi, T., Kodama, Y., & Konno, A. (1998). Characterization of curdlan in
300 aqueous sodium hydroxide. *Polymer*, 39(6-7), 1475-1481.

301 Ogawa, K., & Hatano, M. (1978). Circular dichroism of the complex of a (1→3)-β-D-glucan
302 with Congo Red. *Carbohydrate Research*, 67(2), 527-535.

303 Ogawa, K., Wanatabe, T., Tsurugi, J., & Ono, S. (1972). Conformational behavior of a gel-
304 forming (1→3)-β-D-glucan in alkaline solution, *Carbohydrate Research*, 23(3), 399-405.

305 Pavasars, I., Hagberg, J., Boren, H., & Allard, B. (2003) Alkaline degradation of cellulose:
306 mechanisms and kinetics. *Journal of Polymers and the Environment*, 11(2), 39–47.

307 Prajapat, A. L., Subhedar, P. B., Gogate, P. R. (2016). Ultrasound assisted enzymatic
308 depolymerization of aqueous guar gum solution, *Ultrasonics Sonochemistry*, 29, 84-92.

309 Saito, H., Ohki, T., & Sasaki, T. (1977). A carbon-13 nuclear magnetic resonance study of gel-
310 forming (1→3)-β-D-glucans. Evidence of the presence of single-helical conformation in
311 a resilient gel of a curdlan-type polysaccharide 13140 from *Alcaligenes faecalis* var
312 myxogenes IFO 13140. *Biochemistry*, 16(5), 908-914.

313 [Saito, H., Yoshioka, Y., Yakoi, M., Yamada, J. \(1990\). Distinct gelation mechanism between
314 linear and branched \(1/3\)-β-D-glucans as revealed by high-resolution solid-state carbon-
315 13 NMR. *Biopolymers*, 29, 1689-1698.](#)

316 Tao, Y., & Sun, D. W. (2015). Enhancement of food processes by ultrasound: a review. *Critical*
317 *Reviews in Food Science and Nutrition*, 55(4), 570-594.

318 VanLoon, L.R., & Glaus, M.A. (1997). Review of the kinetics of alkaline degradation of
319 cellulose in view of its relevance for safety assessment of radioactive waste repositories.
320 *Journal of Environmental Polymer Degradation*, 5(2), 97-109.

321 Yan, J. K., Pei, J. J., Ma, H.L., Wang, Z.B. (2015). Effects of ultrasound on molecular properties,
322 structure, chain conformation and degradation kinetics of carboxylic curdlan,
323 *Carbohydrate Polymers*, 121, 64-70.

324 Yang, L. Q., & Zhang, L. M. (2009). Chemical structural and chain conformational
325 characterization of some bioactive polysaccharides isolated from natural sources,
326 *Carbohydrate Polymers*, 76(3), 349-361.

327 Zhang, L., Wang, C., Cui, S., Wang, Z., Zhang, X. (2003). Single-molecule force
328 spectroscopy on curdlan: unwinding helical structures and random coils, *Nano Letters*,
329 3(8), 1119-1124.

330 Zhang, R., & Edgar, K. J. (2014). Properties, chemistry, and applications of the bioactive
331 polysaccharide curdlan, *Biomacromolecules*, 15(4), 1079-1096.

332 Zhi, Z., Chen, J., Li, S., Wang, W., Huang, R., Liu, D., Ding, T., Linhardt, R.J., Chen, S., & Ye,
333 X. (2017). Fast preparation of RG-I enriched ultra-low molecular weight pectin by an
334 ultrasound accelerated Fenton process. *Scientific Reports*, 7(1), 541.